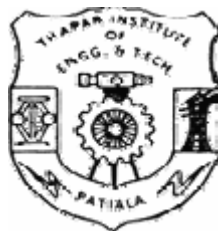


**STUDIES ON THE STATUS OF STARCH
PHOSPHORYLATION IN DEVELOPING TUBERS
FROM
VARIOUS POTATO CULTIVARS**

**A
DISSERTATION
By**

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**Submitted in partial fulfillment of the requirement for the
award**

**of the degree of Masters of Science in Biotechnology
Department of Biotechnology and Environmental Sciences**

Thapar Institute of Engineering and Technology

Patiala –147004

2004

CANDIDATE'S DECLARATION

I, hereby declare that the work presented in the dissertation entitled, "**Studies on the Status of Starch Phosphorylation in Developing Tubers From Various Potato Cultivars** " in partial fulfillment of the requirement for the award of the degree of Masters in Biotechnology, Department of Biotechnology and Environmental Sciences, Thapar Institute of Engineering and Technology, Patiala; is an authentic record of my own work during the period of five months from January 2004 to May 2004, under the supervision of Dr. N. Das ,Assistant Professor, Thapar Institute of Engineering and Technology. I have not submitted the matter embodied in this dissertation for the award of any other degree or diploma.

Place: Patiala TEENA SHUKLA

Date

This is to certify that the above statement made by the candidate is correct and true to the best of our knowledge.

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CERTIFICATE

This is to certify that the thesis entitled "**Studies on the Status of Starch Phosphorylation in Developing Tubers From Various Potato Cultivars** " submitted by Teena Shukla in partial fulfillment of the requirements for the award of Degree of Masters of Science in Biotechnology, to Thapar Institute Of Engineering and Technology (Deemed University), Patiala, is a record of student's own work carried out by her under my supervision and guidance. The report has not been submitted for the award of any other degree or certificate in this or any other university or institute.

(Dr. N. Das) (Dr. Sunil Khanna)

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ACKNOWLEDGEMENT

I thank the almighty whose blessings have enabled me to accomplish my dissertation work successfully.

It is my pride privilege to express my sincere thanks and deep sense of gratitude to **Dr. N. Das**, Assistant Professor, Department of Biotechnology and Environmental Sciences, for his valuable advice, splendid supervision and constant patience through which this work was able to take the shape in which it has been presented. It was his valuable discussions and endless endeavors through which I have gained a lot. His constant encouragement and confidence-imbibing attitude has always been a moral support for me.

My sincere thanks to **Dr. Sunil Khanna**, Head Department of Biotechnology and Environmental Sciences, for his immense concern throughout the project work. A special word of thanks to all the faculty members for their constant encouragement and support throughout this duration.

I take this opportunity as a privilege to thank, **Dr. K.S Barna** Research Scientist, TIFACCORE

and **Mr. Dipal R. Choudhury**, Research Scientist, TIFAC-CORE, and tissue culture department at TIFAC-CORE for his timely help in preparing this rapport in present colorful and brightful way.

I am really grateful for the help rendered the with special mention of Mr. Asnshu, Ms. Sunita Bansal and Mrs. Vijay who was always there for help in completing my project work in a smooth way.

I feel lacunae of words to express my most heartfelt and cordial thanks to my friends Seema, Guneet, Arun, Karan, Rakesh and Amit who have always been a source of inspiration for me, stood by my side at the toughest times.

Finally, I wish to extend a warm thanks to everybody involved directly or indirectly with my work.

The whole credit of my achievements goes to my parents, sister and brother, who were always there for me in my difficulties. It was their unshakable faith in me that has always helped me to proceed further.

DATE: (TEENA SHUKLA)

ABSTRACT

Potato is one of the most important food crops of the world. It is cultivated for its tubers, which are modified stem structures formed by the enlargement of the tips of underground stems. A crucial feature of the post-harvest physiology of potato tubers is an undesirable process called cold-induced sweetening. A comprehensive biochemical and molecular approach need to be undertaken to overcome such problem in the cultivars suitable to our agro-climatic conditions. Currently, phosphorylation status of starch in potato tubers is believed to play a crucial role in the above process as phosphate groups attached to glucose residues at C-6 and C-3 positions alter overall secondary structure of starch hence its degradability at low temperature. We need to have an insight to find correlation between starch phosphorylation and cold sweetening process. Keeping in view with the above aim a few basic experiments were done and executed in this dissertation work. The study was done on several potato cultivars namely Kufri Chipsona-1 (CS-1), Kufri Chipsona-2 (CS-2), Kufri Chandramukhi (KCM), Kufri Jyoti (KJ), Kufri Pukhraj (PR), Kufri Ashoka (AS) and Cultivar Desiree (DE), which were routinely micropropagated in our laboratory. After proper hardening and acclimatization, the potato plantlets of the above mentioned varieties were cultivated in the field in the second week of November 2003. Developing mini-tubers were collected time to time through out the entire period of cultivation. Mature tubers from various cultivars were also transferred to 20°C for a period of four weeks as required in this study. A rapid and convenient method was adopted during isolation of starch granules from various tuber samples and overall yield was calculated in gm per 100 gm of tuber weight. Starch-bound phosphates were converted to inorganic phosphates by complete ashing procedure.

A very sensitive ascorbic acid-ammonium molybdate assay method was adopted for estimation of total phosphorus. The level of free phosphates was also checked in various starch preparations. The purpose was to see the status of starch phosphorylation in the tubers at various stages of development. This study was extended to the several potato cultivars that were routinely micropropagated in our laboratory. With respect to the above biochemical attribute profile was made for each cultivar variety and finally a comparison also made among the cultivars. This type of study could help further to find correlation between starch phosphorylation and cold sweetening phenomenon.

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1. INTRODUCTION

The potato is the most popular and widely used vegetable in the world. It belongs to family Solanaceae. The potato is the only member of the Solonaceae of outstanding agricultural importance. This family includes herbs, shrubs, trees, or vines. The plants in this family are dicotyledons and are of considerable importance for food, drugs, weeds, and poisonous plants. The genus *Solanum* comprises some 2000 species and is very variable in habit, about 170 species produce underground stem tubers. The potato forms an indispensable item of daily food and is an important source of nutrition parity. Today potato is the fourth most important food crop in the world. Potato is one of the highest calorie-yielding crops in the world. Such root crops are particularly valuable in the tropics where most of the population depends on carbohydrate foods as dietary staples.

1.1 ORIGIN

The first edible potatoes are considered to have been originated in the Peruvian and Bolivians Andes of South America, where it was consumed for many millennia. The South American Indians were in fact able to select alkaloid-free potato varieties. Later, Spanish explorers carried these potatoes to Europe in the late 16th century and from there, they spread potatoes throughout the world. By the 19th century it had spread throughout the Indian sub-continent. Initially, the crop was used as a medicinal plant and grown by pharmacists, in Spain in particular. Merchants and kings, who encouraged the cultivation of this efficient plant to increase local agricultural production, later introduced it to other parts of Europe. The successful introduction of this new crop did not only require changes in the dietary habits of the people, but also a biological adaptation of the crop to a new climate. In fact, the

potato plant being originally adapted to short day conditions of the tropical highlands, it

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would yield very little under the long summer days in Europe. Breeding over more than

150 years led to plants tolerating long day conditions. The modern breeding of potatoes

began approximately in 1780, where crossings were performed between local varieties. At

the beginning of the 19th century, the introduction of new potato germplasm, especially

from Chile, contributed highly to the breeding of modern varieties. Towards the end of the

last century, there was already a large array of breeding varieties available to the breeders.

However, because of the need for new resistance genes against pests and diseases, the 20th

century brought about the use of a large population of wild- and cultivated potato species

from South America for backcrossing into European varieties. The potatoes of today in

Europe are largely the result of the intensive breeding programs of the 19th century, but

have benefited greatly from the improvements in breeding techniques of the 20th century to

improve traits like disease resistance, tolerance to environmental factors.

1.2 THE BIOLOGY OF POTATO PLANT

The Scientific name of potato is ***Solanum tuberosum* L.** It belongs to family Solanaceae,

genus *Solanum* and species *tuberosum*. The first cultivated potato species were diploid. The

development of the modern varieties was related to the spontaneous occurrence of

tetraploid species that were superior in yield.

The common potatoes (*Solanum tuberosum* L.) consist of two subspecies. The most

common cultivated species is a tetraploid, with chromosome number of 48 (4n=48), which

is thought to have arisen as a hybrid between the diploid species ***S. stentotomum*** and the

diploid weed ***S. sparsipilum*** with subsequent chromosomal doubling.

1.3 CHARACTERISTICS OF POTATO PLANT

Solanum tuberosum L. is an herbaceous cultivated as an annual crop, and is susceptible to

frost and freezing. It is a cool season crop. The commercially significant portion of the

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plant is the tuber, which is a fleshy, swollen, rounded, oval or oblong distal portion of

underground axillary or adventitious branches. Underground stem is called as stolon.

Swelling causes when stolon ceases to elongate. Swollen region of an underground stem is

modified due to the translocation and storage of photosynthates (carbohydrates), which

occurs as the aerial portion of the plant reaches maturity. Tubers are used in commercial

propagation since the true seeds are heterozygous and highly variable but used primarily in

crop improvement.

Root System - Potatoes produce a fibrous root system arising from initials along the

underground portion of the stem. These roots can extend to a depth of around 20 inches,

but are not highly effective in penetrating the soil layers in search of water. True seed

potatoes produce a tap root system

Eyes - The eyebrow is a leaf scar, which always faces the apical end of the potato.

Eyebrows are highly concentrated at the apical end. The eyes on a potato are the growing

points for new plants. Each tissue layer is connected to each eye. Eyes appear in higher

concentration nearest the "rose end" of the tuber, decreasing in size and concentration

towards the "heel end" where a stolon scar is visible. During the growth of the tuber, the

eyes remain dormant.

Stem (basal) end - the stem (basal) end is the attachment point between the tuber and the

plant. The stem cannot produce new plants. As the plant matures, the stem weakens and lies

prostrate, eventually yellowing and dying back at the end of the growing season.

Leaf – leaf pattern is pinnately compound, alternate, with 7-9 ovate leaflets (one terminal

leaf). The margins are serrated or entire.

Flower - the potato has a perfect flower, which contains both male and female flower parts.

It can be white, purple, lilac, or violet depending on the variety of potato. A significant

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amount of potato cultivars are either pollen sterile or fail to set fruit because of some other means.

True seed - If fertilization is successful, a small green fruit ball (to 1 ¼" diameter) is

produced. The fruit is a berry with seeds in a mucilagenous pulp. Seeds are flattened and

ovate, around 300 seeds per fruit. Cross-pollination is manipulated for crop improvement in

quality and insect or disease resistance. However, many potato cultivars are pollen sterile,

which creates difficulty in breeding programs.

Temperature - Potatoes are affected by differences in temperature. Tuberization occurs

earlier at lower temperature. The optimal temperature for tuberization is 55°F, the process

decreases above 70°F and with certain cultivars, may stop at 85°F. Higher temperatures

may often induce knobiness and secondary growth in tubers. Maximum yields may be

obtained with an average growing temperature between 60° and 65°F. These cooler

temperatures cause the rate of respiration to be lower than the rate of photosynthesis,

resulting in more accumulation of carbohydrates.

Photoperiod - A majority of potato cultivars are not day length sensitive. Cool night

temperatures are important because they affect the accumulation of carbohydrates and dry

matter in the tubers. A misconception is that potatoes contain alkaloid solanine, while the

fact is that only green potatoes contain it. Potato plants and tubers contain the toxic

glycoalkaloids, alpha-solanine, and alpha-chaconine, which act as cholinesterase inhibitors.

When tubers are exposed to light, chlorophyll along with the glycoalkaloids is synthesized.

The amount of glycoalkaloids formed depends on exposure length, intensity and light

quality (mostly ultraviolet), and temperature; little is synthesized at temperatures below 41°F. These compounds taste bitter, and ingestion can cause illness even death in extreme cases, toxicity depends on the amount ingested.

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1.4 NUTRIENT PROFILE OF POTATO

The main nutritional value of and tubers lies in their potential ability to provide one of the cheapest sources of dietary energy, in the form of carbohydrates Potatoes are the most important 'vegetable' in the world today, after cereals the tubers provide the main source of carbohydrate. Potato contains high nutritive value. Potatoes yield 17 - 21% fresh weight of starch and 0.5 - 1.2 % of pure protein.

Food Value Minerals and Vitamins

Moisture - 74.7 % Calcium - 10 mg

Carbohydrates - 22.6 % Phosphorus - 40 mg

Protein - 1.6 % Iron - 0.7 mg

Minerals - 0.6 % Vitamin C - 17 mg

Fat - 0.1 %

Fibre - 0.4 %

*Values per 100 gm edible portion

Small amount of Vitamin B complex, Vitamin A and P is also present.

Misconceptions

regarding the nutritional value of potato need to be dispelled to increase domestic consumption. Most people think that potatoes cause obesity, while it has only 0.1 per cent

fat. It absorbs fat during frying. The potato can be boiled, baked, steamed and cooked with

other vegetables. But it should be cooked in such a way so as to retain all its excellent

qualities. To secure its maximum value, it should always be cooked with its skin as the

most nutritive part of the potato lies just below the skin and this particular layer is very rich

in protein and mineral salts'

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1.5 Natural Benefits and Curative Properties

The potato contains several medicinal virtues. As it is one of the most strongly alkaline of

all foods, it is, therefore, very helpful in maintaining the alkali reserve of the body and a natural antidote for an overdose of acid or acidosis.

However, obese people should avoid fried potatoes as they are fattening. They should also

be omitted from the diet of those suffering from venereal diseases and those afflicted with

aphrodisiac tendencies. The potatoes green in color contains an alkaloid toxin known as

solanine, which affects the nerves controlling the sexual organs.

An exclusive potato diet is considered valuable in the treatment of certain disorders such-as

chronic constipation, intestinal toxæmia, uric acid diseases, renal calculi or stone and

dropsy. The potato is regarded as an excellent food remedy in scurvy. Raw potato juice

helps to eliminate an "an acid condition" and relative rheumatism, skin blemishes and

swellings. Potato juice also relieves gastritis. Stomach ulcers are treated with juice of pink

potatoes.

Potato starch is administered as an anti-inflammatory for gastro-intestinal diseases and

toxins. The skin is exceptionally rich in vital mineral salts and the water in which the

peelings have been boiled is one of the best medicines for the ailments caused by excess of

acid in the system. Raw potato juice externally applied is valuable in the treatment of

swelling and other disordered conditions of the joints and muscles.

1.6 MICROPROPAGATION AND CULTIVATION

Tissue Culture is currently being used to perpetuate disease free seed stock, which can then

be stored "*in vitro*" until needed. This method of micropropagation is meant for rapid

distribution maintenance of existing cultivars and to bulk up new breeding lines for

germplasm storage and transport. Furthermore, the molecular analysis of plantlets, which

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propagate and stored *in vitro*, shows that micropropagation gives genetically stable

plantlets. Another approach for micropropagation of potatoes is the *in vitro* induction of

micro-tubers. Although considerable research has been performed on micro-tubers, their use on commercial basis in the potato industry has been rarely formulated. They can be stored several months and easily transported unlike *in vitro* plantlets. Thus, they are ideal for germplasm distribution as they can conform to long quarantine procedures. They can be directly sown into the soil. Unlike propagation of *in vitro* plantlets, they do not need subculturing to fresh media. They can be produced in bulk in any season. Nevertheless, the cost of producing one micro-tuber is relatively higher than that of an *in vitro* plantlet and has thus restrained its uses only for gene resource preservation. Consequently, several experiments have been aimed on the factors controlling the tuberization process in view of implementing those findings in the development of rapid and cost effective methods of producing micro-tubers on a large-scale basis. These factors are environmental, hormonal, nutritional and physiological in nature. The plants growing *in vitro* have tender, young root lacking root hairs, which need to be hardened. Hardening and acclimatization comprise of transition from completely controlled conditions to uncontrolled field conditions.

1.7 POTATO PROCESSING

The potato (*Solanum tuberosum*) is a major world food crop, surpassed by only wheat, rice, and corn in world production for human consumption. Potato is used in livestock feed and many processed foods, such as potato chips, curls, flakes, fries, thickening agents, and alcoholic beverages. Potato tubers also have a number of industrial uses like potato starch provides a tough resilient coating for paper and textiles.

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In general, tubers with a high dry matter, high amylose to amylopectin ratio, small cell size and low sugar content is preferred for most processing uses, including baking or frying.

Potatoes with a low dry matter are best used boiled because they tend to remain intact. The

starch composition tends to have a low amylose to amylopectin ratio. Such potatoes, when baked, tend to have a moist texture. However, not all potato varieties available in country are suitable for processing.

Potatoes are typically classified into three main types: round whites, red-skinned and russets, red-skinned cultivars generally have low dry matter content (low specific gravity) and are used for fresh market consumption. Round white cultivars with high specific gravity and fewer reducing sugars are used for chip processing, whereas round white types with lower specific gravity are used for table stock. Long russet types (high specific gravity) are used for frozen processing and dehydration. The long russet is the major type used for table stock. In addition, all fresh market types should be uniform and have an attractive appearance: good size, smooth shape and no external defects.

1.8 BIOSYNTHESIS AND OTHER ATTRIBUTES OF STARCH

Keeping in view with the objective of present dissertation work, several attributes of potato starch including its biosynthesis are discussed briefly in the following few sections. The discussion is mostly focused on potato starch.

Starch is very important substance, both to plants and humans. For humans, it is a vital component of our diet, and an important commercial product. Starch and its derivatives are already widely employed in the manufacture of paper, textiles and adhesives, and due to their biodegradable and renewable nature they are increasingly being considered as an environmentally-friendly alternative to the use of synthetic additives in many other

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products, including plastics, detergents, pharmaceutical tablets, pesticides, cosmetics and even oil-drilling fluids.

Starch is the major storage carbohydrate (polysaccharides) in higher plants, being the end product of photosynthesis. As during the growing season, the green leaves collect energy

from the sun. In potatoes this energy is transported as a sugar solution down to the tubers, and down there, the sugar is converted to starch in the form of tiny granules occupying most of the cell interior.

It is believed that starch biosynthesis in potato tubers is developmentally regulated process in the amyloplast of the tubers. The synthesis of starch in plant cells begins with the enzyme ADP-glucose pyrophosphorylase (AGPase), which catalyses the reaction of glucose-1-phosphate with ATP to form ADP-glucose (liberating pyrophosphate). The ADP-glucose is then used a substrate by starch synthase enzymes, which add glucose units to the end of a growing polymer chain to build up a starch molecule (releasing the ADP in the process). Branches in the chain are introduced by starch branching enzymes (SBEs), which hydrolyse 1,4-glycosidic bonds, and in their place, create 1,6 bonds with other glucose units. Starch is composed of two polymers, an essentially linear polysaccharide amylose, and a highly branched polysaccharide - amylopectin.

Amylose - The constituent of starch in which anhydroglucose units are linked by α -(1 \rightarrow 4)-D glucosidic bonds to form linear chains. Amylose molecules are typically made from 200-2000 anhydroglucose units. Aqueous solutions of amylose are very unstable due to intermolecular attraction and association of neighboring amylose molecules. This leads to viscosity increase, retrogradation and, under specific conditions, precipitation of amylose particles.

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Amylopectin - A constituent of starch having a polymeric, branched structure. In addition to α -(1 \rightarrow 4)-D bonds that are present in amylose and the linear segments of amylopectin, the amylopectin molecule has α -(1 \rightarrow 6) bonds, which occur every 20-30 anhydroglucose units. Aqueous solutions of amylopectin are characterized by high viscosity, clarity,

stability, and resistance to gelling.

For many commercial uses it is desirable to alter the proportions of amylose and

amylopectin found in starch. High-amylose starches are useful in confectionery (because

they thicken rapidly), in fried snacks (because they resist the penetration of cooking oil),

and in photographic film (because of their toughness and transparency). It has also been

suggested that the nutritional properties of bread can be improved by the use of flour high

in amylose. Amylopectin is preferred in paper-making and adhesives (because its branched

chains give it greater binding power), and in frozen food (because it enhances stability and

shelf-life.

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1.8.1 Composition of Potato Starch

Potatoes are well known for being starchy, and a potato tuber comes close to being 100%

starch storage parenchyma. Potato starch is second most important and abundant starch of

the world. It is the major carbohydrate reserve in plant tuber in the form of granules. Starch

is used as a raw material in the food, pharmacy, cosmetics, paper and plastics industries.

The composition of potato starch is given below:

Starch, dry substance 80 %

Water 20 %

Ash 0.3 %

Sand 0.02 %

Protein 0.09 %

Phosphor, P 0.07 %

Calcium, Ca 0.03 %

Iron, Fe 3 ppm

Cold water solubility 0.1 %

1.8.2 Phosphorylation of starch

Starches contain phosphorus in some form or another. The nature of bound phosphorus

affects starch performance. The phosphate content in potato tubers is exceptionally high.

Small starch granules contain 25 % more ester-bound phosphate per glucose residue than

large starch granules. The phosphate groups are located as monoesters at mainly C-3 & at the C-6 position of the glucose monomers. It is believed that the phosphate content

influences the overall secondary structure of starch granules.

Starch biosynthesis is accomplished by different form of starch synthase, which polymerize

the glucose monomer using ADP- glucose, and isoforms of branching enzymes, which

introduce the branch points. Other enzymes are needed, to determine the final starch

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structure, e.g. the presence of a debranching enzyme is a prerequisite to synthesize the

semi-crystalline starch granules. More phosphorylation in the starch is related to its more

degradability, particularly at low temperature leading to more cold sweetening. However

the mechanism of starch phosphorylation is not completely known.

1.9 NEED AND PROBLEMS

Potato plants are grown in moderate climate, therefore, continuous production of tubers is

not possible round the year. In summers as the temperature rises sprouting, microbial

spoilage and senescence-induced sweetening, weight loss, rottage etc. cause the damage to

the tubers. As a result, major part of potato tubers need to be stored at lower temperatures

for a certain period of time to prevent damage.

A crucial feature of the post-harvest physiology of tubers of the potato plant is a process called cold-induced sweetening, i.e. the accumulation of the reducing sugars

fructose, glucose and sucrose during tuber storage at low temperatures. It results from

partial starch breakdown. Potatoes with high reducing sugar levels pose problem for the

food industry. The reducing sugars react with proteins and amino acids to form dark

products in a non- enzymatic browning reaction after frying. Hence high levels of these

sugars cause discoloration, changes in flavor and loss of essential amino acids in processed

products made from stored tuber slices. Therefore, due to the growing demand of the

processed potato products, important parameters for the selection of raw material in potato

processing industries should have dry matter of the tuber up to 21 to 23 % and reducing

sugar content below 150 mg / 100 g fresh tuber weights.

At present we need to focus on agricultural research to ensure adequate productivity of

potato crop, both for fresh consumption and processed potato products. At the same time

we need to find out proper means to overcome cold storage problems of the harvested

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tubers. At the moment, we need to understand the phenomena of cold-sweetening process

at biochemical levels. We should focus on the potato cultivars suitable to Indian agroclimatic

conditions.

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2. REVIEW OF LITERATURE

As potato is not grown round the year, a major portion is required to be stored at low

temperature (2- 4°C). This storage prevents sprouting, microbial spoilage and senescence

induced sweetening, weight loss and rotting etc.

Potatoes are the most important 'vegetable' in the world today - after cereals the tubers

provide the main source of carbohydrate. Secondary uses include the production of starch

and dextrose, industrial alcohol by fermentation, and spirits. Potatoes yield 17 - 21 % fresh

weight of starch and 0.5 - 1.2 % of pure protein. The importance of this crop was realized

soon after independence. The Central Potato Research Institute, Shimla since its

establishment in 1949 has been doing comprehensive potato breeding programme to

develop improved cultivar varieties. They bred and released many high yielding varieties

so far, which are suitable for different agro-climate regions. Most of these varieties have

been bred for consumption as fresh potatoes. However fresh varieties required for

processing should have the following criteria. Tuber dry matter in the range of 21 to 23 %.

Reducing sugar content below 150 mg per 100 gm fresh tuber weight and reasonably good

yield to provide economic returns to the farmer (Singh *et al.*, 1999).

In general, the potato tubers from different cultivars vary with respect to the following

characteristics: dry matter content, starch content, protein content, starch yield and mean

particle diameter (μm) of the starch granules (Jansen *et al.*, 2001) etc.

2.1 STARCH IN POTATO TUBERS

Starch is the major predominant storage form in potatoes. Potatoes starch is, in many ways,

a superior starch and its many interesting properties make it attractive for both food and

industrial application (Alexander, 1995). Starch synthesis in storage organs occurs over a

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long developmental period, during which there are usually considerable changes in the

complement of starch-synthesizing enzymes (Smith and Martin, 1993; Burton *et al.*, 1995).

Starch is composed of two polymers amylose and amylopectin. The amylose molecules are

essentially linear α -(1 \rightarrow 4)D glucan chains, whereas the amylopectin molecules are highly

branched and often contain covalently-bound phosphate (Hizkuri *et al.*, 1970).

2.2 PHOSPHATE CONTENT IN STARCH

In this section phosphorylation of potato starch is mainly discussed. The phosphorus in

tuber starches is in the form of phosphate monoesters. The phosphate groups are located as

monoesters at the C-6 and at the C-3 position of the glucose residues in the amylopectin are

phosphorylated (Hizkuri *et al.*, 1970).

The low degree of phosphorylation has a profound effect on the qualitative properties of

the starch changing e.g. the gelatinization temperature, viscosity, retrogradation and solubility

(<http://www.plbio.kvl.dk/plbio/starch.htm>). Starch phosphorylation proceeds concurrently

with the *de novo* biosynthesis of starch in potato tubers (Nielson *et al.*, 1994).

Small starch granules contain approximately 25 % more bound phosphate per glucose

residues than large granules, whereas the overall level of phosphorylation does not depend

upon the tuber size (Nielsen *et al.*, 1994). Majority of the phosphate is bound to the amylopectin fraction of starch (Hizkuri *et al.*, 1970). The starch-bound phosphate constitutes a major part of the total phosphate pool in the potato tubers (Quick *et al.*, 1976). The level of phosphorylation may vary approximately two folds among different potato cultivars and depends upon the growth conditions (Nikuni *et al.*, 1969). On an average, one of every 200-500 glucose residues is phosphorylated (Nielsen *et al.*, 1994).

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2.3 COLD INDUCED SWEETENING

Potatoes are stored in cold-stores (2-4°C). This storage prevents the moisture loss, decay, and early sprouting while removing respiratory heat. Storage extends the availability and thereby assists with orderly marketing, distribution, and utilization. Whereas storage can extend the usefulness of harvested potato crops, quality does diminish proportionally to the length of storage. As during storage at low temperatures, a process known as **cold sweetening** occurs in the tuber, i.e. starch is broken down to soluble sugars such as sucrose, glucose and fructose. Hence, it is very desirable to reduce cold-sweetening as potatoes with high sugar levels pose problem for the food industry. It is thought that the cold-induced hexose accumulation is caused by an imbalance between starch breakdown and glycolytic activity. Several explanations are also there related to cold-induced sweetening in the potato tubers.

^ An increased concentration of hexose phosphates as a result of cold lability of phosphofructokinase and other glycolytic enzymes.

^ The starch is thought to be degraded mainly by the phosphorylytic route, based on the relative activities of phosphorylase and amylolytic enzymes. In other words, an increase in the activity of one or more starch degrading enzymes leads to above

process.

^ Increased activity of invertases or the enzymes involved in sucrose synthesis.

(Isherwood, 1973; Pollock and Rees, 1975; Guy *et al.*, 1992; Deiting *et al.*, 1998)

The only reported example of turnover (the simultaneous occurrence of synthesis and

degradation) in storage organs is in transgenic potatoes in which the flux of carbon into

starch was increased 6-fold by elevating ADP-Glc pyrophosphorylase activity 17

(Sweetlove *et al.*, 1996). However, little, if any, turnover was observed in the wild-type

tubers. This is consistent with earlier findings (Dixon and ap Rees, 1980).

2.4 Possible correlation between starch phosphorylation and

cold-sweetening

Various attempts have been made currently in order to identify enzymes/proteins

involved in starch synthesis and its modification. For this, starch bound proteins were

isolated from potato tubers and antisera raised against them for screening of potato cDNA

expression libraries. Recently, a cDNA clone has been isolated and characterized. It is 4851

bp in size encoding starch-granule-bound protein R1, found to cause phosphorylation.

Starch-granule-bound R1 protein involved in starch biosynthesis and modification is an

alpha-glucan, water dikinase (GWD). The enzyme catalyses the phosphorylation of starch

by a dikinase-type reaction in which the beta-phosphate of ATP is transferred to either the

C-6 or the C-3 position of the glycosyl residue of amylopectin. Starch phosphorylation

seems to be an integrated activity in starch biosynthesis. Moreover, potato tubers with

repressed GWD showed significant changes in starch structure.

The R1 protein is involved in determining the phosphate content of potato starch. A major

observation made is the reduction of the phosphate monoester content of the starch

synthesized in the transgenic lines down to 10 % as compared with wild-type plants. This

indicates that the R1 protein is responsible for the phosphorylation of starch, which is also supported by the fact that the expression of the protein in *Escherichia coli* leads to elevated phosphate content of the synthesized glycogen. Similarities to PEP synthase in the C-terminal part of the R1 sequence could be taken as an indication that R1 mediates an ATP-dependent starch phosphorylation by a Dikinase type of reaction. The biochemical mechanism by which the R1 protein phosphorylates amylopectin remains to be elucidated. It is not known which substrate acts as phosphoryl donor, nor which type

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of glucans are phosphate acceptors. Evidence that phosphoenol pyruvate may act as a phosphoryl donor comes from slight sequence homology of the R1 protein to the PEP synthases from different bacteria.

The overall secondary structure of starch from transgenic tubers with lowered levels of R1 protein is altered, that led to less starch degradation during cold storage. It was found that

the level of reducing sugars was up to nine folds less after two months of storage at 4°C as

compared to the wild type tubers. In other words, repression of cold sweetening was noted

in transgenic tuber (Loberth *et al.*, 1998).

In our country, no comprehensive biochemical and molecular approaches have been

adopted so far to understand and inhibit the process of cold-induced sweetening in potato

tubers suitable to our own agro-climatic conditions. However, the details of starch

phosphorylation in the developing potato tubers are not clearly known at biochemical and

molecular level. Now rigorous attempts should be made to understand the cold sweetening

phenomenon at biochemical level. Attempts to be made to understand the overall starch

phosphorylation status in the developing mini-tubers from the above varieties and to find

out the varietal differences with respect to this attribute. At the same time, we need to

understand the metabolic aspects of starch phosphorylation at biochemical and molecular level.

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Keeping in view with the cold sweetening problem and its possible correlation with starch phosphorylation status in potato tubers, the following objectives were formulated for

this dissertation work:

Micropropagation of various potato cultivars, hardening, acclimatization and cultivation in the field

Harvesting of developing and mature mini-tubers from field grown plants

Isolation of starch granules from the mini-tubers of various potato cultivars

Determination of total phosphorus in starch samples among different potato cultivars

Comparison of the status of the starch phosphorylation in the various developing tubers

with in a particular variety as well as among different varieties

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3. MATERIALS AND METHODS

3.1 MATERIALS

3.1.1 PROCUREMENT OF THE MATERIALS

The germplasm of various potato cultivars such as Kufri Chipsona-1 (CS-1), Kufri

Chipsona-2 (CS-2), Kufri Chandramukhi (KCM), Kufri Jyoti (KJ), Kufri Pukhraj (PR)

Kufri Ashoka (AS) were procured from central potato research institute (CPRI), Shimla

except cultivar Desiree and routinely maintained in our laboratory on modified MS

medium.

The required chemicals were purchased from Sisco Research Laboratory Pvt. Ltd. Mumbai,

Qualigens Fine Chemicals, Merck, CDH, New Delhi, HiMedia Laboratories Pvt. Ltd.

Mumbai and Various glasswares from Borosil and plasticwares from Tarson Pvt. Ltd.

Kolkata. All the chemicals used in this study were of analytical grade (AR).

3.1.2 MEDIA USED IN MICROPROPAGATION

Shoot Multiplication Medium (SM-1):

Major salts concentration (mg/l)

KNO₃ 1900
NH₄NO₃ 1650
MgSO₄.7H₂O 370
CaCl₂.2H₂O 440
KH₂PO₄ 170

Minor Salts

MnSO₄.H₂O 22.3
ZnSO₄.7H₂O 8.6
H₃BO₃ 6.2

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KI 0.83

Na₂MoO₄.2H₂O 0.25

CuSO₄.5H₂O 0.025

CoCl₂.6H₂O 0.025

Na₂Fe-EDTA 30.0

Additives

Thiamine HCl 0.4 mg

Nicotine Acid 0.5 mg

Pyridoxin HCl 0.5 mg

Ca-Pantothenate 2.0 mg

Glycine 2.0 mg

Myo-inositol 100 mg

Sucrose 30 mg

NAA 0.01 mg

BAP 0.01 mg

GA₃ 0.25 mg

Agar 0.7 % (w/v)

pH adjusted to 5.8

3.1.3 Phytohormones

For Cytokinins: Dissolve cytokinin such as BAP (benzylaminopurine), in 2.5 ml of 0.5N

HCl, heat gently and made to the volume. Adjust the pH to about 5.0. Concentration of

stock solution of BAP was 0.1 mg/ml.

For Auxins: Dissolved auxin such as 2,4-D (2,4-Dichlorophenoxyacetic acid) and NAA

(α -Naphthaleneacetic acid) in 2.5 ml of 95 % ethanol or 2.5 ml of 1N KOH, stirred gently

and made up the volume by adding distilled water. Adjusted the pH to 5.0 and stored at

4°C. Concentration of stock solution of 2,4-D was 1 mg/ml and concentration of stock

solution of NAA was 0.1 mg/ml.

Gibberellic acid was prepared by dissolving it in distilled water.

3.2 METHODS

3.2.1 PREPARATION OF SHOOT MULTIPLICATION MEDIUM

→ SM-1 is a modified MS medium and was prepared by adding all the components in

required amounts.

→ The pH of the medium was adjusted at 5.8 and then agar was added to it at a

concentration of 0.7 % (w/v).

→ This was then melted and ~50.0 ml of the medium was dispensed in each tissue

culture bottle.

→ These bottles were then autoclaved at 121°C for 20 minutes at 15 psi, followed by

their storage at 25°C.

3.2.2 MICROPROPAGATION OF POTATO CULTIVARS

Micropropagation of different potato cultivars such as Kufri Chipsona-1, Kufri

Chipsona-2, Kufri Ashoka, Kufri Pukhraj, Kufri Chandramukhi, Kufri Jyoti, and CV

Desiree was done on SM-1 media on a routine basis.

→ The laminar airflow chamber was thoroughly cleaned with alcohol.

→ Under aseptic conditions, young and tender plantlets of the above said varieties

were taken out on a sterile glass plate, with the help of sterile forceps. These plantlets were not surface sterilized as these were already being maintained under *in vitro* conditions.

→ Using a sterile scalpel, the roots of these plantlets were excised.

→ The leaves were removed and finally the shoot part was cut into small segments,

each segment retaining at least one node.

→ Maintaining the correct polarity of the cut fragments, these were individually

inoculated in the solidified MS-1 medium (About 10-12 such explants were inoculated per bottle).

→ The bottles were sealed and labeled carefully and were finally kept in the culture

room under maintained conditions of temperature (25°C) and light (16 hrs. light,

8 hrs. dark) and the growth of the inoculated explants was monitored regularly.

3.2.3 HARDENING AND ACCLIMATIZATION

→ All the above rooted potato varieties maintained by tissue culture was gently

washed with warm water to remove the agar from the roots.

→ These plantlets were treated with 1 % (w/v) Bavistine for 5- 10 and were transferred to the plastic protrays containing the potting mix Solirite.

→ These plantlets were further shifted to highly humid conditions in a polyhouse for

two weeks with the temperature maintained at 25°C and relative humidity of 85 to

90 %.

→ These hardened plantlets were then shifted to shade house for 6-7 days with

temperature maintained at 25°C and relative humidity of 65 to 70 %.

3.2.4 CULTIVATION OF PLANTLETS IN THE FIELD

→ These plantlets were then cultivated in the field in different bed for different varieties.

→ Their growth was monitored regularly along with the application of water, fungicides and fertilizers whenever required.

→ Initially after two months, minitubers of varying sizes from each variety were

harvested, and then tubers were harvested after regular interval of time of 14-20

days with their increasing developmental stages.

→ After harvesting, every time minitubers were washed gently with water, dried on

the folds of blotting paper carefully, to avoid mixing.

→ Finally various studies were carried out on them.

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All the potato varieties were cultivated in the same agroclimatic conditions of sunlight,

relative humidity, water balance, fertilizer dose etc.

3.2.5 ISOLATION OF STARCH GRANULES FROM POTATO TUBERS

The protocols that were followed for the isolation of starch granules is as follows:

→ Weighed 6.0 g – 10 g of potato tubers and crushed them using pestle and mortar to

form a paste; formed a uniform using ice-cold double distilled water.

→ Filtered the homogenous slurry through a double-layered muslin cloth (which was

washed with double distilled hot water) into a 50 ml of centrifuge tubes and made

up the volume to 50.0 ml using ice-cold water.

→ Incubated the tubes in ice for at least 4-5 hours and then decanted the supernatant carefully to avoid any loss of starch granules.

→ The white colored starch granules were then mixed with nearly 30.0 ml of ice-cold double distilled water, mixed thoroughly and centrifuged at 5000 to 6000 rpm for 10 minutes to give the first water wash.

→ Similarly gave a second water wash and decanted the supernatant without disturbing the pellet.

→ To the pellet added 15.0 ml of acetone, mixed and vortexed the solution and centrifuged at 6500rpm for 15 minutes to give the first acetone wash. Similarly gave second wash and finally decanted the supernatant carefully.

→ Air-dried the starch granules by incubating the tubes at 37°C overnight.

→ Finally weighed the dried starch granules and quantified them per 100 g fresh tuber weights.

3.2.6 EXPERIMENT TO CHECK THE PRESENCE OF ANY FREE PHOSPHATE IN STARCH SAMPLES

The phosphate content in potato tubers is exceptionally high as compared with other plant storage organs. The phosphate groups are bound as monoesters at the C-6 and at the C-3 positions of the glucose residues. Therefore, the phosphate present in the

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potato starch is the bound phosphate. However, to check if there is any free phosphates in starch, following experiment was carried out:

→ Nearly 10.0 mg of starch powder was thoroughly suspended in water and kept for 20-25 minutes at room temperature, so that the free phosphate, if any, would come in to the solution.

→ Then the suspension was centrifuged at full speed and the supernatant was assayed for phosphorus by ascorbic acid-ammonium molybdate assay. However, the results showed that amount of free phosphate in the supernatant was negligible and that was almost comparable to the blank sample.

3.2.7 DETERMINATION OF TOTAL STARCH-BOUND PHOSPHORUS

In this method the phosphomolybdate complex is reduced by ascorbic acid. Essentially, this method is based on the principle of colorimetric estimation of inorganic

phosphate. Inorganic phosphate reacts with ammonium molybdate in an acid solution to form phosphomolybdate complex, addition of a reducing agent like ascorbic acid, reduce the molybdenum, in the phosphomolybdate complex, to give a blue color. In this reaction, the uncombined molybdic acid will remain unaffected. This ascorbic acid method is very sensitive, by which one can easily determine inorganic phosphate as low as 0.01 micromole.

Here, we need to determine the total starch-bound phosphate. For this ashing procedure of starch was adopted to determine bound phosphorus. Complete ashing enables the conversion of organic phosphates to inorganic phosphates. Basically, the ashing procedure was coupled with the very sensitive inorganic phosphate method.

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(A) REAGENTS

REAGENT A: Ascorbic acid, 10 %

Weighed 1.0 g of ascorbic acid and dissolved in 10 ml of distilled water.

REAGENT B: 0.42 % Ammonium molybdate.4H₂O in 1 N H₂SO₄.

Weighed 0.084 g of ammonium molybdate, to it added 0.572 ml of concentrated H₂SO₄

and then volume made up to 20 ml.

MIX: Mixed 1.0 ml of reagent A and 6.0 ml of reagent B before use.

STANDARD KH₂PO₄ (100 mg P in 1.0 liter solution i.e. 100 ppm)

Dissolved 87.9 mg of KH₂PO₄ in 80 ml of distilled water, then added 5.0 ml. of 7 N H₂SO₄ to it and then volume made up to 200 ml.

MAGNESIUM NITRATE: Weighed 0.4 g of Magnesium nitrate [(MgNO₃)₂.6H₂O]

dissolved in 4.0 ml of 95 % alcohol.

OTHERS (as describe in the methods):

0.1 N H₂SO₄, Conical flask, Pipettes, Micropipette, Water bath, Dispenser, Centrifuge

tubes, Glass rod, microfuge tubes, Centrifuge, Test tubes, Bunsen burner, Weighing

balance, Butter paper, Spatula etc.

(B) PROCEDURE

→ Weighed 5.0 to 7.0 mg of starch granule, isolated from potato tubers, and suspended

in 1-2 drops of water. The material was taken to dryness and completely charred

over a strong flame of gas burner.

→ The tubes were then allowed to cool and 0.4 ml of 0.1 N H₂SO₄ was added. The

tubes were tightly capped with plastic caps and heated in boiling water bath for 10

minutes, to ensure solubilization as well as conversion of any pyrophosphate formed during ashing, to inorganic phosphate.

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→ The solution was allowed to cool and then transferred to 1.5 ml microfuge tubes

from glass tubes, then rinsed the glass tubes by addition of 0.1 N H₂SO₄ and volume

made up to 0.5 ml.

→ Sequential addition was done in the following way, for a reaction volume of 0.5 ml:

~ ^ 300 µl aliquots of the sample taken for assay.

~ ^ 180 µl of 0.1N H₂SO₄

~ ^ Made up the volume to 900.0 µl by addition of water

~ ^ In case of blank, 900 µl of 0.1N H₂SO₄ and 900 µl of water were mixed together

→ To each reaction tube, added 2.1 ml of the MIX and to the blank 4.2 ml of MIX,

vortexed the tubes and incubated in water bath at 45°C for 20 minutes.

→ Cooled the tubes and read absorbance at 820nm.

The procedure was repeated for three times with almost reproducible results.

NOTE:

(i) The Standard Curve: For the preparation of standard curve KH₂PO₄ (100 ppm)

solution was used and the following amounts of phosphorus were taken in different tubes as

given here: 1.5 µg, 3.0 µg, 4.5 µg, 6.0 µg, 7.5 µg, 9.0 µg. The other ingredients and

reaction volume in each case were same as followed in the unknown samples.

(ii) Other ashing procedures:

→ Few starch samples were treated with few drops of water or conc. H₂SO₄ and then

charred in the muffle furnace at very high temperature of 600°C – 615°C for two

hours using silica crucible. The results were almost comparable as obtained by the

ashing procedure discussed in the previous section.

→ The charring of a few starch samples was also carried out by making suspension in

alcoholic magnesium nitrate [$\text{Mg}(\text{NO}_3)_2 \cdot 6\text{H}_2\text{O}$] solution followed by assay of Pi as described earlier. The purpose was to check the sensitivity of the assay procedure.

Nearly comparable data was obtained in all these ashing procedures.

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4. RESULTS AND DISCUSSION

Tuber development in potatoes is a very complex biological event. Various types of

metabolic activities are operational, till date detailed mechanism both at biochemical and

molecular level is not known clearly. In addition varietal differences are also there. Starch

phosphorylation occurs during tuber development and status of starch phosphorylation is

believed to play a crucial role with respect to cold-sweetening phenomenon in the potato

tubers during prolong storage at low temperatures. So far we do not know clearly the status

of starch phosphorylation in the tubers of our own potato cultivars. Keeping in view with

the above, here attempts have been made to know the status of starch phosphorylation in

the developing as well as mature tubers. In addition, the starch phosphorylation was also

studied in the potato tubers after transferring to 20°C for four weeks. Mini-tubers at various

stages of development were harvested from field-grown seven cultivar varieties for this

study. The results obtained are given in the following sections.

4.1 MICROPROPAGATION

The following potato varieties namely Kufri Chipsona-1, Kufri Chipsona-2, Kufri Ashoka,

Kufri Pukhraj, Kufri Chandramukhi, Kufri Jyoti, CV Desiree were routinely micropropagated on the shoot multiplication medium (SM-1), which was modified MS

medium. It contains various phytohormones such as auxin, cytokinin and gibberlin. Profuse

shoot growth was observed within three weeks of inoculation, along with the root growth to

some extent. Young and tender leaves started appearing on the stem within the same time

period as shown in the **Fig. 1, 2, 3 and 4**. The growth of the inoculated explants was

observed regularly and these were checked for contamination. Usually contamination was less frequent. Further subculturing was carried out for every four to five weeks.

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4.2 HARDENING, ACCLIMATIZATION AND CULTIVATION

The micropropagated plantlets with tender roots were hardened and acclimatized in the potting mix-soilrite for two weeks. Soilrite consisted of peat moss to absorb water and

vermiculite along with perlite helps in aeration. While this step was carried out, the

plantlets were carried under high humid conditions, to prevent desiccation. Finally, these

were planted on the trial plots within our own campus. Weeds were manually removed

when required. The moisture level was maintained by spraying water almost twice a day,

under the dry weather conditions.

Growth characteristics of different cultivars were noted time to time are given below:

Kufri Chipsona-1 (CS-1): Green short plants with moderately thin stems, tripinnately compound leaves with smooth lamina.

Kufri Chipsona-2 (CS-2): Dark green, erect and tall plants with compound leaves, undivided and rough lamina and smooth margins. Dense growth was observed.

Kufri Chandramukhi (KCM): Short and light green plants with thin stem, tripinnately compound leaves and plane lamina.

Kufri Jyoti (KJ): Short to moderate tall, light green plants with thin stem, small to very large, shiny, compound leaves. Lamina was curved inside with pointed apex.

Kufri Pukhraj (PR): Moderately tall, dark green plants with thick stem compound leaves with thin and large lamina and smooth margins. Apex was slightly blunt.

Kufri Ashoka (AS): Light green plant with bushy appearance, thick stem, thin compound leaves with pointed apex and smooth lamina and margins.

Cultivar Desiree (DE): Short to moderately tall, light green plants with both simple and

compound leaves. Lamina was not smooth.

A few field-grown plants are shown in **Fig. 5, 6, 7 and 8**

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4.3 HARVESTING OF MINI-TUBERS AT VARIOUS STAGES OF DEVELOPMENT

In this study seven important potato cultivars were selected to carry out experiments. Minitubers

were harvested from the field-grown plants of the above varieties. It may be noted

here that properly hardened and acclimatized plantlets of various potato cultivars served as

planting material in the field. Plantation was carried out in the second week of Nov. 2003.

The first maturing tuber samples (referred to as TS-01) from various cultivars were

collected after nearly eight weeks of cultivation. The weight of these TS-01 mini-tubers

from different varieties was in the range of 0.3 g – 0.4 g. Similarly, at different time

intervals growing mini-tubers from all these varieties were collected (referred to as TS-02

to TS-07). The final harvesting of tuber samples (referred to as TS-08) was done in the first

week of March 2004 (the entire period of cultivation in the field was nearly four months).

The mature tubers were also transferred to 20°C for 4 weeks, which are referred to as

TS-09. A few harvested mini-tubers are shown in **Fig. 9,10,11 and 12.**

Various potato tuber

samples as harvested above along with their range of weights are given in the **Table 4.1**

Table 4.1 Various potato mini-tuber samples

S.No. Tuber Sample Range of Tuber Weight

1. TS-01 0.3 g - 0.4 g
2. TS-02 0.8 g - 1.0 g
3. TS-03 1.3 g - 1.5 g
4. TS-04 2.5 g - 2.7 g
5. TS-05 10.0 g - 11.0 g
6. TS-06 14.0 g - 15.0 g
7. TS-07 20.0 g - 22.0 g
8. TS-08 30.0 g - 32.0 g
9. TS-09 28.0 g to 30.0 g

35
36
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In each potato variety, the weight of the developing minitubers was found to be proportional to their size. Morphologically differences were noted with respect to the shape and size of minitubers from different cultivars. The minitubers were more or less spherical in shape with delicate tuber skin and light brown in color, except in the case of cultivar Desiree of which tubers were reddish in color.

4.4 QUANTIFICATION OF STARCH GRANULES IN THE TUBERS

Here a rapid, convenient and simple method was adopted for isolation of good quality of starch granules (section 3.2.5) from various tuber samples (TS-01 to TS-09) of each potato cultivar. It may be mentioned here that extensive washing steps were employed using both cold water and organic solvent like acetone during the isolation of starch granules. This helped to remove various organic and inorganic impurities in the starch preparations. Weight of each starch preparation was taken after extensive drying at 37°C. The starch granules recovered were clean, white-colored, and powdery in form. The starch yield was then calculated in gm per 100 gm of tuber weight as shown in the **Table 4.2**.

Table 4.2 Starch yield from potato tubers at different developmental stages

(Calculated in gm per 100 gm of tuber weight)

Variety	TS-01	TS-02	TS-03	TS-04	TS-05	TS-06	TS-07	TS-08	TS-09
CS-1	5.7	4.5	4.5	4.8	6.5	8.9	9.8	13.9	6.3
CS-2	5.6	4.7	6.2	7.0	6.6	10.8	12	6.8	7.5
KJ	5.0	3.8	4.7	7.8	4.2	6.5	6.8	6.1	4.7
KCM	3.4	5.6	5.3	5.9	7.7	11.6	9.3	8.8	7.8
PR	3.0	5.3	3.7	4.1	4.9	5.3	8.0	7.7	7.5
AS	3.5	3.0	7.5	4.2	7.3	8.1	6.6	11.5	9.0
DE	4.1	5.0	4.7	4.5	7.8	9.8	9.6	12	7.6

TS – Tuber Sample

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Close inspection of the data given in the above table shows that in almost all varieties starch yield was found to increase in the later stages of development. The maximum starch yield of was noticed in TS-08 of Kufri Chipsona-1 (13.9 g /100g). Whereas minimum starch yield (3.0 g /100 g) was noted in TS-01 of Kufri Pukhraj and TS-02 of Kufri Ashoka. However, suitable processing varieties like Kufri Chipsona 1, Kufri Chipsona 2 and Kufri Ashoka showed more starch yield as tuber dry matter could be directly correlated with starch content. In all the above varieties storage of tubers samples at 20°C leads to decrease of starch yield as compared with the tubers before storage. Although it is not true for all, the low yield is possibly due to starch degradation, respiration and other metabolic activities functional in the potato tubers.

4.5 DETERMINATION OF TOTAL PHOSPHORUS FROM STARCH GRANULES

Starch is composed of two polymers, amylose and amylopectin. The amylose molecules are essentially linear, whereas amylopectin molecules are highly branched and often contain some amount of covalently bound phosphate. In addition, amylopectin depending upon the plant organ where it is manufactured contains different level of phosphate monoesters. The phosphate content in potato tubers is exceptionally high as compared with other plant storage organs. The phosphate groups are localized as monoesters at the C-6 and C-3 positions of the glucose residues in starch. Here attempts have been made to determine the total starch-bound phosphate in the tubers of different potato cultivar in its various developmental stages. The purpose is to know the phosphorylation status in various starch preparations. The total bound-phosphate i.e. organic phosphates were converted into

inorganic phosphate through complete ashing procedure on the gas burner. Then phosphate

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was determined by the Ascorbic acid-ammonium molybdate method. Another ashing

procedure carried out was by, charring of few starch samples in the muffle furnace by using

silica crucible at 600-612°C for two hours. Nearly comparable data was obtained in both of

the ashing procedures The starch bound-phosphate was estimated in the tubers at various

stages of development from seven varieties. Here, the amount of total starch-bound

phosphorus was determined per 100 g of starch granule preparations. For each variety as

discussed earlier, the amount of bound phosphorus was measured from all starch

preparations as shown in **Table 4.2**.

In case of Kufri Chipsona-1, the amount of total phosphorus was found to vary

considerably as shown in **Fig. 13**. The bound phosphorus content was found to be higher in

the starch preparations isolated from very early maturing tubers (around 60 mg per 100 g of

starch). In case of Kufri Chipsona 2, the phosphorus content was found to be in the range of

30 mg to 40 mg per 100 g of starch (**Fig. 14**). The four cultivars namely Kufri Jyoti, Kufri

Chandramukhi, Kufri Pukhraj and Kufri Ashoka showed similar pattern. The phosphorus

content more or less was lying in the range of 30 to 50 mg per 100 g of starch as shown in

Fig. 15, 16, 17 and 18. It may be noted that phosphorus content appeared to be in the

higher side as compared with Kufri Chipsona-1 except its early maturing stages. In the case

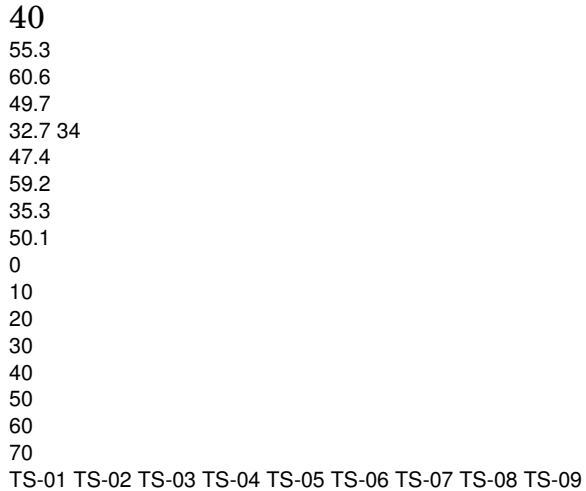
of Kufri Ashoka maximum phosphorus content was noted at the early stages of tuber

development. It is of interest to note that phosphorus content was highest i.e., 70.0 mg in

the very early stage and then decreased notably in the other stages of development in case

of cultivar Desiree as shown in **Fig. 19**. All these data collectively suggest that varietal

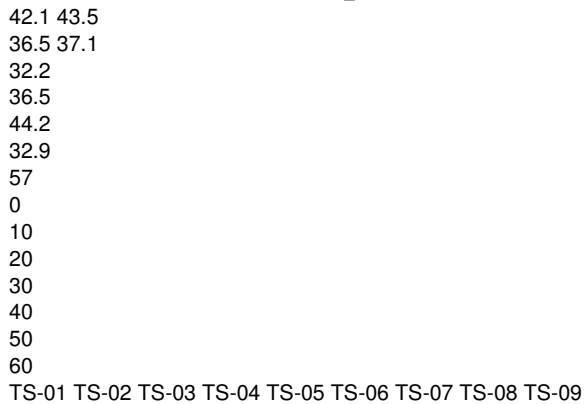
difference does exist with respect to starch-bound phosphorus content. It is apparent that starch phosphorylation occurs throughout the tuber development. The level of this biochemical activity seems to be higher in the early stage of development. At present the



Various stages of tuber development
Amount of Starch-bound P
(in mg)

Fig. 13. Comparison of total phosphorus content in various starch samples

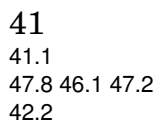
Cultivar: Kufri Chipsona-1



Various stages of tuber development
Amount of Starch-bound P
(in mg)

Fig. 14. Comparison of total phosphorus content in various starch samples

Cultivar: Kufri Chipsona-2



46.5 46.1
 34.9
 46.3
 0
 10
 20
 30
 40
 50
 60
 TS-01 TS-02 TS-03 TS-04 TS-05 TS-06 TS-07 TS-08 TS-09

Various stages of tuber development

Amount of Starch-bound P

(in mg)

Fig. 15. Comparison of total phosphorus content in various starch samples

Cultivar: Kufri Jyoti

39.8
 36.2
 34
 40
 43.8
 36.5 37.5
 34.6
 41.1
 0
 5
 10
 15
 20
 25
 30
 35
 40
 45
 50
 TS-01 TS-02 TS-03 TS-04 TS-05 TS-06 TS-07 TS-08 TS-09

Various stages of tuber development

Amount of Starch-bound P

(in mg)

Fig. 16. Comparison of total phosphorus content in various starch samples

Cultivar: Kufri Chandramukhi

42
 37.5
 34.1
 40.3
 36 36.7
 39
 34.7
 29
 36.8
 0
 5
 10
 15
 20

25
30
35
40
45

TS-01 TS-02 TS-03 TS-04 TS-05 TS-06 TS-07 TS-08 TS-09

Various stages of tuber development

Amount of Starch-bound P

(in mg)

Fig. 17. Comparison of total phosphorus content in various starch samples

Cultivar: Kufri Pukhraj

50.8 51.4

35.2 35.5

44.9

41.4 41.2

50.7

46.5

0

10

20

30

40

50

60

TS-01 TS-02 TS-03 TS-04 TS-05 TS-06 TS-07 TS-08 TS-09

Various stages of tuber development

Amount of Starch-bound P

(in mg)

Fig. 18. Comparison of total phosphorus content in various starch samples

Cultivar: Kufri Ashoka

43

70.6

50

43.4 42

35.6

38.2

40.5

31

43

0

10

20

30

40

50

60

70

80

TS-01 TS-02 TS-03 TS-04 TS-05 TS-06 TS-07 TS-08 TS-09

Various stages of tuber development

Amount of Starch-bound P

(in mg)

Fig. 19. Comparison of total phosphorus content in various starch samples

Cultivar: Desiree

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role of starch granule-bound protein (R1) possibly the known member causing phosphorylation of starch. Apart from this, other genes could be functional during tuber development. As R1 protein is known, we may focus on its expression both at the level of transcription and translation in the developing tubers. It may answer partly the varietal difference with respect to starch-bound phosphorus content. Starch-bound phosphorus content appeared to be higher in potato tubers stored at 20°C (here TS-09) for a few weeks. The molecular mechanism pertaining to starch degradation as well as phosphorylation under storage condition of potato tubers needs to be further understood. The differential degradation of amylose and amylopectin fractions may be involved during prolonged storage. Finally, a comparative study was also made among different cultivars as shown in **Fig. 20**. In all the potato cultivars, the amount of total bound phosphorus was found to be higher along with considerable variation in the early stages of tuber development if compared with respective mature tubers. The bound phosphorus content in the starch preparations isolated from 20°C stored tubers (TS-09) were almost comparable among the varieties and similar to the trend noted in the very early maturing tubers (TS-01). The variation within a variety may be due to alteration of the amylose to amylopectin ratio. Apart from this, it may be considered that during tuber development some other components or non-phosphorus impurities could be associated with starch granules, which were co-purified with starch preparations. The present study gives us an idea on the status of starch phosphorylation in the developing tubers from potato cultivars suitable to our own agro-climatic conditions. This will help us further to undertake biochemical and molecular studies on starch phosphorylation at developmental stages and

eventually to find out possible correlation between the status of starch phosphorylation and cold-induced sweetening.

45

0

10

20

30

40

50

60

70

80

CS-1 CS-2 KJ KCM PR AS DE

Potato Varieties

Amount of Starch-bound P

(in mg)

TS-01

TS-08

TS-09

Fig. 20. Comparison of total starch-bound phosphorus content in the tubers of various potato cultivars

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