

**DESIGN OF ION TRANSPORTER-BASED ON POLYPEPTIDE
MARKERS FOR DETECTION OF SALT TOLERANCE IN
PLANTS**

Thesis

Submitted by

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Under the supervision of

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DECLARATION

I Sambit Majumdar, hereby declare that the thesis entitled “**Design of ion transporter-based on polypeptide markers for detection of salt tolerance in plants**” submitted by me, for the degree of Master of Technology in Biotechnology of Thapar Institute of Engineering and Technology, Patiala is a record of honest work carried by me under the supervision and guidance of supervision of Dr. Debajyoti Dutta, Assistant Professor at the Department of Biotechnology, Thapar Institute of Engineering and Technology, Patiala. I have not submitted the matter embodied in this thesis for the award of any other degree.

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CERTIFICATE

It is to be certified that the thesis entitled “**Design of ion transporter-based on polypeptide markers for detection of salt tolerance in plants**” submitted by Sambit Majumdar (602204016) to the Department of Biotechnology, Thapar Institute of Engineering and Technology Patiala, in partial fulfilment for the award of the degree Master of Technology in Biotechnology, is a record of original work carried out by him from 1st August 2023 to 29th July 2024 under the supervision and guidance of supervision of Dr. Debajyoti Dutta, Assistant Professor at the Department of Biotechnology, Thapar Institute of Engineering and Technology, Patiala. The matter embodied in the thesis work has not been submitted for the award of any degree.

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LIST OF ABBREVIATION USED

At	<i>Arabidopsis thaliana</i>
ATP	Adenosine Triphosphate
BLAST	Basic Local Alignment Search
DNA	Deoxyribose Nucleotide Phosphate
H ₂ O ₂	Hydrogen Peroxide
SOS1	Salt Overall Sensitivity 1
HKT1	High Affinity K ⁺ Transporter 1
NHX1	Sodium/ Hydrogen Exchanger 1
MAFFT	Multiple Alignment Using Fast Fourier Transform
MSA	Multiple Sequence Alignment
NaCl	Sodium Chloride
NCBI	National Center for Biotechnology Information
PDB	Protein Data Bank
PM	Plasma Membrane
ROS	Reactive Oxygen Species
SS	Salinity Stress
AtSOS1	<i>Arabidopsis thaliana</i> Salt Overall Sensitivity
TtSOS1	<i>Triticum turgidum</i> Salt Overall Sensitivity
OsSOS1	<i>Oryza sativa</i> Salt Overall Sensitivity
NtSOS1	<i>Nicotiana tabacum</i> Salt Overall Sensitivity
StSOS1	<i>Solanum tuberosum</i> Salt Overall Sensitivity
AtHKT1	<i>Arabidopsis thaliana</i> High Affinity K ⁺ Transporter
GmHKT1	<i>Glycine max</i> High Affinity K ⁺ Transporter

GaHKT1	<i>Gossypium arboretum</i> High Affinity K ⁺ Transporter
SIHKT1	<i>Solanum lycopersicum</i> High Affinity K ⁺ Transporter
StHKT1	<i>Solanum tuberosum</i> High Affinity K ⁺ Transporter
RMSD	Root Mean Square Deviation
RMSF	Root Mean Square fluctuation
Rubisco	Ribulose 1,5 biphosphate carboxylase/ oxygenase
PEPCase	Phosphoenolpyruvate carboxylase
NADP-ME	NADP- malic enzyme

Abstract

Salinity is one of the environmental stresses that impact growth and development of plants. Sodium chloride is one of the predominant salt responsible for salinity. Beyond the allowable limit in the soil sodium chloride exerts osmotic and ionic stresses. Upon perception of the salinity stress plants undergo different mechanisms such as ion homeostasis, signaling, redox homeostasis and transport of the saline solute in the cell to overcome the salinity. It is managed by the different membrane proteins present in the plant cell that help with sodium distribution, sodium extrusion, sodium compartmentalization, osmotic balance and cellular damage repair. Some of the plant's conserved membrane proteins that help to maintain the plants in salinity stress are Aquaporins, HKT1 (High-Affinity K⁺ Transporter 1), NHX1 (Na⁺/H⁺ exchangers 1), and SOS1 (Salt Overly Sensitive 1). In this thesis, we have studied two membrane proteins SOS1 and HKT1 protein that help to maintain salinity stress in plants. For the above studies, we have selected a few plants where the role of SOS1 and HKT1 are shown in salt tolerance including the model plant *Arabidopsis thaliana*. We have used different bioinformatics tools and servers to identify the conserved amino acid sequence and design the most appropriate polypeptide markers. The designed polypeptide markers can be used to determine the salinity stress in multiple crops.

Introduction

Plant stress can be defined as biotic or abiotic stresses that affects by disrupting of proper functioning of the plant's membrane (Zhang *et al.*,2022; Dresselhaus *et al.*, 2018). Soil salinity, drought and temperature are some examples of abiotic stress whereas pathogens, parasites, allelopathy and weeds are some examples of biotic stress. (Zhu *et al.*, 2016; Roeber *et al.*, 2021).

Salinity stress in plants is a significant issue for crop productivity around the world. These stresses impact the plant productivity in the same way as the abiotic stress. Other than the climate change, degradation of the land and droughts are responsible for increase in soil salinity (Zhu *et al.*, 2016). Sometimes, these stresses affect the plants so severely that they get over damaged or die due to stresses. It also creates conductivity and porosity in soils, which causes low water potential in plants. Salinity stress impacts the plants by creating osmotic stress, ion toxicity, oxidative stress thereby affecting photosynthesis and carbon assimilation, stomatal regulation and transpiration, secondary metabolism and hence leading to loss in crop productivity.

As data reported in 2016 shows approx. 60% of the world's total land area is suitable for cultivation, of which 80% are used for crops and 6% are used for rangelands (Mythili *et al.*, 2016). According to estimates, salinity will impact 50% of the world's arable land by 2050 (Bartels *et al.*,2005; Kumar *et al.*, 2020). According to data reported in 2017, approximately 2.1% of the India's total land area or 7 million hectares is impacted by salt of these 3 million hectares are salty and the remaining 4 million hectares are sodic (Arora *et al.*, 2016;). In all developing countries, intensive agricultural practices are responsible for deteriorating agricultural lands. One of the crucial factors for the deduction of plant growth and production is the salt stress (Szabolcs *et al.*, 1974).

Soil salinity has various types depending on the types of salt, such as sodic soils, saline soils, and saline-sodic soils. When the concentration of sodium ions increases as compared to other cations in the soil, it is considered sodic soil. The decrease of water flow in the soil initiates the accumulation of salts (Qadir *et al.*,2007). When water-soluble salts are present in the soil, in amounts that they affect normal crop production, then it is considered saline soil. This type of soil pH is always less than 8.5 (Negacz *et al.*, 2022). If the concentration of both saline and sodic levels is high in soils, it is considered saline-sodic soils. This type of soil is poor in soil

structures, has an imbalance in nutrient content, moderately alkalinity in pH and many other characteristics (Qadir *et al.*,2007).

Salinity stress started in crops due to the solubilization of salts in soil. There are some positively and negatively charged ions such as Ammonium (NH_4^+), Phosphate (PO_4^{3-}), Potassium(K^+), Calcium (Ca^{2+}), Magnesium (Mg^{2+}), Phosphate (PO_4^{3-}), Sulfate (SO_4^{2-}), Nitrate (NO_3^-), Sodium (Na^+) and Chlorine (Cl^-). Out of these ion's potassium, sodium, chloride and sulfate are the main ions present in the soil that salinity stress. NaCl containing soil causes various types of stresses in plants which leads to osmotic stress, ion toxicity, root damage, nutrient imbalance, physiological disorders, reduces growth and yields.



Figure 1: Land affected by soil salinity

Salinity effects the plants by osmotic stress and then by phytotoxic ions. Continuous intake of salt from roots creates a decrease in water conductivity in cells. Long exposure of plant roots to this process creates toxicity and nutrient imbalance in plants.

There are two different categories of plants that reveal different tolerability to salinity and also to adaptability, they are glycophytes and halophytes. Halophytes are the ones that can grow in saline soils. These types of plants can be grown in coastal areas. Some of the halophyte plants are mangroves, glasswort, seagrasses and many others. Glycophytes are simply the opposite, meaning they grow in saline soils; they are less tolerant as compared to halophytes. Some of the glycophyte plants are wheat, rice, corn, soybeans and many others.

There are various physiological responses to salinity stress such as ionic imbalance and osmotic stress, reactive oxygen species accumulation in cells, changes in photosynthesis and

carbonic metabolism, variations in water uptake and transport. Inside the cells plants maintain the low concentration of Na^+ and the high concentration of K^+ . High concentration of K^+ helps the plants for proper growth and development. During the salt stress Na^+ levels increase in the cells and create ion imbalance in plants, which disrupts the normal function of plants (Li, J. *et al.*, 2023). This imbalance can be maintained by segregating sodium, compartmentalization of sodium or by distribution of sodium from cells. There are various transmembrane protein that helps to maintain the concentration.

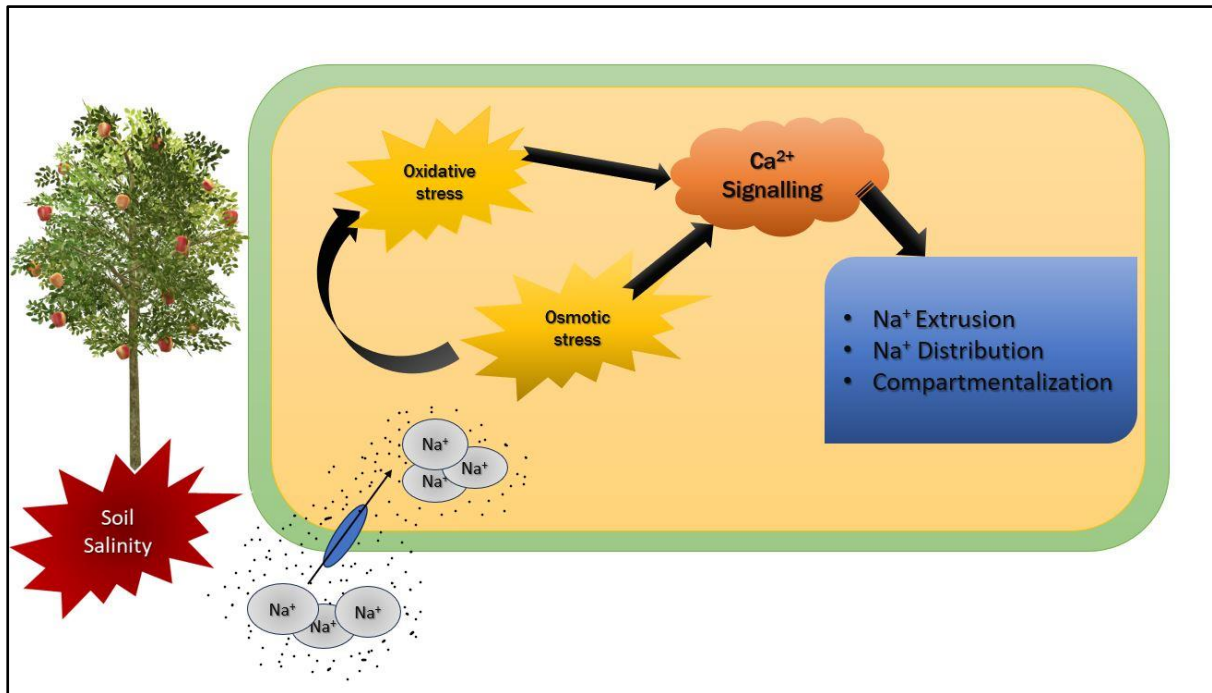


Figure 2: Mechanism to salinity stress in plants

Salinity stress is managed by Na^+ distribution, extrusion and compartmentalization, osmotic balance and cellular damage repair by the membrane proteins (Dutta *et al.*, 2023). Some critical plant membrane protein that helps in the management of salinity stress are HKT1 (High Affinity K^+ Transporter 1), NHX1 (Sodium/ Hydrogen Exchanger 1), SOS1 (Salt Overall Sensitivity 1), Aquaporins and various ion channels. Different membrane protein has different function in various membrane parts in plants. During salinity stress, certain membrane protein in plants recognize osmotic stress and activate calcium ions which signal to manage salinity stress and produce reactive oxygen species (ROS) by activating the plasma membrane enzyme RBOH, aquaporins membrane transporter transport reactive oxygen species (ROS) into the cytosol and causing oxidative stress which signals to manage salinity stress (Banik and Dutta, 2023).

During salinity stress in soil, the water uptake in plants decreases. Referring to figure 2, we can understand that concentration of salt creates the salinity in soil. This pressure impacts plants by water absorption and by plant development. Osmotic stress develops due to the variation of water potential in the soil. Reactive oxygen species (ROS) is another physiological response to salinity stress. It is a common byproduct of cells produced at low rates. During salinity stress, it is produced at high rate in the cell (Moller *et al.*, 2007; Miller *et al.*, 2010; Hossain *et al.*, 2016). They are the free radicals like superoxide and hydroxyl radicals as well as non-radical molecules. Excess amounts of reactive oxygen species (ROS) can damage the cells by damaging and by metabolic disorders. It helps to maintain the biotic and abiotic stresses in the plants (Mittler *et al.*, 2004; Hossain *et al.*, 2016).

SOS1 (Salt Overly Sensitive 1) is one of the major membrane proteins associated with the salinity tolerance. It is localized in the plasma membrane to extrudes the sodium ion from the cell. The SOS1 membrane protein is composed of 13 transmembrane domains. It helps to maintain the cell viability and it is expressed in the parenchyma cells present in the boundary between the xylem and symplast and also in the epidermal cells at the tips of the roots (Zhang *et al.*, 2023). It is responsible for the excretion of sodium ions out of the cell across the plasma membrane. The SOS1 pathway involves the binding and exchange of the Na^+/H^+ antiporter. SOS1 is a large integral membrane protein it has multiple transmembrane domains in various locations, different domains have different functions. It has been seen that on overexpression of SOS1 protein can also show the potential activity for salt tolerance in plants. Various SOS membrane protein is also available which shows a similar function to other membrane protein.

HKT1 (High affinity K^+ Transporter 1) is also a membrane protein associated with the salinity tolerance. It is also primarily localized to the plasma membrane and acts as a Na^+ transporter in the membrane (Horie *et al.*, 2001). It is composed of 4 transmembrane domain-pore loop- transmembrane domain patterns (Hamamoto *et al.*, 2015). They are expressed not only present in xylem parenchyma but also present in roots, shoots, leaves and flowers (Riedelsberger *et al.*, 2021). It functions as a uniporter or symporter for the transport of potassium ions across the plasma membrane. When the concentration of the sodium ions is high and potassium ions is less inside the cell, it helps to uptake the potassium ions from the soils to maintain the physiological characteristics inside the cells.

In this project, we have used the SOS1 and HKT1 membrane protein. We attempt to use the conserved membrane protein transporter sequences, involved in plant salt tolerance, to design polypeptide markers for the identification of salinity stress.

Review of literature

Salt stress is a major issue in the environment that affects the growth and development of plants. Presence of the sodium concentration affects the plant's growth and developments. Like animals, plants also do not require excess Sodium ions inside the cells. If present in excess amount, it affects the plant in various physiological ways such as oxidative stress, nutrient imbalance, delayed growth and development of death of the plant's cells (Assaha *et al.*, 2017; Cabot *et al.*, 2014).

Restriction of accumulation of the sodium ions and ion homeostasis are the key function of plant tolerance. Roots of the plant gets exposed to saline soil first. Sodium is then absorbed from soil to plants through roots by certain non-selective cation channel (Assaha *et al.*, 2017). Sodium ions are then transported to the shoot through the xylem with the support of different transporters and channels and then to the leaves where they impact the most (Munns *et al.*, 2007). It decreases the agricultural lands for production and increases the demand for foods with the increase in the population (Kumar *et al.*, 2020). Some of the ways by which plant manages the salinity are distribution, exclusion and compartmentalization of the sodium ions (Munns *et al.*, 2008).

Ion homeostasis in plants is defined as how plants respond to the different ions in their cells and tissues, such as potassium, sodium, calcium, magnesium, and chloride. Some of the characteristics of ion homeostasis are the uptake of ions from the soils through different transport mechanisms such as ion channels and transporters and ion partitioning by different cellular compartments to maintain the ion concentration and toxicity inside the cells. Salinity stress affects plants by disrupting the ionic balance inside the cells. When the imbalance occurs in plants due to an excess of Na^+ and Cl^- ions, they uptake the other essential ions from the soils to recover the plants from the imbalance. On accumulation of the Na^+ and Cl^- ions inside the cells it disrupts the plant cells and creates the osmotic stress. Finally, the plant's metabolism gets affected by hinderance of enzyme activities, protein synthesis and membrane integrity.

For detecting stress some markers are used such as markers, such as physiological markers, morphological markers, oxidative stress markers and molecular stress markers (Soltabayeva *et al.*, 2021). Morphological and physiological changes are followed by the orders of visualization of the stress in the plants and then by the physiologically. On the other hand, ROS molecules play a role in signaling stress and the oxidative stress markers changes are

detected with the changes in the molecular markers (Ellouzi *et al.*, 2011). Salt stress can be more easily detected in the older plants leaves than in the other parts of the plants by morphologically, physiologically and molecular changes.

During salinity stress, certain ion's transporters in plant's membrane gets activated. These transporters help to control the uptake, transport or compartmentalization of ions in the plant's cells. Some of the ion transporters that plays the critical role in regulation of the salinity stress are SOS1 protein which is known as the salt overly sensitive 1 and localized in the plasma membrane and helps in extrusion of sodium ions from cytoplasm in the exchange of the proton (H^+); NHX protein which is known as the Na^+/H^+ exchanger, and localized in the intracellular membrane in the sequestration of sodium ions inside the vacuoles; HKT1 which is also known as the High affinity K^+ transporter 1 and localized in the plasma membrane and regulates the sodium ions toxicity; AKT1 which is known as the Arabidopsis potassium ion transporter 1 localized in plasma membrane and maintain the osmotic stress in salinity condition; CNGC which is known as Cyclic Nucleotide Gated Ion Channel and maintain the calcium signaling pathways in cells during salinity stress condition; GORK which is known as the Guard Cell Outward Rectifying potassium channels localized in the plasma membrane in guard cells and helps during water loss in salt stress condition by mediating the potassium ions the cells; CAX protein which is known as Ca^{2+}/H^+ exchanger and transport the calcium ions during the salinity stress conditions.

In this project, we will mainly focus on SOS1 and HKT1 membrane transporter protein which on expression causes drastic changes in the salinity stress in plants.

The SOS (salt overly sensitive 1) system is an important mechanism that can be used by plants to resist salinity. There are three SOS proteins that are generally involved in the SOS pathway SOS1, SOS2 and SOS3 (Figure 3). All three types of protein have different functions in the SOS pathway. SOS1 is activated by the collaborative reaction of the SOS2 and SOS3. SOS3, which is also known as a calcium-binding protein, binds to the stress activated released calcium and activates the SOS2 kinase (Ali *et al.*, 2023). SOS2, which is known as the serine/threonine kinase, creates the complex kinase of the SOS2-SOS3, which combinedly interacts with the SOS1 and activates it to release Na^+ from the cells, promoting salt tolerance (Ali *et al.*, 2023).

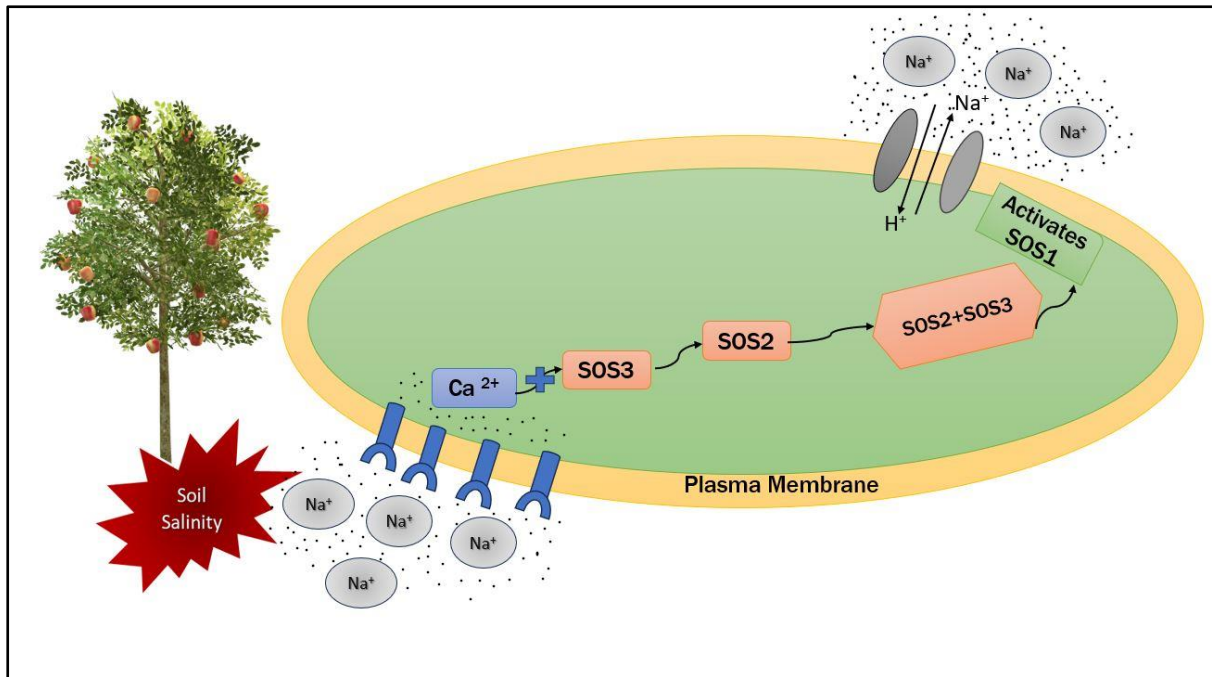


Figure 3: SOS pathway

SOS1 pathways can also be activated by the SCaBP8-SOS2 complex in shoots. SCaBP is known as the SOS3 Like Calcium Binding Protein, also known as the CBL (Calcineurin B-like), which has been identified in the Arabidopsis genome (Laun *et al.*, 2002; Gong *et al.*, 2004; Lin *et al.*, 2009). It has been observed that CBL protein can also control ion homeostasis. The interaction of the CBL1 and CBL9 activates the CIPK 23 (CBL- Interacting Protein Kinase 23). Activation of CIPK23 helps to control the K⁺ homeostasis by activating the activity of the AKT1, plasma membrane K⁺ channel.

Table 1: Different SOS1 genes studied in plants

Sl.No.	Protein Name	Organism Name	Comments	Reference
1	<i>AtSOS1</i>	<i>Arabidopsis thaliana</i>	They have identified the SOS1 gene, which functions by removing sodium from the cell in the exchange of hydrogen ions. They show that a mutation of the SOS1 gene makes the plants more sensitive to salinity stress.	Shi <i>et al.</i> ,2000

2	<i>TdSOS1</i>	<i>Triticum turgidum</i>	On overexpression of <i>Arabidopsis thaliana</i> protein complex (SOS2/SOS3) with <i>TdSOS1</i> it shows the salt tolerance in the drum wheat plant.	Feki <i>et al.</i> ,2011
3	<i>OsSOS1</i>	<i>Oryza sativa</i>	They suggested that the efflux of sodium ions in cytosolic was greater in the wild type as compared to the mutant type.	Shahzad <i>et al.</i> ,2022
4	<i>GmSOS1</i>	<i>Glycine max</i>	The SOS1 protein, as an antiporter, has an important role in sodium ion homeostasis by improving salinity in crops.	Zhang <i>et al.</i> , 2022
5	<i>NtSOS1</i>	<i>Nicotiana tabacum</i>	They have used the overexpressed <i>AtSOS1</i> gene in <i>Nicotiana tabacum</i> and compared it with non-transgenic plants and found that transgenic crops can effectively grow in salinity stress.	Yue <i>et al.</i> ,2011
6	<i>GhSOS1</i>	<i>Gossypium hirsutum</i>	They observed that the efflux of sodium ions shows the tolerance to salinity stress.	Cushman <i>et al.</i> , 2020
7	<i>SoSOS1</i>	<i>Spinacia oleracea</i>	It demonstrates that the highly expressed of SOS1 in plants shows elution of the sodium ions and its tolerability to salts in plants.	Katschnig <i>et al.</i> ,2015
8	<i>SpSOS1</i>	<i>Sesuvium portulacastrum</i>	They identified <i>SpSOS1</i> tolerability to salinity in transgenic plants. It was also observed that expression of <i>Arabidopsis thaliana</i> plants in <i>SpSOS1</i> shows better tolerance to salinity.	Fan <i>et al.</i> ,2019

9	<i>StSOS1</i>	<i>Solanum tuberosum</i>	The position of <i>StSOS1</i> was identified and recognised as a cis-element that helps in the various types of stress.	Liang and Guo, 2023
10	<i>KcSOS1</i>	<i>Karelinia caspia</i>	They state that <i>KcSOS1</i> regulates sodium transport and also regulates the potassium transporter in plants.	Guo <i>et al.</i> , 2020
11	<i>PeSOS1</i>	<i>Populus euphratica</i>	They state that <i>PeSOS1</i> helps in the control of the Na ⁺ /H ⁺ antiporter in plants.	Wu <i>et al.</i> , 2016
12	<i>CqSOS1</i>	<i>Chenopodium quinoa</i>	They show that <i>CqSOS1</i> present in plant membranes can help tolerate quinoa in saline environments.	Adolf <i>et al.</i> , 2012
13	<i>McSOS1</i>	<i>Mesembryanthemum crystallinum</i>	Expression of Na ⁺ /H ⁺ antiporters response to salinity in plants. It shows the function of antiporter in chloroplasts in plants.	Cosentino <i>et al.</i> , 2009
14	<i>NtSOS1</i>	<i>Nitraria tangutorum</i>	Overexpression of <i>NtCIPK9</i> from <i>Nitraria tangutorum</i> in <i>Arabidopsis</i> increased seed germination rate under salt stress.	Zhu <i>et al.</i> , 2011

HKT1 (High affinity K⁺ Transporter) is a non-specific cation transporter protein primarily located in xylem parenchyma plasma membrane in plants (Horie *et al.*, 2005). It belongs to superfamily of proteins such as HKT/Trk/Ktr, which are present in plants, bacteria and fungi (Corratge *et al.*, 2010). This transporter was first cloned from wheat roots (Schachtman and Schroeder, 1994). Heterologous expression of *AtHKT1* was also studied in *Xenopus laevis* oocytes, *Saccharomyces cerevisiae* and *Escherichia coli* (Uozumi *et al.*, 2000). HKT1 function by influx and distributes the Na⁺ into the root cells and prevents them from moving into the shoots. It has been reported that plant roots are getting tolerant to salinity on overexpression and transgenic overexpression of HKT1 (Han *et al.*, 2018). Based on the structure and ions selectivity, the HKT transporter has been divided into two sub-classes with differences in the amino acids of the pore domain of the proteins (Platten J.D. *et al.*, 2006; Ali

et al., 2019). HKT class 1 is considered a symporter as it transports both sodium or potassium ions in the same direction, whereas HKT class 2 is considered an antiporter as it transports both sodium and potassium ions in the opposite direction (Horie T. *et al.*, 2001; Platten J.D. *et al.*, 2006; Kato Y. *et al.*, 2001; Ali *et al.*, 2019).

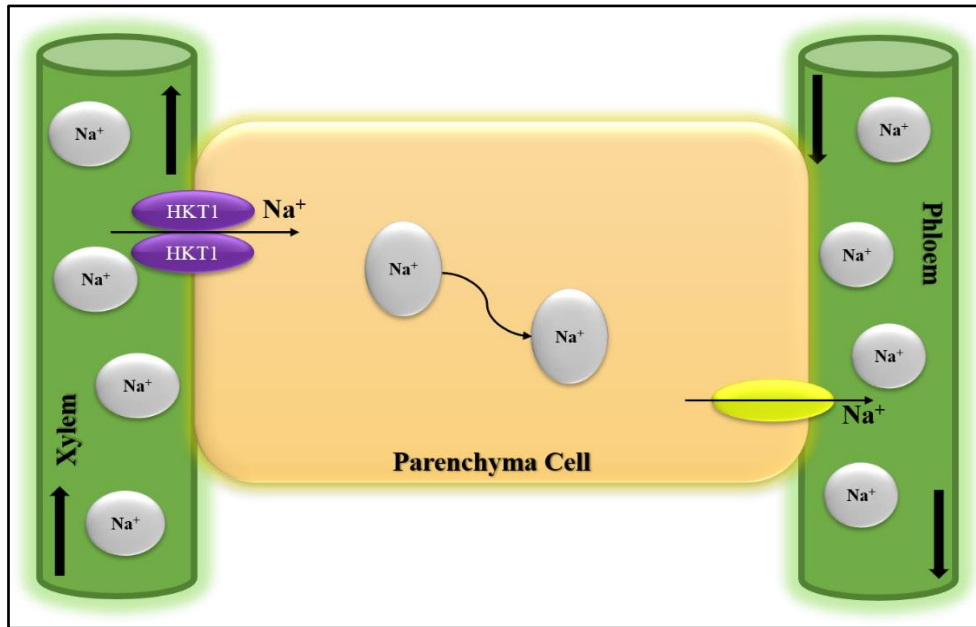


Figure 4: HKT pathway

From the figure 4 we can understand that the Na⁺ retrieve from xylem sap through shoot into the plant cell with the help of HKT1 and it experiences salinity stress then with the membrane protein Na⁺ enrich in phloem sap and returns to root.

Table 2: Different HKT1 genes studied in plants

Sl.No.	Protein Name	Organism Name	Comments	Reference
1	<i>AtHKT1</i>	<i>Arabidopsis thaliana</i>	HKT1 protein efflux the sodium ions from the xylem parenchyma cells and helps to protect them from tissue damage.	Ali <i>et al.</i> ,2021
2	<i>BoHKT1</i>	<i>Brassica oleracea</i>	Expressing the HKT in low potassium ion concentration in a salinity environment shows a	Yang <i>et al.</i> , 2023

			positive role in maintaining the potassium ion concentration.	
3	<i>GmHKT1</i>	<i>Glycine max</i>	It has been observed that on expression of HKT1, it helps to maintain the ionic ratio in roots in saline environment.	Chen <i>et al.</i> ,2014
4	<i>CaHKT1</i>	<i>Cicer arietinum</i>	On the expression of HKT1 could maintain homeostasis, osmolytes and enzymatic antioxidants in roots help them to tolerate the saline environment for the survival of plants.	Kaur <i>et al.</i> , 2022
5	<i>GhHKT1</i>	<i>Gossypium hirsutum</i>	It has been observed that on overexpression of <i>Salicornia bigelovii</i> HKT1 (<i>SbHKT1</i>) in <i>Gossypium</i> plant shows increase the uptake capacity of K ⁺ and ion homeostasis in saline environment.	Guo <i>et al.</i> , 2020
6	<i>SoHKT1</i>	<i>Spinacia oleracea</i>	It shows the HKT1 helps in the tolerance of the salinity by the movement of the sodium ions.	Katschnig <i>et al.</i> , 2015
7	<i>NtHKT1</i>	<i>Nicotiana tabacum</i>	It has been observed that on overexpression of the <i>Arabidopsis</i> HKT1 gene in root, stem and leaf, it reduced the sodium ion toxicity and make the plant tolerable to salinity.	Wang <i>et al.</i> ,2018
8	<i>SlHKT1</i>	<i>Solanum lycopersicum</i>	They demonstrate the improvement of the transport of the sodium ions in the plants.	Almeida <i>et al.</i> , 2013
9	<i>StHKT1</i>	<i>Solanum tuberosum</i>	Overexpression of the <i>AtHKT1</i> in <i>Solanum tuberosum</i> maintains the sodium ion and potassium ion concentration in the leaves.	Wang <i>et al.</i> ,2019

10	<i>HvHKT1</i>	<i>Hordeum vulgare</i>	On the expression of the HKT gene, it was observed that <i>Hordeum vulgare</i> leaves show sodium or potassium homeostasis and tolerance to salinity.	Fu <i>et al.</i> ,2018
11	<i>CaHKT1</i>	<i>Capsicum annuum</i>	The expression of <i>AtHKT1</i> controls the sodium ion influx from roots. It is distributed among the shoots of the plants in salinity stress conditions.	Yang <i>et al.</i> ,2008
12	<i>SsHKT1</i>	<i>Suaeda salsa</i>	They show that overexpression of <i>SsHKT1</i> shows the tolerability to salinity and also increases the concentration of the potassium ions.	Shao <i>et al.</i> , 2014
13	<i>VvHKT1</i>	<i>Vitis vinifera</i>	It has been observed that the accumulation of salt stress forms chlorotic leaves, but on the expression of <i>HKT1</i> , it regulates the sodium import and retransform to other lower parts of leaves and from roots, it excretes out from plants.	Lu <i>et al.</i> ,2022
14	<i>BrHKT1</i>	<i>Brassica rapa</i>	Expressing the <i>HKT</i> in low potassium ion concentration in a salinity environment shows a positive role in maintaining the potassium ion concentration.	Yang <i>et al.</i> , 2023
15	<i>LsHKT1</i>	<i>Lactuca sativa</i>	It shows that it improves the growth, water holding capacity and ratio of sodium and potassium.	Santander <i>et al.</i> ,2020

Aim of the study

To study the SOS1 (Salt Overall Sensitivity 1) and HKT1 (High-affinity K⁺ Transporter 1) protein of plants that is important for plant salt tolerance.

The objective of the study: -

- Literature survey with active role of on SOS1 and HKT1 in plant salinity.
- Designed of SOS1 and HKT1 based polypeptide markers using computational tools.

Materials and Methods

1. SOS1 and HKT1 protein responsible for plant salinity (Literature Survey)

The literature survey was done to identify the important membrane proteins HKT1 and SOS1 which have reported role in salt tolerance in plants. Additionally, identifying different plants species effective with respective proteins we have used the BLAST server. In the BLAST server [<https://blast.ncbi.nlm.nih.gov/Blast.cgi>], we have used the SOS1 and HKT1 sequences from the model plant *Arabidopsis thaliana* for identifying the respective proteins in different plants. It is to be noted that in *Arabidopsis thaliana* has only one copy of SOS1 and HKT1 is present. The sequence was selected from the server on the basis of percentage identity. The FASTA format sequence was downloaded from NCBI [<https://www.ncbi.nlm.nih.gov/>] database and UniprotKB database [<https://www.uniprot.org/>] for the respective proteins in different plants. Refer to Figure- 5.

2. Analysis of FASTA sequences

The downloaded FASTA sequences of the various plant species were aligned using multiple sequence alignment tools. Different multiple sequence alignment tools were used such as MAFFT (Multiple Alignment using Fast Fourier Transform) server [<https://www.ebi.ac.uk/Tools/msa/mafft/>], CLUSTALW server [<https://www.genome.jp/tools-bin/clustalw>], MUSCLE server [<https://www.ebi.ac.uk/jdispatcher/msa/muscle>]. We downloaded the result and for easier interpretation of the conserved region, we have used the ESPript 3.0 server [<https://esprict.ibcp.fr/ESPript/ESPript/>]. The above server provides the conserved region in graphical enhancements. Refer to Figure- 5.

3. Identification of highly conserved amino acid

Conserved regions in protein sequences are sections of the protein that remain relatively unchanged or exhibit a high degree of similarity across different species or within a set of related protein sequences. We have used computational tools like MEME (Multiple Em for Motif Elicitation) software [<https://meme-suite.org/meme/tools/meme>] are used to identify the highly conserved motif. Refer to Figure- 5.

4. Identification of the motif predicted position in common plants

We have selected a few plants that show salinity tolerance. Downloaded the FASTA sequences from NCBI or UniProt. The sequences including a selected stable sequence (Motif) were aligned using multiple sequence alignment tools. Different multiple sequence alignment tools were used such as MAFFT (Multiple Alignment using Fast Fourier Transform) server. We downloaded the result and for easier interpretation of the conserved region, we have used the ESPript 3.0 server [<https://esprict.ibcp.fr/ESPript/ESPript/>]. Refer to Figure- 5.

5. Designing the polypeptide sequence based on stability and plotting the secondary structure

From the Meme software we have noted the most conserved amino acid residues and aligned those residues to form motifs. Afterwards, a different combination of motifs was done to get the polypeptide sequence. Computational tools like ExPasy-Prototam [<https://robeta.bakerlab.org/>] and RaptorX [<http://raptorx6.uchicago.edu/>] were used to identify the highly stable polypeptide sequences based on their instability index and helix percentage. After the prediction of a stable polypeptide sequence, a secondary structure was designed with the help of the Robetta server [<https://robeta.bakerlab.org/>] and Pymol software [<https://pymol.org/>] was used to visualize the predicted structure. In the Robetta server there were five results from which we have selected the less error rate structure. Refer to Figure- 5.

6. Prediction the 3D polypeptide structure designed

Designed structure was predicted using the Ramachandran plot [<http://vadar.wishartlab.com/>]. It shows the potential of the confirmation of the peptide structure. Accordingly, they plot the phi ϕ and the psi ψ angle in X and Y axis respectively. After designing the structure, we validate the sequence by plotting the Ramachandran Plot. They plot an angle spectrum ranging from -180° to $+180^\circ$. Refer to Figure- 5.

7. Molecular Dynamic Simulation

We have validated the results by using the online molecular dynamics simulator servers. For these Molecular dynamics we preferred to use the UAMS Simlab (WEBGRO Macromolecular Simulation). In this server there are two types of tools 'Protein in Water

Simulation' and 'Protein with Ligand Simulation'. They use the GROMACS simulation for predicting the solvated molecular dynamics simulations. Out of the above tools we preferred to use the 'Protein in Water Simulation' where we have just uploaded the predicted SOS1 and HKT1 structure one at a time. In input parameters, we have used the default parameters where 'Forcefield: GROMOS96 43a1'; 'Water model as SPC'; 'Box Type as triclinic'; 'Salt type as NaCl'. In Energy Minimization parameters, we have used the default parameters where 'Integrator as Steepest Descent'; 'Steps as 5000'. In Equilibration and MD run Parameters, we have used the default parameters where 'Equilibration type as NVT/NPT'; 'Temperature as 300K'; 'Pressure as 1.0bar'; 'MD integrator as Leap frog'; 'Simulation time as 50ns'; 'Approximate number of frames per simulation as 5000' and then submit to run the MD simulator. Refer to Figure- 5.

8. Predicting the antigenicity part in the sequence and mapping of the antigenicity on the sequences

Potential antigenicity was analyzed on the predicted sequences. It helps to predict computationally the B-cells and the T-cells in the sequence. Various computational tools were used such as ABCpred, Bcepred, BepiPred and SVMTriP.

ABCpred [<https://webs.iitd.edu.in/raghava/abcpred/>] is a computational tool used to predict the B-cell epitopes from the protein sequences. These epitopes are specifically part of the proteins which can bind to the antibodies. This tool uses the artificial neural network to predict the likely part of epitopes in the predicted protein sequence.

Like ABCpred, Bcepred [http://crdd.osdd.net/raghava/bcepred/bcepred_instructions.html] and BepiPred [<https://services.healthtech.dtu.dk/services/BepiPred-2.0/>] is also a computational tool used for prediction of B-cell epitopes. BepiPred uses the Hidden Markov Models (HMMs) and propensity scale methods to predict the B-cell epitopes from the protein sequences.

SVMTriP [<http://sysbio.unl.edu/SVMTriP/prediction.php>] is a computational tool used to predict the T-cell epitopes from the protein sequences. These epitopes are specifically a part of the proteins that are identified by T-cells of the immune system. This tool uses the Support Vector Machine algorithm to predict the T-cell epitopes.

After the prediction of the antigenicity by computational methods, the area of the antigenicity was marked in the predicted peptide structure. For marking the antigenicity, the Pymol

software [<https://pymol.org/>] was used. By marking it we can analyze the part which is more suitable for which epitopes. Refer to Figure- 5 depicts the workflow of the methodology used to design polypeptide marker for both SOS1 and HKT1.

Workflow for the designing of the Polypeptide markers

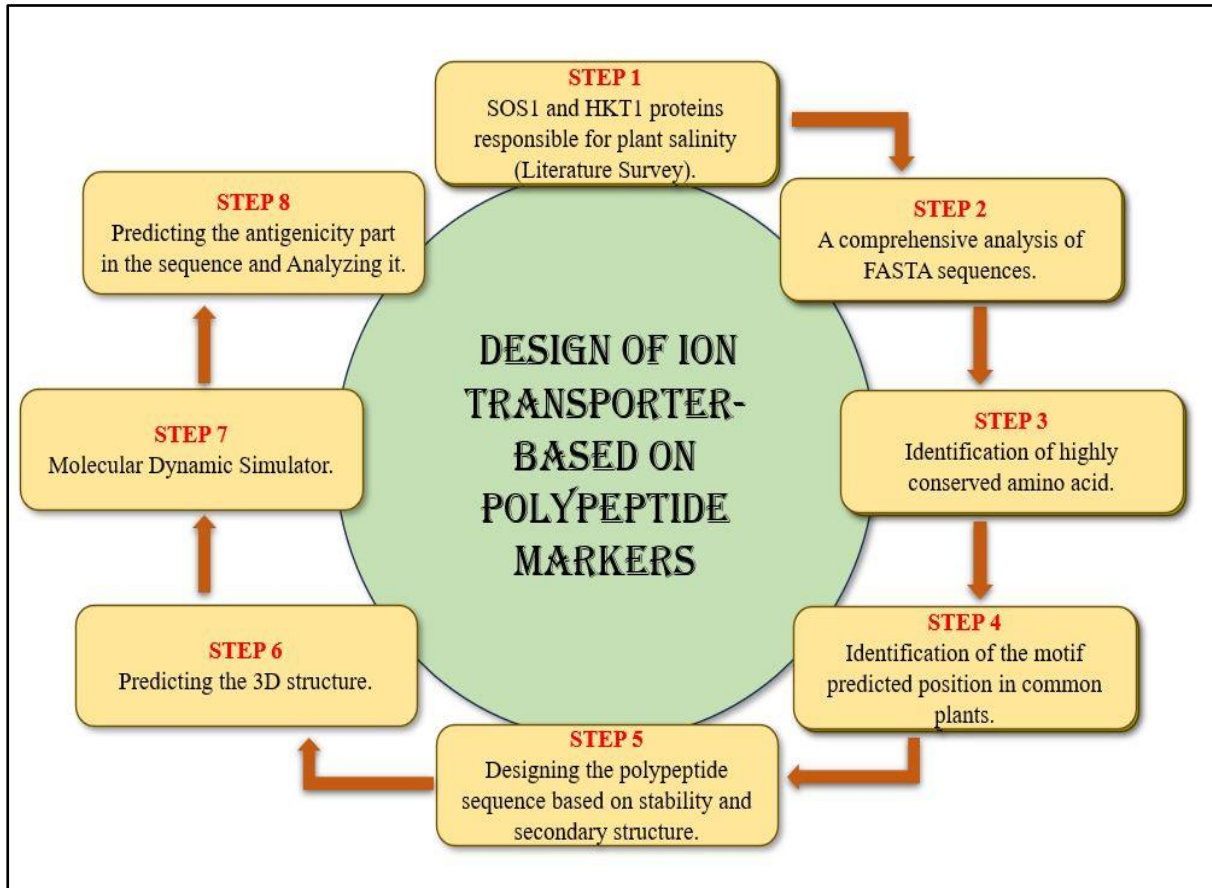


Figure 5: Flowchart to design an ion transporter based on polypeptide markers

Results

1. Analysis and comparison of the SOS1 and HKT1 proteins in crops

We have selected two proteins that are responsible for salinity tolerance SOS1 and HKT1. For both proteins, we have selected a common model plant *Arabidopsis thaliana*. The accession number of *Arabidopsis thaliana* SOS1 is NP_178307.2. The accession number of *Arabidopsis thaliana* HKT1 is OAO98616.1. The sequences of the above crops were used to identify other crops that are homologous to SOS1 and HKT1 proteins. Then, we go through the literature survey so that we can predict the crops with respect to the above proteins that have more tolerance to salinity. Accordingly, we have designed the table below which contains the organism's name, accession number and the homologous crops with respect to the *Arabidopsis thaliana* SOS1 (Table 3) and HKT1 (Table 4).

Table 3: SOS1 proteins from different crops with homology with *Arabidopsis thaliana* SOS1

Sl.No.	Protein Name	Scientific Name	Common Name	Accession Number	Query Cover (%)	Percentage Identity (%)
1.	AtSOS1	<i>Arabidopsis thaliana</i>	Thale cress	NP_178307.2	100	100
2.	TtSOS1	<i>Triticum turgidum</i>	Durum Wheat	ACB47885.1	96	60.18
3.	OsSOS1	<i>Oryza sativa</i>	Rice	ATU90113.1	97	60.31
4.	ZmSOS1	<i>Zea mays</i>	Maize	QOI16621.1	97	59.54
5.	GhSOS1	<i>Gossypium hirsutum</i>	Cotton	AKN19929.1	96	62.14
6.	GmSOS1	<i>Glycine max</i>	Soybean	NP_001244939.1	96	64.85
7.	NtSOS1	<i>Nicotiana tabacum</i>	Tobacco	XP_016466679.1	96	66.01
8.	SoSOS1	<i>Spinacia oleracea</i>	Spinach	CDL70805.1	92	63.44
9.	VvSOS1	<i>Vitis vinifera</i>	Grape	CBI26761.3	99	65.63
10.	SpSOS1	<i>Sesuvium portulacastrum</i>	Sea purslane	AFX68848.1	99	62.28
11.	StSOS1	<i>Solanum tuberosum</i>	Potato	XP_006364070.1	98	63.73
12.	KcSOS1	<i>Karelinia caspia</i>	Capsicum	QOS02253.1	97	64.56
13.	PeSOS1	<i>Populus euphratica</i>	Poplar	AQN76669.1	99	64.56

14.	<i>CqSOS1</i>	<i>Chenopodium quinoa</i>	Inca wheat	ABS72166.1	98	62.24
15.	<i>McSOS1</i>	<i>Mesembryanthemum crystallinum</i>	Baby sun rose	ABN04858.1	97	63.85
16.	<i>NtSOS1</i>	<i>Nitraria tangutorum</i>	Nitre bushes	AGW30210.1	99	63.87
17.	<i>LgSOS1</i>	<i>Limonium gmelinii</i>	Statice	ACF05808.1	99	60.19
18.	<i>SpSOS1</i>	<i>Schrenkiella parvula</i>	Salt cress	ADQ43186.1	99	78.71

Table 4: HKT1 proteins from different crops with homology with *Arabidopsis thaliana* HKT1

Sl.No	Protein Name	Scientific Name	Common Name	Accession Number	Query Cover (%)	Percentage Identity (%)
1.	<i>AtHKT1</i>	<i>Arabidopsis thaliana</i>	Thale cress	OAO98616.1	100	100
2.	<i>GmHKT1</i>	<i>Glycine max</i>	Soybean	XP_003517618.3	94	49.7
3.	<i>CcHKT1</i>	<i>Cajanus cajan</i>	Pegion pea	XP_020222038.2	94	51.76
4.	<i>CaHKT1</i>	<i>Cicer arietinum</i>	Chickpea	XP_004513109.1	94	48.5
5.	<i>AhHKT1</i>	<i>Arachis hypogaea</i>	Groundnut	XP_025697586.1	94	49.11
6.	<i>GaHKT1</i>	<i>Gossypium arboretum</i>	Cotton	XP_017644471.1	94	52.95
7.	<i>SiHKT1</i>	<i>Sesamum indicum</i>	Sesame	XP_011094146.1	94	52.52
8.	<i>SoHKT1</i>	<i>Spinacia oleracea</i>	Spinach	XP_021850306.1	94	48.21
9.	<i>NtHKT1</i>	<i>Nicotiana tabacum</i>	Tobacco	XP_016459984.1	94	51.96

10.	<i>S</i> HKT1	<i>Solanum lycopersicum</i>	Tomato	NP_001289833.1	94	49.8
11.	<i>St</i> HKT1	<i>Solanum tuberosum</i>	Potato	XP_006359731.1	94	49.7
12.	<i>Ha</i> HKT1	<i>Helianthus annuus</i>	Sunflower	XP_022009341.1	93	51.13
13.	<i>Hv</i> HKT1	<i>Hordeum vulgare</i>	Barley	KAE8777465.1	89	47.87
14.	<i>Rc</i> HKT1	<i>Ricinus communis</i>	Castor bean	XP_015576583.1	94	52.44
15.	<i>Ca</i> (pepper) <i>HKT1</i>	<i>Capsicum annuum</i>	Pepper	XP_016577037.2	94	49.8
16.	<i>Dc</i> HKT1	<i>Daucus carota</i>	Carrot	XP_017257369.1	94	48.13
17.	<i>Ec</i> HKT1	<i>Eucalyptus camaldulensis</i>	River red gum	AAF97728.1	94	51.29
18.	<i>Ss</i> HKT1	<i>Suaeda salsa</i>	Seepweed	AAS20529.2	93	48.01
19.	<i>Vv</i> HKT1	<i>Vitis vinifera</i>	Grape	XP_010656787.2	93	47.62
20.	<i>Br</i> HKT1	<i>Brassica rapa</i>	Turnip mustard	XP_009128719.1	99	77.17

2. Analysis of FASTA sequences

Multiple sequence alignments were done of the plants that were selected from the SOS1 and HKT1 sequences. It was done with the help of MAFFT, CLUSTALW, and MUSCLE tools respectively. The aligned sequences were interpreted with ESPript 3.0 tool as illustrated below for SOS1 refer to Figure 6 and for HKT1 refer to Figure 7.



Figure 6: Multiple Sequence Alignment for SOS1 Protein

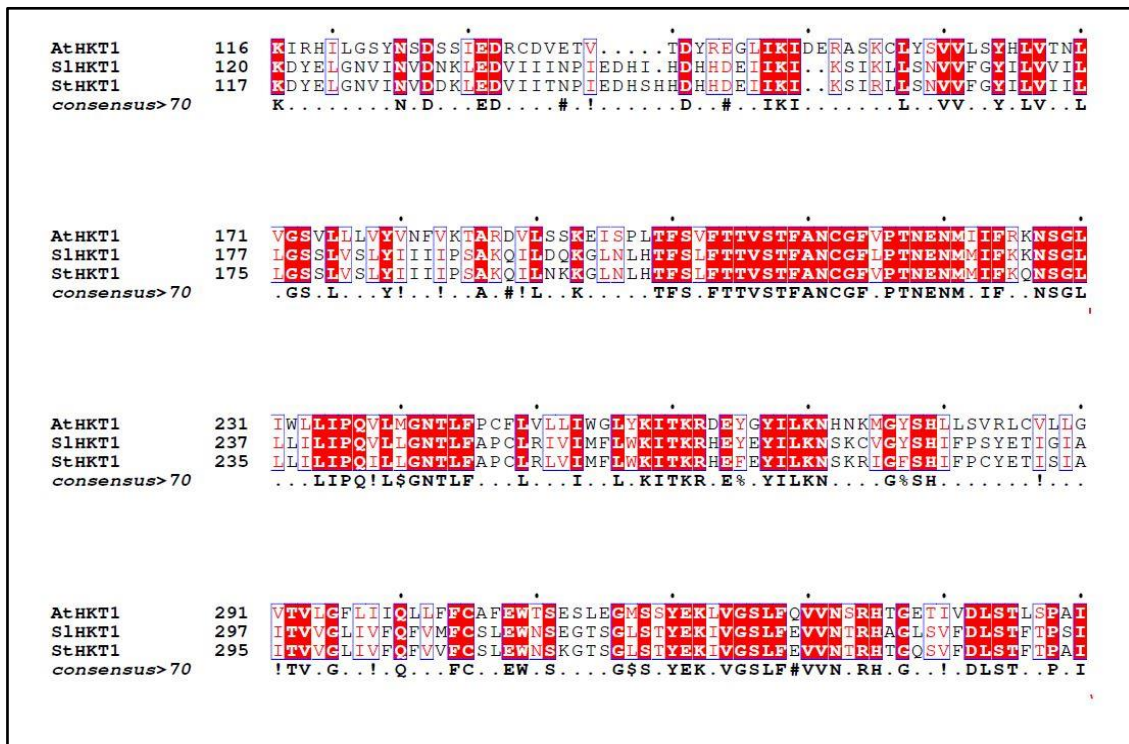


Figure 7: Multiple Sequence Alignment for HKT1 Protein

3. Identification of highly conserved amino acid

After running multiple sequence alignments using MAFFT software, we have identified the conserved motif for SOS1 and HKT1 sequences from the alignment using the MEME server. It designated five motifs as Motif 1, Motif 2, Motif 3, Motif 4, Motif 5. The motifs of the conserved amino acid were identified on the basis of the E-value. For SOS1 refer to Figure 8 and for HKT1 refer to Figure 9.



Figure 8: Conserved motif for SOS1

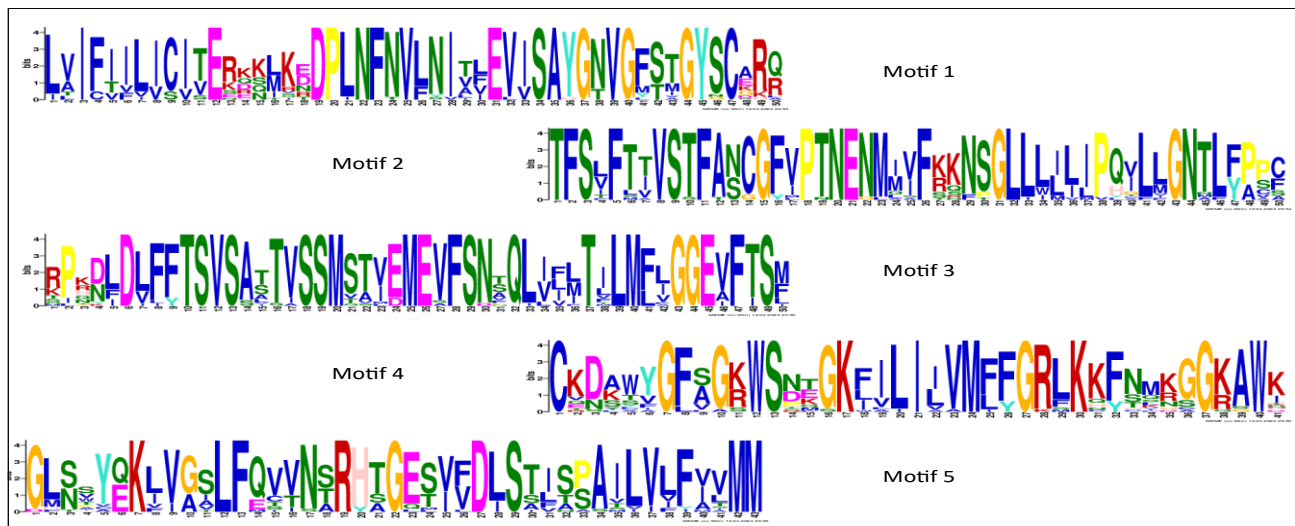


Figure 9: Conserved motif for HKT1

4. Identification of the motif predicted position in common plants

Multiple sequence alignments were done on the plants that were selected from the SOS1 and HKT1 sequences with selected stable sequences (Motif). It was done with the help of MAFFT. The aligned sequences were interpreted in ESPrnt 3.0 tools. For SOS1 refer to Figure 10 and for HKT1 refer to Figure 11.

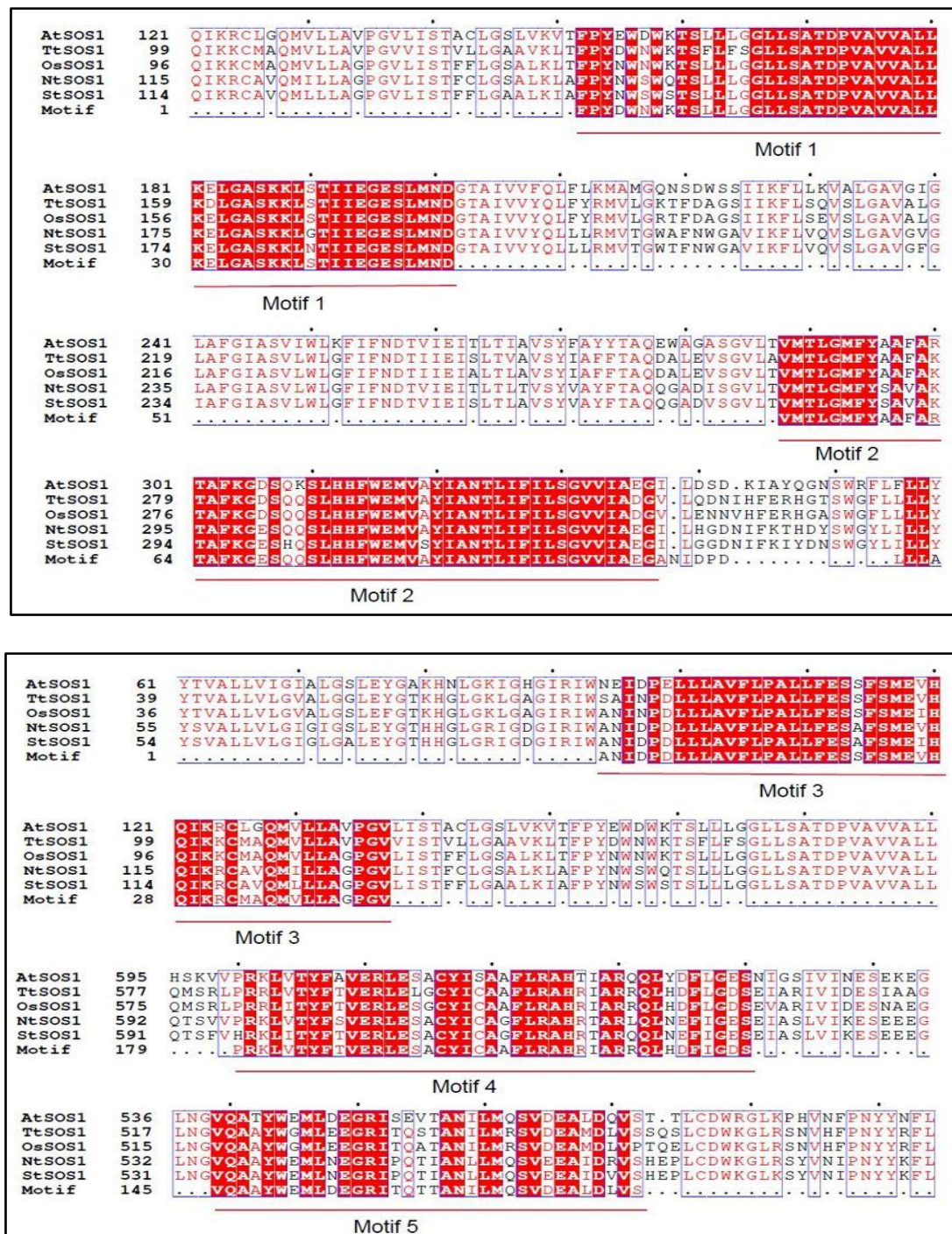


Figure 10: Alignment with Selected sequence SOS1

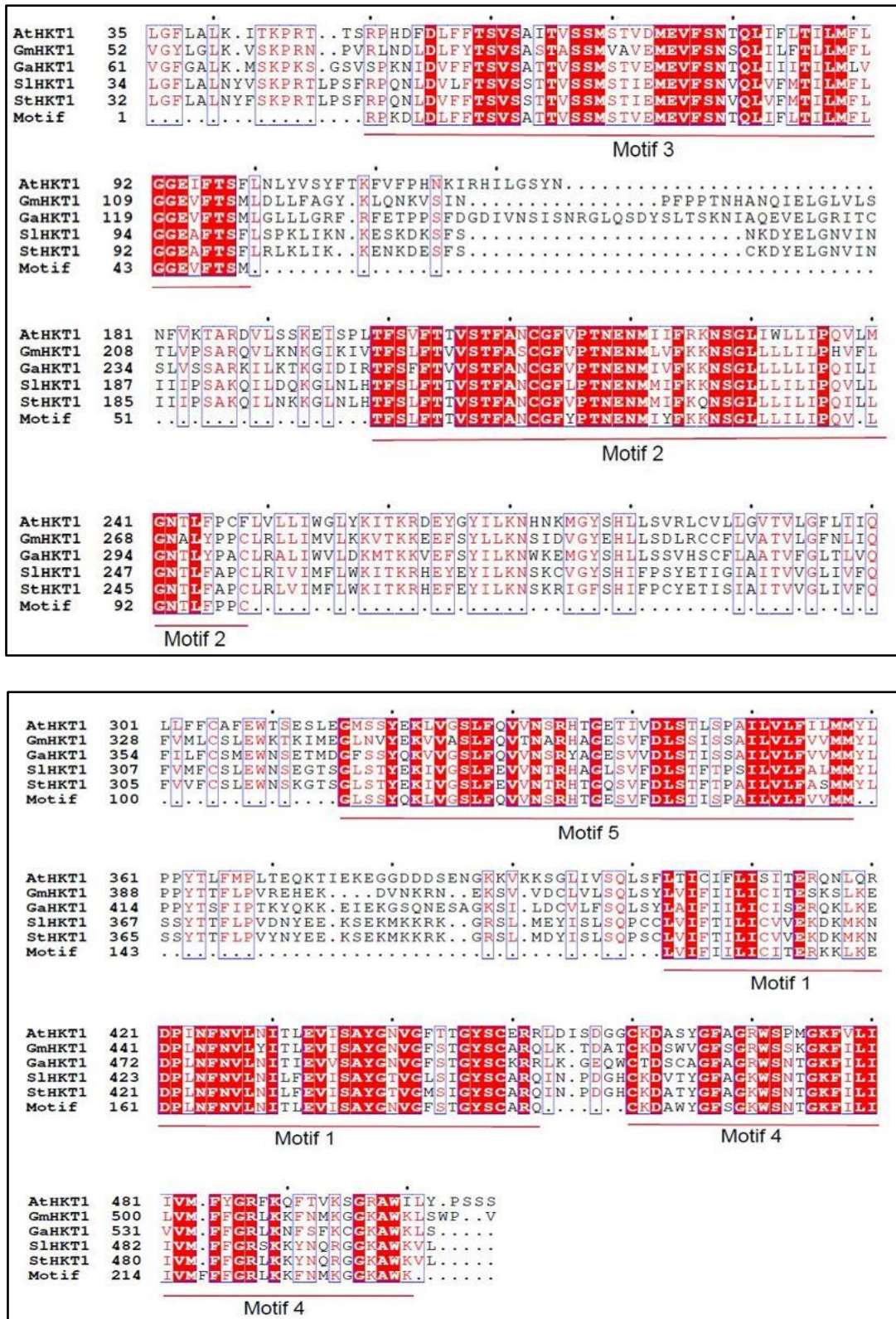


Figure 11: Alignment with Selected sequence HKT1

5. Designing of the polypeptide sequence on the basis of the instability index and the helicity and secondary structure plotted.

For designing the SOS1 and HKT1 sequences, from the motifs the conserved amino acids were identified. After identification, the amino acid was arranged to form one sequence for SOS1 and one sequence for HKT1. The various combination of sequence was arranged and stability was checked on the basis of the instability index and the helicity. By instability index, we can predict the stability of the peptide sequence. It is calculated on the basis of the peptide composition. During prediction if the instability index is less 40 then we can estimate that the peptide sequence is stable or if the instability index is greater than 40 then we can estimate that the sequence form is unstable. Around 120 combinations of sequences were verified from the above conserved amino acid sequences. On successful verification we were able to establish the 5 stable conserved amino acid sequences for SOS1 (Table 5) and HKT1 (Table 6). Later, the secondary structure for SOS1 and HKT1 sequences was plotted on the Robetta server, which had the more stability [Refer to ANNEXURE-I SOS1 (Table 10), (Figure 27); HKT1 (Table 11), (Figure 27)]. In this server, 5 models were formed from the above most stable sequence on the basis of the energy, stability and the error rate. Out of which we have selected the more stable and less error rate-based models.

Table 5: Instability index and Helix Percentage of SOS1.

SL.No.	Motif's Sequence's	Instability Index	Helix Percentage (%)
1	A1, A2, A3, A5, A4	35.35	80
2	A1, A4, A5, A3, A2	37.09	77
3	A2, A3, A4, A5, A1	37.58	77
4	A4, A1, A2, A3, A5	37.79	75
5	A4, A2, A3, A1, A5	37.79	75

#Here **A1**= FPYDWNWKTSLLLGGLLSATDPVAVVALLKELGASKKLSTIIE GESLMND;
A2= VMTLGMFYAAFARTAFKGESQQSLHHFWEMVAYIANTLIF ILSGVVIAEG;
A3= ANIDPDLLLAVFLPALLFESSFSMEVHQIKRCMAQMV LLA GPGV;
A4= PRKLVTYFTVERLESACYICAAFLRAHRIARRQLHDFIGDS;
A5= VQAAYWEMLDEG RITQTTANILMQSVDEALDLVS.

Table 6: Instability index and Helix Percentage of HKT 1

SL.No.	Motif's Sequence's	Instability Index	Helix Percentage (%)
1	A1, A4, A5, A3, A2	22.69	57
2	A1, A4, A5, A2, A3	23.01	61
3	A4, A1, A5, A3, A2	23.01	57
4	A3, A1, A4, A5, A2	23.01	60
5	A5, A3, A2, A4, A1	23.01	62

#Here **A1**= LVIFIILCITERKKLKEDPLNFNVLNITLEVISAYGNVGFSTGYSCA RQ;

A2= TFSLFTTVSTFANCGFYPTNENMIYFKKNSGL LLILIPQVLGNTLFPP C;

A3= RPKDLDLFFTSVSATTVSSMSTVEMEVFSNTQLIFLITLMFLGGEVFTS M;

A4= CKDAWYGFSGKWSNTGKFILIVMFF FGRLKKFNMKGGKAWK;

A5= GLSS YQKLVGSLFQVVNSRHTGES VFDLSTISPAILVLFVMM

6. Prediction of the 3D polypeptide structure designed

After the 3D models were created from the above selected conserved amino acid the potential of the confirmation of the peptide structure was done with the help of the Vadar server. This server mainly confirms the structure potential by plotting the “Ramachandran Plot” (Figure-12) on which it plots the phi (ϕ) and the psi (ψ) angle in X and Y axis respectively. On the basis of that residue in phi (ϕ) psi (ψ) core, residue in phi (ϕ) psi (ψ) allowed, residue in phi (ϕ) psi (ψ) generous and residue in phi (ϕ) psi (ψ) outside were also observed for both SOS1 (Table 7) and HKT1 (Table 8) protein.

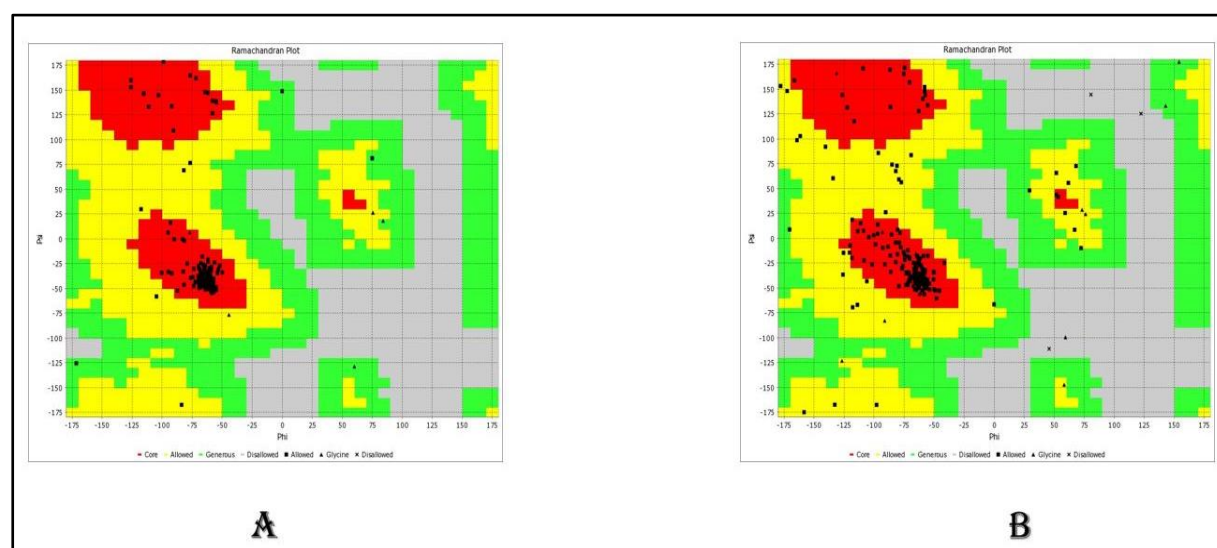


Figure 12: Ramachandran plot of secondary Structure; **A.** SOS1; **B.** HKT1

Table 7: Different Ramachandran plot statistics for residue in phi-psi in SOS1 structure

Sl.No.	Ramachandran plot statistics	Observed	Expected
1	Residue in phi (ϕ) psi (ψ) core	212 (96%)	197 (90%)
2	Residue in phi (ϕ) psi (ψ) allowed	5 (2%)	15 (7%)
3	Residue in phi (ϕ) psi (ψ) generous	2 (0%)	2 (1%)
4	Residue in phi (ϕ) psi (ψ) outside	0 (0%)	0 (0%)

Table 8: Different Ramachandran plot statistics for residue in phi-psi in HKT1 structure

Sl.No.	Ramachandran plot statistics	Observed	Expected
1	Residue in phi (ϕ) psi (ψ) core	211 (90%)	211(90%)
2	Residue in phi (ϕ) psi (ψ) allowed	16(6%)	16(7%)
3	Residue in phi (ϕ) psi (ψ) generous	4(1%)	2(1%)
4	Residue in phi (ϕ) psi (ψ) outside	3(1%)	0(0%)

7. Molecular Dynamic Simulation

After predicting the SOS1 and HKT1 membrane protein structure and then predicting the 3D structure of the protein using the Ramachandran plot, we were finally able to establish a suitable structure. For further validation, we have done the molecular dynamics simulation in which we focus on the different parameters of a graph such as radius of gyration vs time, solvent accessible vs time, RMSD value vs time and RMS fluctuation vs residues. For SOS1 refer to figure 13,14; [ANNEXURE-I (Figure 28)] and for HKT1 refer to figure 15,16); [ANNEXURE-I (Figure 29)].

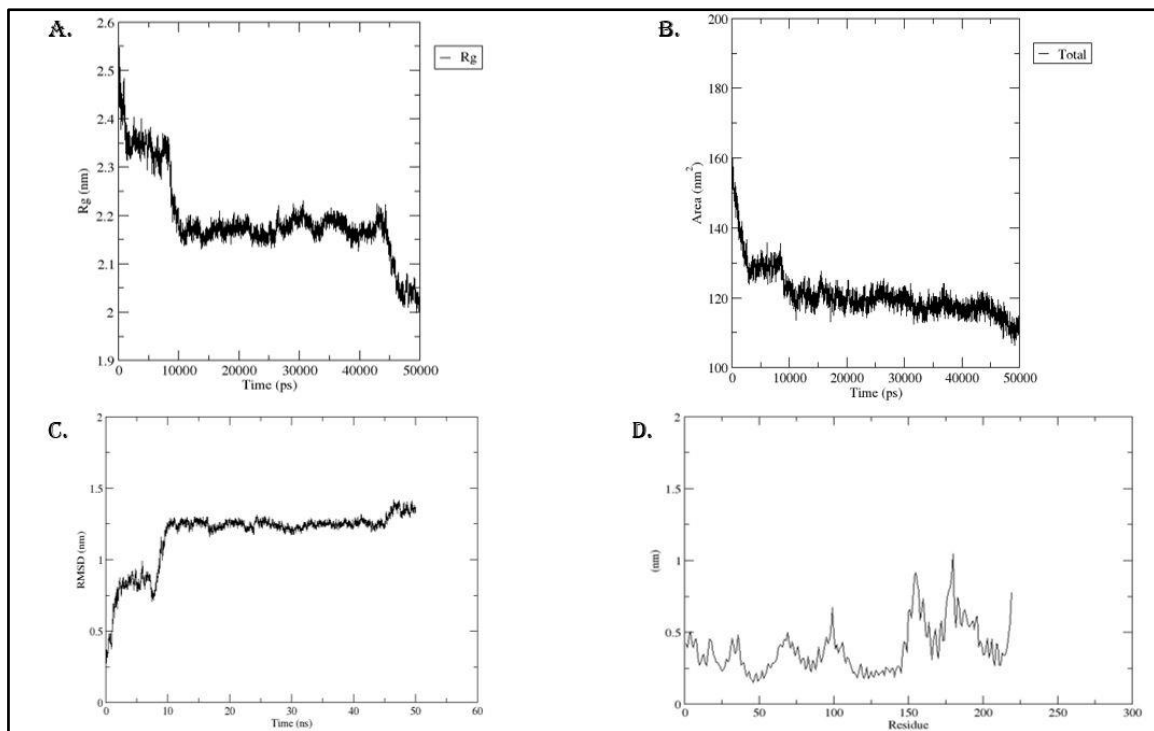


Figure 13: Molecular dynamic simulation of SOS1 sequence **A.** Rg vs Time [Radius of gyration]; **B.** Area vs Time [Solvent Accessible surface]; **C.** RMSD vs Time [RMSD]; **D.** RMS fluctuation.

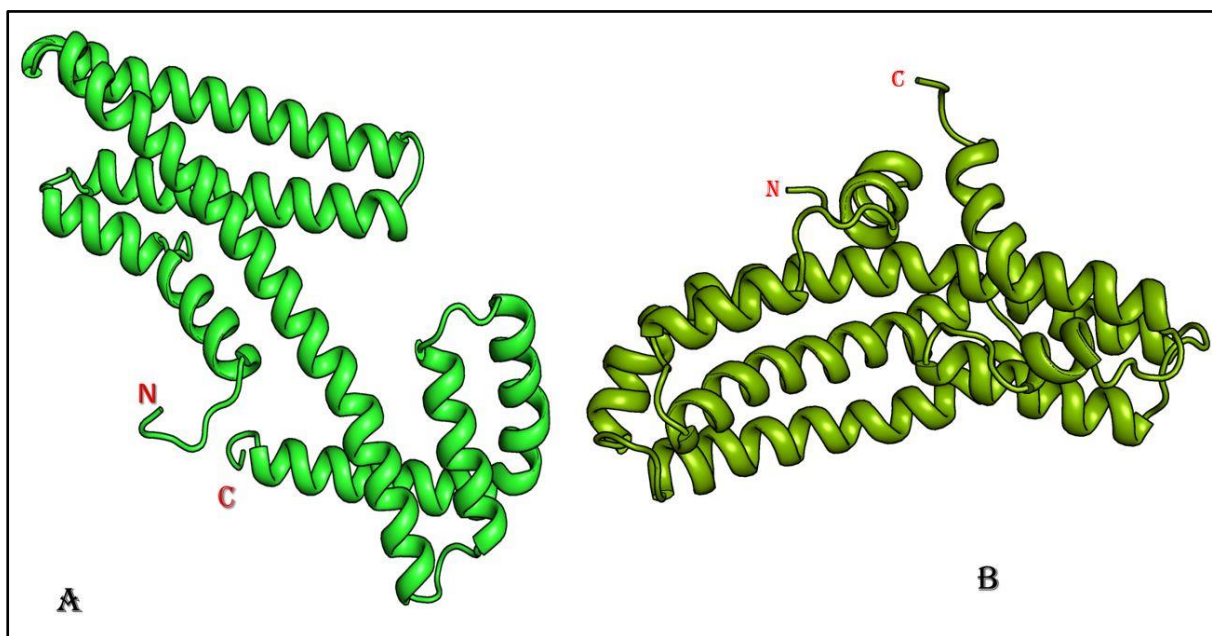


Figure 14: Comparison of SOS1 peptide structure with **A.** Before Simulation; **B.** After Simulation

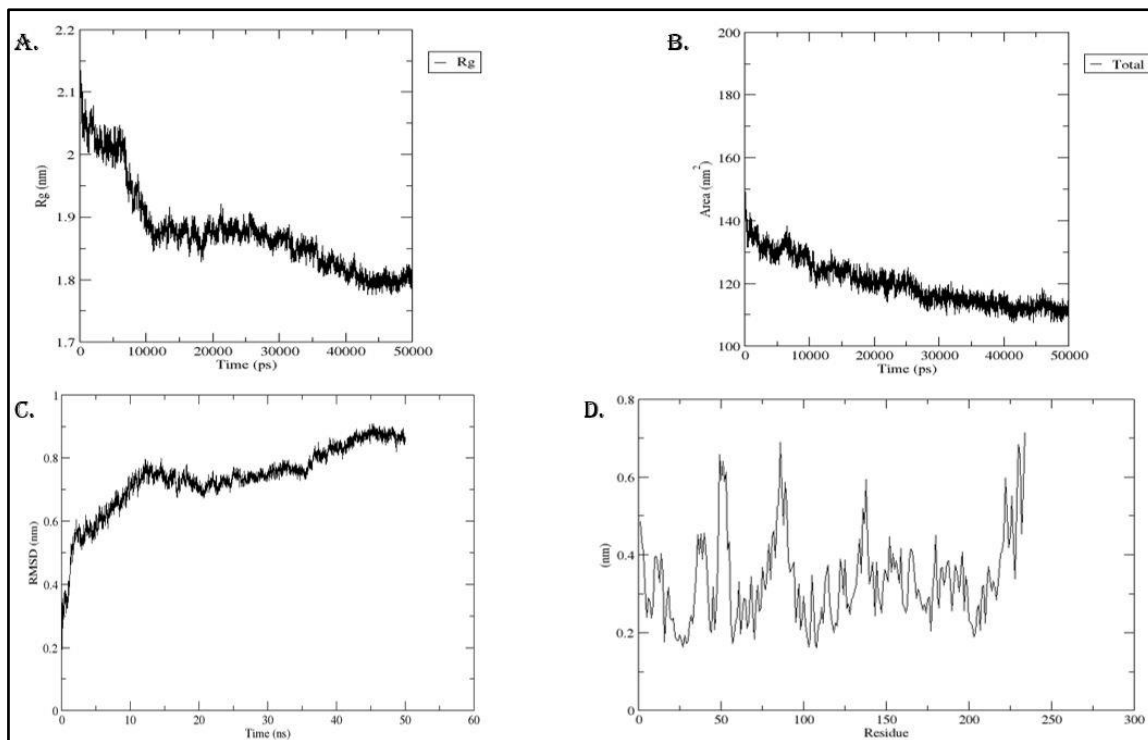


Figure 15: Molecular dynamic simulation of HKT1 sequence **A.** Rg vs Time [Radius of gyration]; **B.** Area vs Time [Solvent Accessible surface]; **C.** RMSD vs Time [RMSD]; **D.** RMS fluctuation.

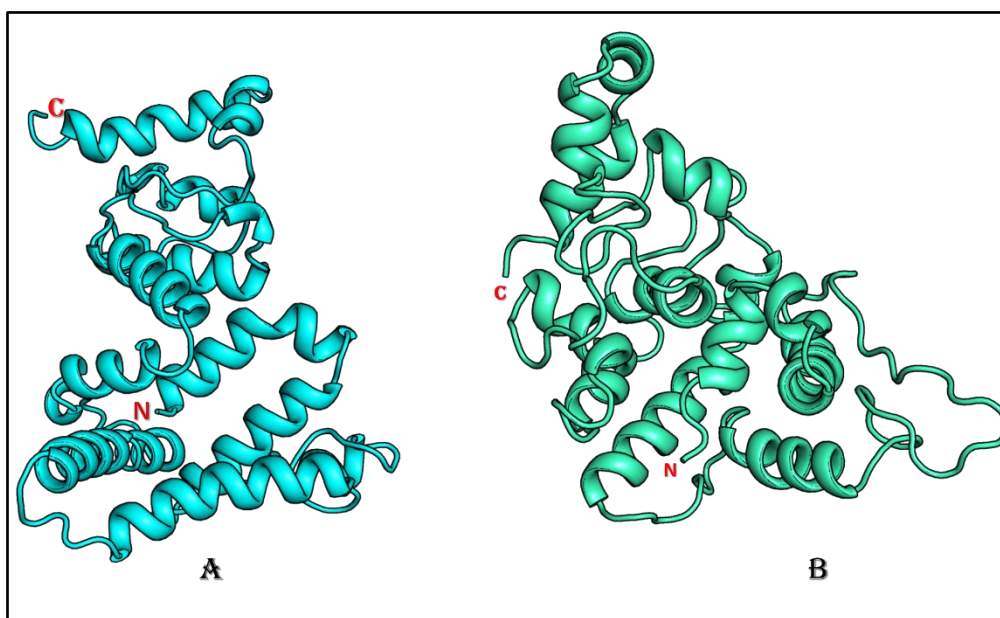


Figure 16: Comparison of HKT1 peptide structure with **A.** Before Simulation; **B.** After Simulation

8. Predicting the antigenicity part in the sequence and mapping of the antigenicity on the sequences

Predicted sequences of SOS1 and HKT1 proteins antigenicity were analysed using the servers ABCpred, Bcepred, BepiPred and SVMTriP. Different servers were used to improve the accuracy of the antigenic parameter. On this server, we have analyzed the results on the basis of the threshold value 0.5 to predict the more proper binders. It was also observed in Bepi pred server that in SOS1 and HKT1 predicted sequences mostly had overlapping which specifies its potential antigenicity.

For SOS1, we have selected the part of the sequence on the basis of the score such as in SVMTriP (Table 9) (Figure 17,18); [ANNEXURE-I (Table 12); (Figure 29)]; we have preferred the threshold value of 1.0, in ABCpred (Figure 17,18); [ANNEXURE-I (Table 14); (Figure 30)]; we prefer the highest threshold value and in BepiPred we prefer the threshold value greater than 0.5 (Table 9); (Figure17,18), [ANNEXURE-I (Figure 31)].

Similarly, for HKT1, we have selected the part of the sequence on the basis of the score such as in SVMTriP (Table 9) (Figure 17,18); [ANNEXURE-I (Table 13); (Figure 29)], we have preferred the threshold value of 1.0, in ABCpred (Table 9) (Figure 17,18); [ANNEXURE-I (Table 15); (Figure 30)] we prefer the highest threshold value and in BepiPred we prefer the threshold value greater than 0.5 (Table 9); (Figure17,18), [ANNEXURE-I (Figure 31)].

We have also used the BcePred in which we have predicted the B-Cell epitope on the basis of physiochemical properties such as hydrophilicity, flexibility/mobility, accessibility, polarity, exposed surface and turns. The tables are attached in ANNEXURE-I (Table 16,17).

After plotting of the secondary structure and prediction of 3D structure in the Ramachandran plot, we started to map the antigenicity part of the structure to find the most capable binding site. This part was analysed using a different server such as SVMTriP. ABCpred and BepiPred.

Table 9: Selected sequence of SOS1 and HKT1 from server on the basis of score

Sl.No.	Predicting Server	Protein Name	Selected sequence	Score
1	SVMTrip	SOS1	DWNNWKTSLLLGGLLSATDPV	1.0
		HKT1	VFDLSTISPAILVLFVVMR	1.0
2	ABCpred	SOS1	AAYWEMLDEGRITQTT	0.84
		HKT1	TVSSMSTVEMEVFSNT	0.91
3	BepiPred	SOS1	WNW/ GASKK/ AFKGESQQLHFWEMVA/ ANIDP/ AYWEMLDEGR/ EALDLVSP/ HDF	Greater than 0.5
		HKT1	KLKEDPLN/ SCARQCKDA/ LKKFNMKGGKAWKGLSSYQK/ VNSRHTGESVFD/ TTVSSMSTVEMEVFSN/ N- GFYPTNENMIYFKK/ NTL	Greater than 0.5

SVMTrip [SOS1]

FPYDWNNWKTSLLLGGLLSATDPVAVVALLKELGASKKLSIIIEGESLMNDVMTLGMFYAAFARTAFKGESQQLH
HFWEMVAYIANTLIFILSGVVIAEGANIDPDLLAVFLPALLFESSFSMEVHQIKRCMAQMVLVLAGPGVVQAAAYWE
MLDEGRITQTTANILMQSVDEALDLVSPRKLVTYFTVERLESACYICAAFLRAHRIARRQLHDFIGDS

SVMTrip [HKT1]

LVIFILICITERKLLKEDPLNFNVLNITLEVISAYGNVGFSTGYSCARQCKDAWYGFSGKWSNTGKFILIVMFFFGR
LKKFNMKGGKAWKGLSSYQKLVGSLFQVVNSRHTGESVFDLSTISPAILVLFVVMRPKDLDLFFTSVSATTVSS
MSTVEMEVFSNTQLIFLITLMFLGGEVFTSMTFSLFTTVSTFANCGFYPTNENMIYFKKNSGLLLILIPQVLGNTLFP
PC

ABCpred [SOS1]

FPYDWNNWKTSLLLGGLLSATDPVAVVALLKELGASKKLSIIIEGESLMNDVMTLGMFYAAFARTAFKGESQQLH
HFWEMVAYIANTLIFILSGVVIAEGANIDPDLLAVFLPALLFESSFSMEVHQIKRCMAQMVLVLAGPGVVQAAAYWE
MLDEGRITQTTANILMQSVDEALDLVSPRKLVTYFTVERLESACYICAAFLRAHRIARRQLHDFIGDS

ABCpred [HKT1]

LVIFILICITERKLLKEDPLNFNVLNITLEVISAYGNVGFSTGYSCARQCKDAWYGFSGKWSNTGKFILIVMFFFGR
LKKFNMKGGKAWKGLSSYQKLVGSLFQVVNSRHTGESVFDLSTISPAILVLFVVMRPKDLDLFFTSVSATTVSS
MSTVEMEVFSNTQLIFLITLMFLGGEVFTSMTFSLFTTVSTFANCGFYPTNENMIYFKKNSGLLLILIPQVLGNTLFP
PC

BepiPred [SOS1]

FPYDWNNWKTSLLLGGLLSATDPVAVVALLKELGASKKLSIIIEGESLMNDVMTLGMFYAAFARTAFKGESQQLH
HFWEMVAYIANTLIFILSGVVIAEGANIDPDLLAVFLPALLFESSFSMEVHQIKRCMAQMVLVLAGPGVVQAAAYWE
MLDEGRITQTTANILMQSVDEALDLVSPRKLVTYFTVERLESACYICAAFLRAHRIARRQLHDFIGDS

BepiPred [HKT1]

LVIFILICITERKLLKEDPLNFNVLNITLEVISAYGNVGFSTGYSCARQCKDAWYGFSGKWSNTGKFILIVMFFFGR
LKKFNMKGGKAWKGLSSYQKLVGSLFQVVNSRHTGESVFDLSTISPAILVLFVVMRPKDLDLFFTSVSATTVSS
MSTVEMEVFSNTQLIFLITLMFLGGEVFTSMTFSLFTTVSTFANCGFYPTNENMIYFKKNSGLLLILIPQVLGNTLFP
PC

Figure 17: Selected Sequence of SOS1 and HKT1 from different server

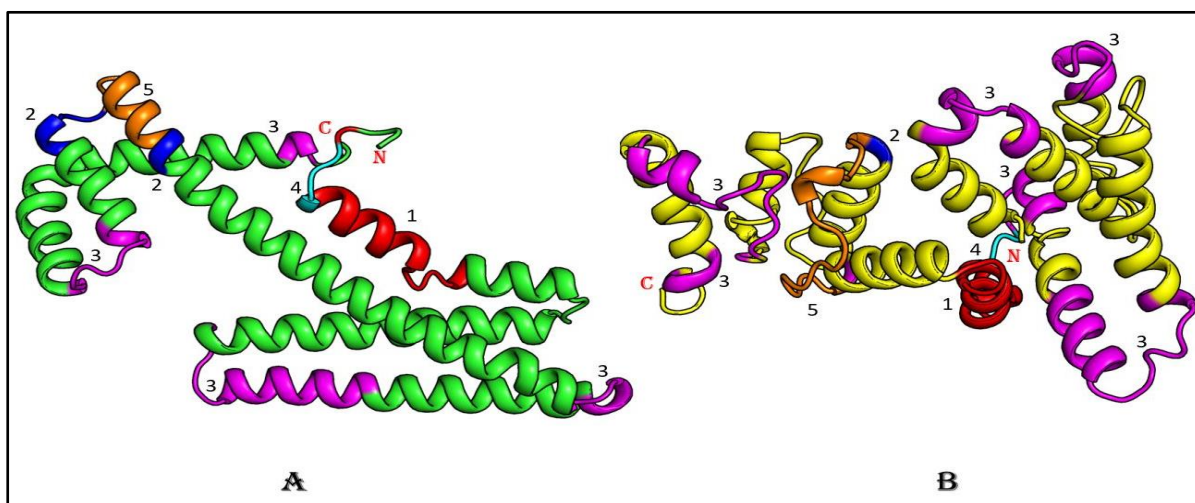


Figure 18: Mapping of antigenicity by different servers in designed structure **A.** SOS1; **B.** HKT1. Both structures were mapped and labeled as 1. SVMTrip [Red]; 2. ABCpred [Blue]; 3. BepiPred [Magenta]; 4. SVMTrip + BepiPred [Cyan]; 5. ABCpred + BepiPred [Orange]

Discussion

SOS1 is one of the major membrane proteins associated with the salinity tolerance. SOS1 protein are uses electrochemical gradients to extract the sodium ions from the internal of plant cell and extruding it, thereby maintaining low concentration of sodium in the cytoplasm of the cells. HKT1 is also a membrane protein associated with the salinity tolerance. It functions as a uniporter or symporter for the transport of potassium ions across the plasma membrane.

We have selected SOS1 and HKT1 membrane protein sequences of different plants respectively that were used for multiple sequence alignment and predicted the conserved part from the aligned sequences. The conserved motif 1, motif 2, motif 3, motif 4 and motif 5 were combined to form 120 amino acid sequences. We have also predicted the conserved motif parts position in some common plants such as in SOS1 compared plants were taken as *AtSOS1*, *TiSOS1*, *OsSOS1*, *NtSOS1*, *StSOS1* and for HKT1 compared plants were taken as *AtHKT1*, *GmHKT1*, *GaHKT1*, *SlHKT1*, *StHKT1*. This prediction was done to identify the region of conserved motif in plants.

Later, from 120 combination we have selected the most stable sequence as per the instability index and helix percentage. This prediction shows that for SOS1 sequence predicted combination were A1, A2, A3, A5, A4 whose instability index was 35.35 and helix percentage was 80%; for HKT1 sequence predicted combination were A1, A4, A5, A3, A2 whose instability index was 22.69 and helix percentage was 57%. After getting the stable sequences for SOS1 and HKT1 membrane protein, we have plotted a de novo model using the above selected sequences for SOS1 and HKT1. During modelling we get 5 models for SOS1 sequence and HKT1 sequence on the basis of the energy, stability and the error rate. Out of which we have selected the more stable and less error rate-based models.

After predicting the stable sequences and models for SOS1 and HKT1 protein, we predicted the structure using Ramachandran Plot. On the basis of the phi ϕ and the psi ψ angle in X and Y axis respectively and different statistics for residue in phi (ϕ) psi (ψ) core, residue in phi (ϕ) psi (ψ) allowed, residue in phi (ϕ) psi (ψ) generous and residue in phi (ϕ) psi (ψ) outside were enabled us to confirming the 3D structure for both SOS1 and HKT1 protein. This shows that we have successfully able to design the polypeptide sequences for SOS1 with length of 219 amino acid and HKT1 with length of 234 amino acid.

For further validation we have also run the molecular dynamic simulation of SOS1 and HKT1 stable structure. In this simulator, we focus on the different parameters such as radius of gyration, solvent accessibility, RMSD value and RMS fluctuation. In the designed SOS1 structure, from a radius of gyration we were able to predict from the graph that the molecular structure formed was compact; from RMSD we can predict that the structure formed is in an equilibrium state with little fluctuation by which we can presume the deviation of the atomic position with time; from RMSF graph we can predict some part of the sequence is flexible and some parts are rigid indicates about the fluctuation of each atom in its average positions. In designed HKT1 structure, from radius of gyration, we were able to predict from the graph that some part in the starting of the molecular structure is in a compact state while the rest of the parts is less compact or expansion; from RMSD graph, we can predict that most of the structure is in equilibrium state by which we can state the atomic position is stable overtime; from RMSF graph we can predict some part of the sequence is flexible and some parts are rigid indicates about the fluctuation of each atom in its average positions. They have also replotted the structure of the peptide designed where, we found that the after-simulation peptide structure formed is somehow compact structure. By this, we can assume that structure formed after simulation is more stable on the basis of the reducing the likelihood of degradation. This simulator helps to predict the different properties of the above two designed protein sequences on the basis of parameters.

For further analysis with respect to future aspects, we have gone for predicting the antigenicity part of the above designed SOS1 and HKT1 sequence. From this prediction we were able to analyse the binding site of the antibody in the above polypeptide designed sequences. It was observed that most of the antigenic sites in SOS1 and HKT1 sequence are having overlapping sites respectively. This analysis was done to predict the accurate polypeptide binding sites, which might bind to the predicted region when antibody is produced in rabbits or mice or rat.

Conclusion

We have selected SOS1 and HKT1 membrane protein sequences of different plants that show some functions in salinity stress. From the selected sequences we have done the multiple sequence alignment and predicted the five conserved motifs. We have also gone for multiple sequence alignment with some common plants to identify the region of conserved motif in such plants. Then the predicted conserved motifs were combined to form the one hundred twenty sequences. Out of those sequences we predicted the most stable sequences and plotted a de novo model and selected the most accurate model on the basis of the less error rate. After predicting the model and to validate it, we use Ramachandran Plot. On the basis of the phi ϕ and the psi ψ angle in X and Y axis respectively it established that we have successfully able to design the polypeptide sequences for SOS1 with length of 219 amino acid and HKT1 with length of 234 amino acid.

Later, we have also validated the above designed polypeptide structure using the molecular dynamic simulation on the basis of different parameters. Structure formation after simulation, we can assume that the structure is more stable. For future prospects, we have gone for predicting the antigenicity part of the above designed SOS1 and HKT1 sequence.

After various prediction, designing, interpreting and finally validating by various methods we can expect that the designer of ion transporter-based polypeptide markers can be applied in multiple plants for detection of salinity stress.

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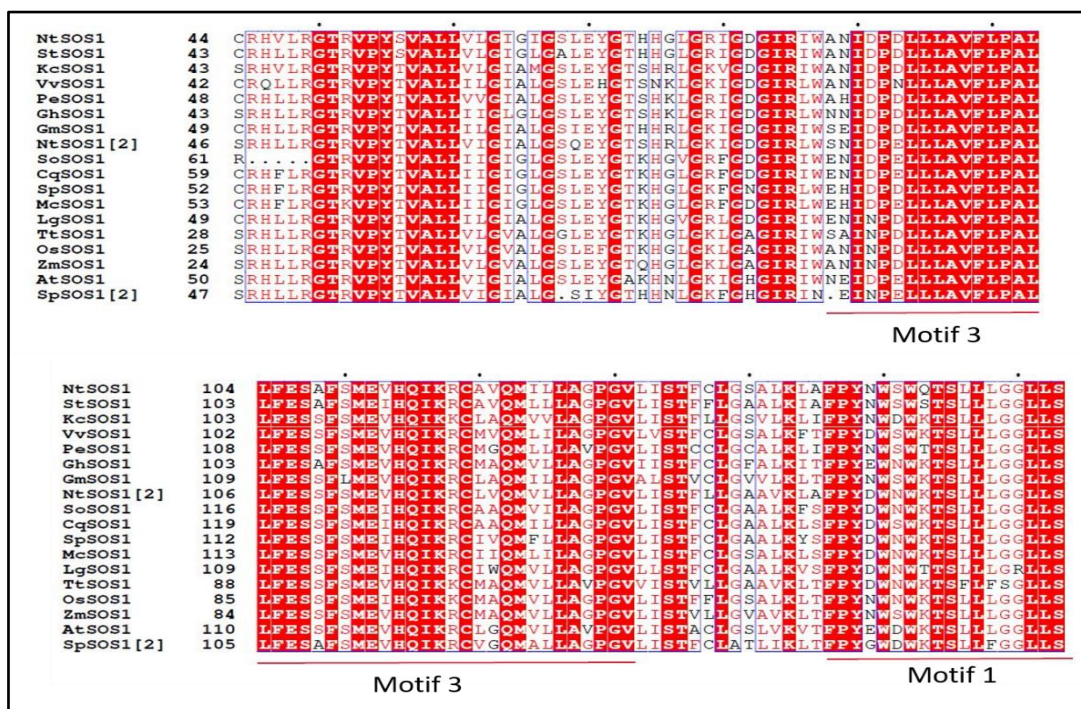
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ANNEXURE-I



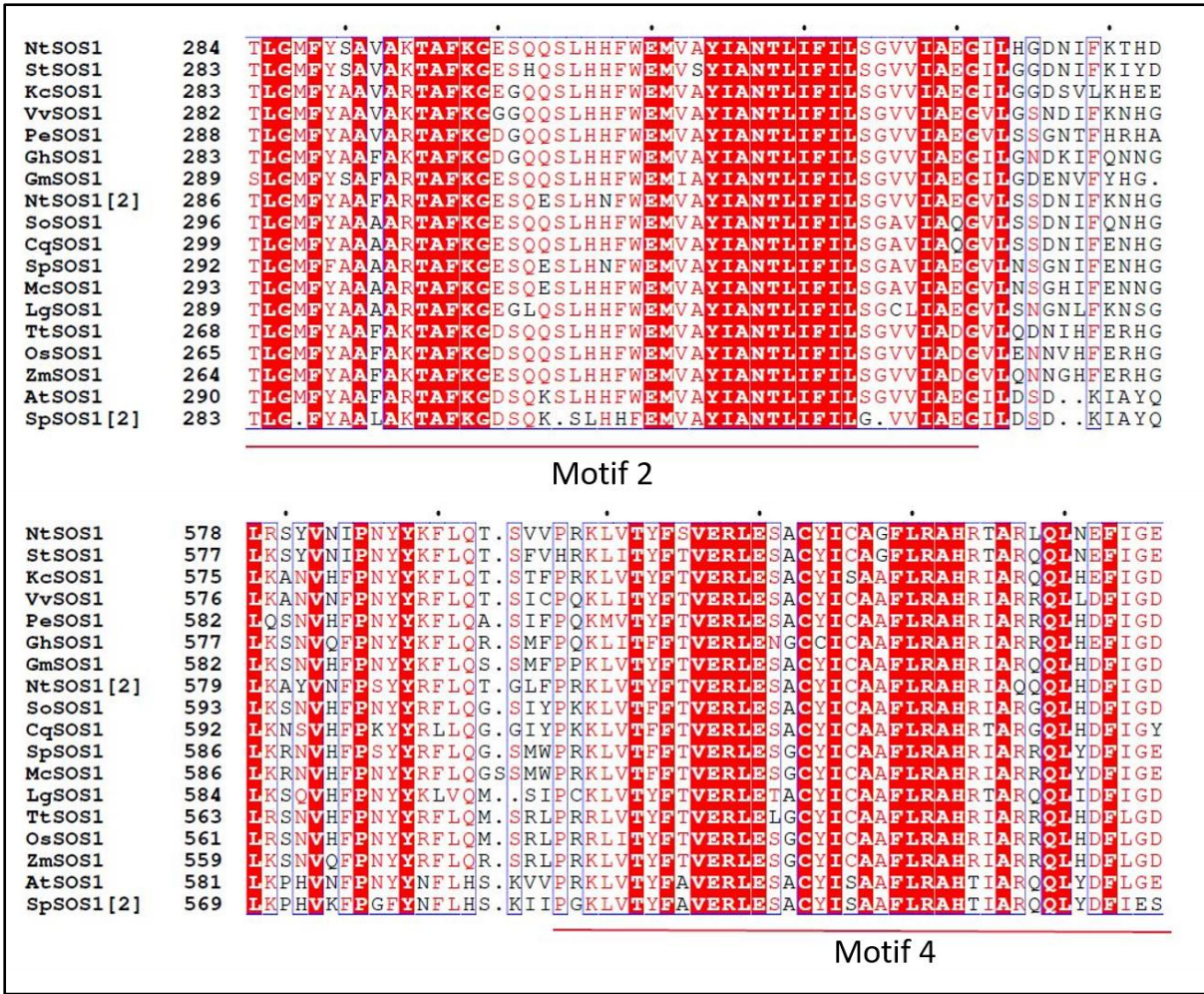


Figure 21: Highly conserved amino acid motif after multiple sequence alignment of SOS1 with motif 2 and motif 4

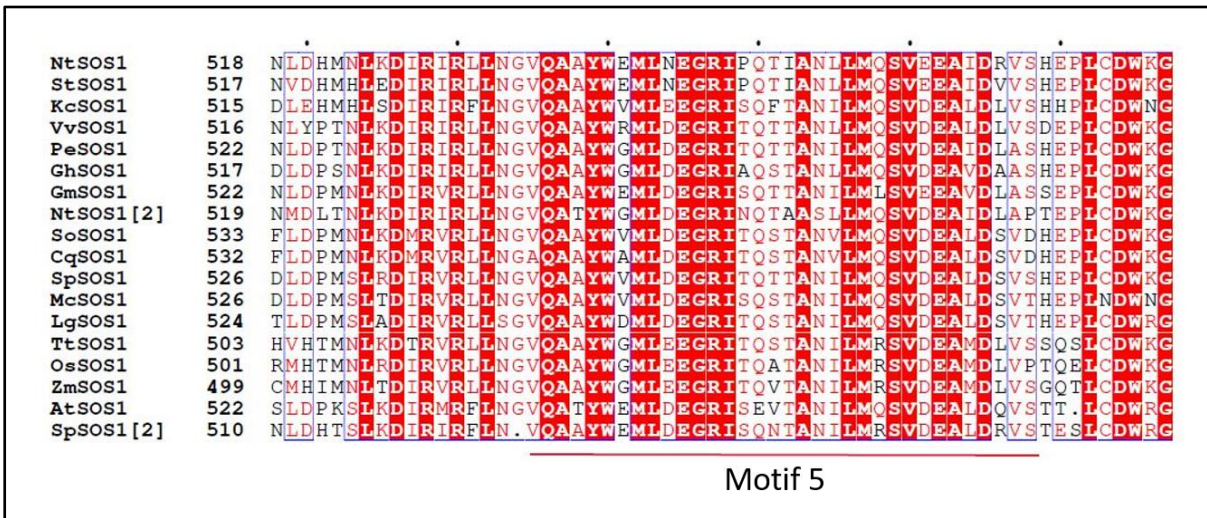


Figure 22: Highly conserved amino acid motif after multiple sequence alignment of SOS1 with motif 5

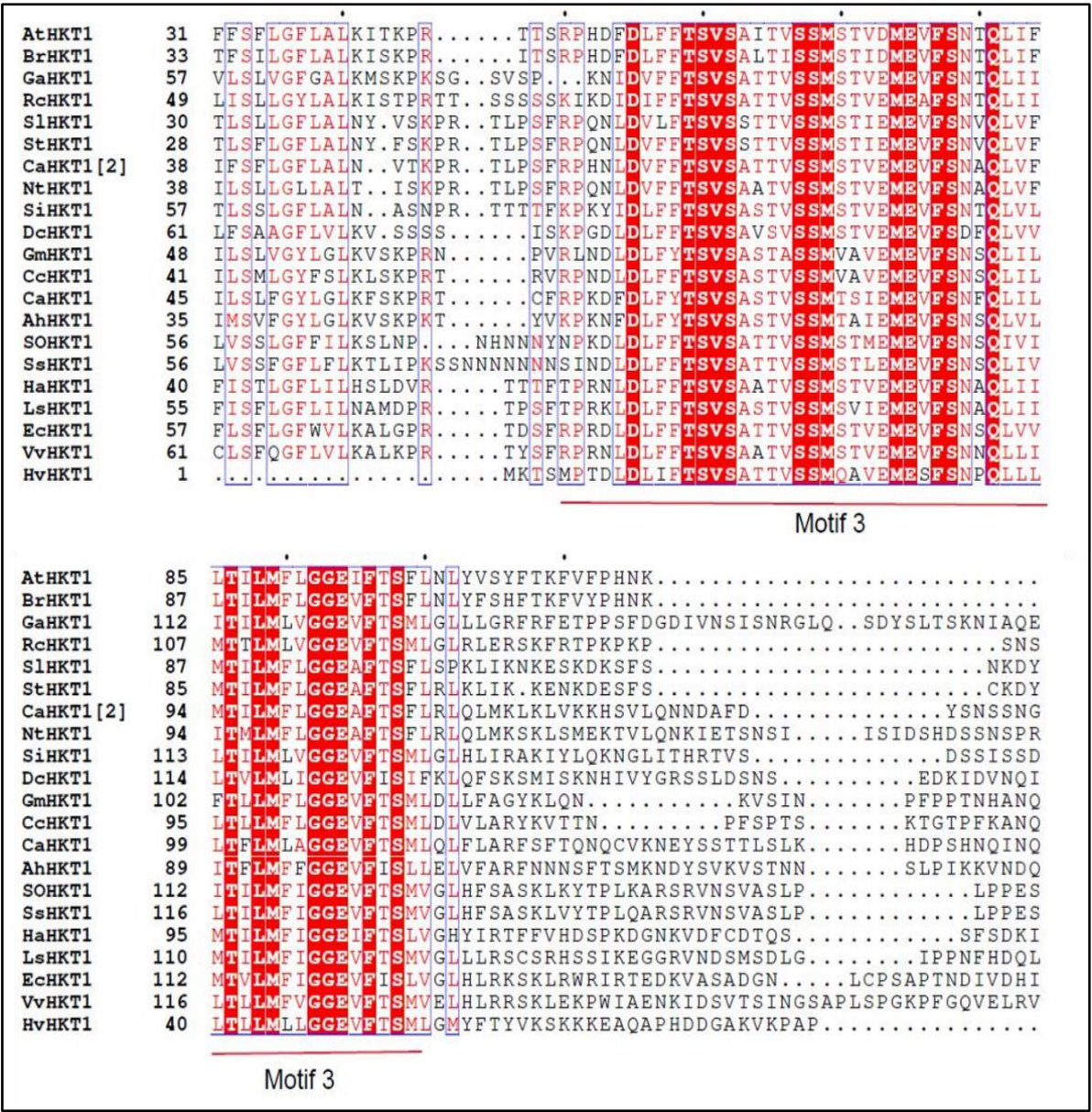


Figure 23: Highly conserved amino acid motif after multiple sequence alignment of HKT1 with motif 3

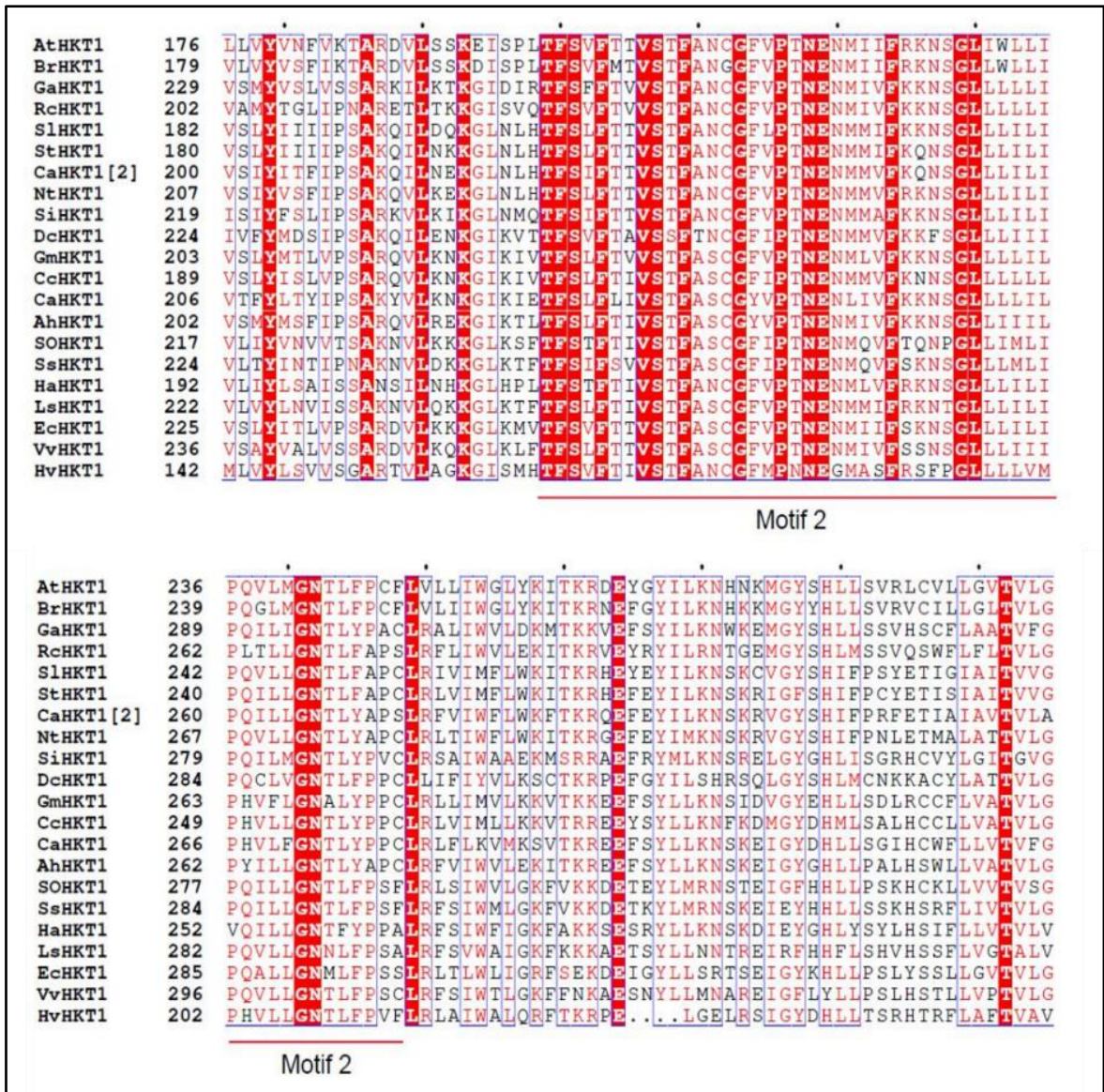


Figure 24: Highly conserved amino acid motif after multiple sequence alignment of HKT1 with motif 2

AtHKT1	296	FLIIQLLFFCAFEWI	SESL	GMSSYEKLVGS	LFQ	VVNSRHT	GETIV	DLSTLS	PAIVL	LVF
BrHKT1	299	FFLFQFLLFCTFEWS	SEPLE	GMSWYEKLVGS	LFQ	VVNSRHT	GETIV	DLSTLS	PATLIF	F
GaHKT1	349	LTLVQFILFCSMEWN	SETMD	GFSSYQKVVGS	LFQ	VVNSRYA	GESVVD	DLSTISS	AILV	LVF
RcHKT1	322	FIVLQLIVFCCMEWN	NSQGM	GLSWVEKLVAV	LFQ	VVSTRHS	GESVFD	LSAIS	PAIV	VIF
SlHKT1	302	LIVFQFVMFCSLEWN	SEGTS	GLSTYEKIVGS	LFQ	VVNRHAG	LSVFD	DLSTFT	PSIL	LVF
StHKT1	300	LIVFQFVVFCSLEWN	SKGTS	GLSTYEKIVGS	LFQ	VVNRHTG	QSVFD	DLSTFT	PAIV	LVF
CaHKT1 [2]	320	FISVQFVAFCSLEWN	SEAA	GLSTYQKLVGS	LFQ	VVNRHTG	QSVFD	DLSTFT	PAIV	LVF
NtHKT1	327	FISIQFVVFCLEWN	SEAT	GLSSYEKLVGS	LFQ	VVNRHTG	ESVFD	DLSTLT	PAIV	LVF
SiHKT1	339	FVIVQLVAFCLLEWK	SEAT	GLSAYQKLVGS	LFQ	VVNSRHT	GESVFD	LSAIS	PAIV	LVF
DcHKT1	344	FTLIQFVIVFVSMWNS	GAME	GLSSYEKLVAT	LFQ	VVNRHTG	ELVFD	DLSTIS	PAV	LVVF
GmHKT1	323	FNLIQFVMLCSLEWK	TKIME	GLNVYEKVVAS	LFQ	VVNARHAG	ESVFD	DLSSIS	AILV	LVF
CcHKT1	309	LNVVQFVMLCSMEWN	SNIME	GLNVYQKLVAS	LFQ	VVNARHS	GESVFD	DLSSIS	AILV	LVF
CaHKT1	326	FIVIQLVLFCSMEWN	SNIME	GLNPYQKLVAS	LFQ	VVNARHS	GESVFD	LSAIS	AILV	VIF
AhHKT1	322	FNIIQFVIFCSMEWG	TQITD	GLNPYQKLVAS	LFQ	VVNARHS	GESVFD	DLSTIS	AILV	LVF
SoHKT1	337	FIMVQFILFCAMEWGF	EGLN	GLNVYQKIIGIL	LFQ	CINSRHT	GESILD	LSIAS	AILV	VVF
SsHKT1	344	FILVQFIMFCSMEWN	FDGLN	DHNIYQKLVGIL	LFQ	CVNSRHT	GESIVD	LSSINS	AMLV	VIF
HaHKT1	312	FIMVQYILFSSMEWSES	SLN	GLNVYQKLVGIL	LFQ	TVNSRYT	GESIVD	DLSTISS	AILV	LVF
LsHKT1	342	FIFVQYILFSSLEWN	ADPMS	GLNHYEKFIGV	LFQ	TINRHTG	ETIVD	DLSTIAA	ASLV	LVI
EcHKT1	345	FVGIQFIMFCSMOWD	SESLN	GLSSCEKIVGAL	LFQ	CVNSRHT	GETVVD	LSTVAP	AILV	LVF
VvHKT1	356	FILIQFTLLCSMDWN	SEGLN	GLNSYQKIIIGAL	LFQ	SVNSRHT	GETIVD	LSILS	PAIV	LVF
HvHKT1	258	FVLAQLSLFCAMEWGS	SDGLR	GLTAAQKLVAA	LFMS	VNSRHAG	EMVVD	DLSTVS	SAVV	VVVY

Motif 5

AtHKT1	355	ILMM...YLPPYT	LFMPL	TEQKTIEKEGGDD	SENGK	KVKKSG	LIVS	QLSFLT	ICIFL	IS			
BrHKT1	358	ITMM...ELPPYT	LFMPL	TEENNNKDE..	EDDSGNGN	KRKRSG	FFVS	QLSFLV	ICIFL	VS			
GaHKT1	408	VVMM...YLPPYTS	FIPT	TKYQKKEIE..	KGSQNESAG	KSILDC	VLF	QLSYLA	IFILIC				
RcHKT1	382	IVMM...YLPPYTS	SFLPV	KQEEELVPRNG	RERKNEK	TLFQD	LLFS	QLSYLV	IFIVL	IC			
SlHKT1	361	ALMM...YLSSYT	TFLPV	.DNYEE...KSEK	MKKRKR	GRSLMEY	ISLS	QPCCLV	IFITIL	IC			
StHKT1	359	ASMM...YLSSYT	TFLPV	.YNYEE...KSEK	MKKRKR	GRSLMDY	ISLS	QPSCLV	IFITIL	IC			
CaHKT1 [2]	379	ALMM...YLSSYT	TFLPV	.DNHEERLKT	KEKINRKR	RRSLVKY	ISFS	QPSCLV	IFITIL	IC			
NtHKT1	386	AVMM...YLSPYTS	SFLPI	.DSNEERLET	TERMNRK	GSSLVEY	VSF	SHLSYL	VFITIL	IC			
SiHKT1	398	VAMM...YLPPYTS	SFLPI	.DEAEMKGE	GEKWRKKK	G..LGEH	IMFS	QLTYLL	IFVIL	IC			
DcHKT1	404	ILMM...YLSPYTS	SFLLD	.GENKGP	DGEDKIT	CTTRKT	TWVEY	LRSE	PTYLT	IFIVL	IC		
GmHKT1	382	VVMM...YLPPYT	TFLPV	REHEKD.....	VNKRNE	KSVV	DCLV	LSQLSY	LVIFIL	IC			
CcHKT1	368	IVMM...YLPPYT	TFLPV	NREKEN.....	DVKRKE	KSIVE	CVMS	QLSYLV	IFILIC				
CaHKT1	385	IVMM...YLPPYT	TFLPL	RDHEIN.....	DAKKDQ	KCLV	DCLV	FSQLSY	LLIFIL	IC			
AhHKT1	381	VVMM...YLPPYT	TTFMP	IMDHKTEN.....	DAKRDK	RS	SLVE	CLLFS	QLSYLV	IFILIC			
SoHKT1	396	VVMM...YLPPYTS	SFLPI	IEEQGE	FAHIMLW	KGVKKE	RKIV	KNFIF	SQSYLA	IFITIL	IC		
SsHKT1	404	IVMM...YLPPYTS	SFLPI	KDEEKEY	PNMVL	FNREK	RRKIL	KNFL	SQSGY	IAIFII	IC		
HaHKT1	371	VLMM...YLPPYTS	SFLPV	AE.ESSEQ	RKRSRK.....	IMDN	LIFS	QPIYL	VIFVIL	V			
LsHKT1	401	VVMM...YLPPYTS	SFLPL	SG.KQSS	QRNSD	QRKKSR	RVLF	ENIV	FSQLAY	LVIFVIL	V		
EcHKT1	404	VVMM...YLPPYTS	SFLPV	KGNRFP	ENGERR	KPKQSY	RLL	LENL	KFSQ	LSYLA	IFIVIC		
VvHKT1	415	VVMM...YLPPYTS	SFLPI	IEDYE	QSSPT	CSGRR	.RRKR	GKIV	ENLIF	SQSYLV	IFILIC		
HvHKT1	317	MVMMCVTYL	PPYT	TFLPV	EDSNQ	QVGTD.....	QKR	TSI	WHKL	LMSP	LSCIA	IFIVV	C

Motif 5

Motif 1

Figure 25: Highly conserved amino acid motif after multiple sequence alignment of HKT1 with motif 5 and motif 1

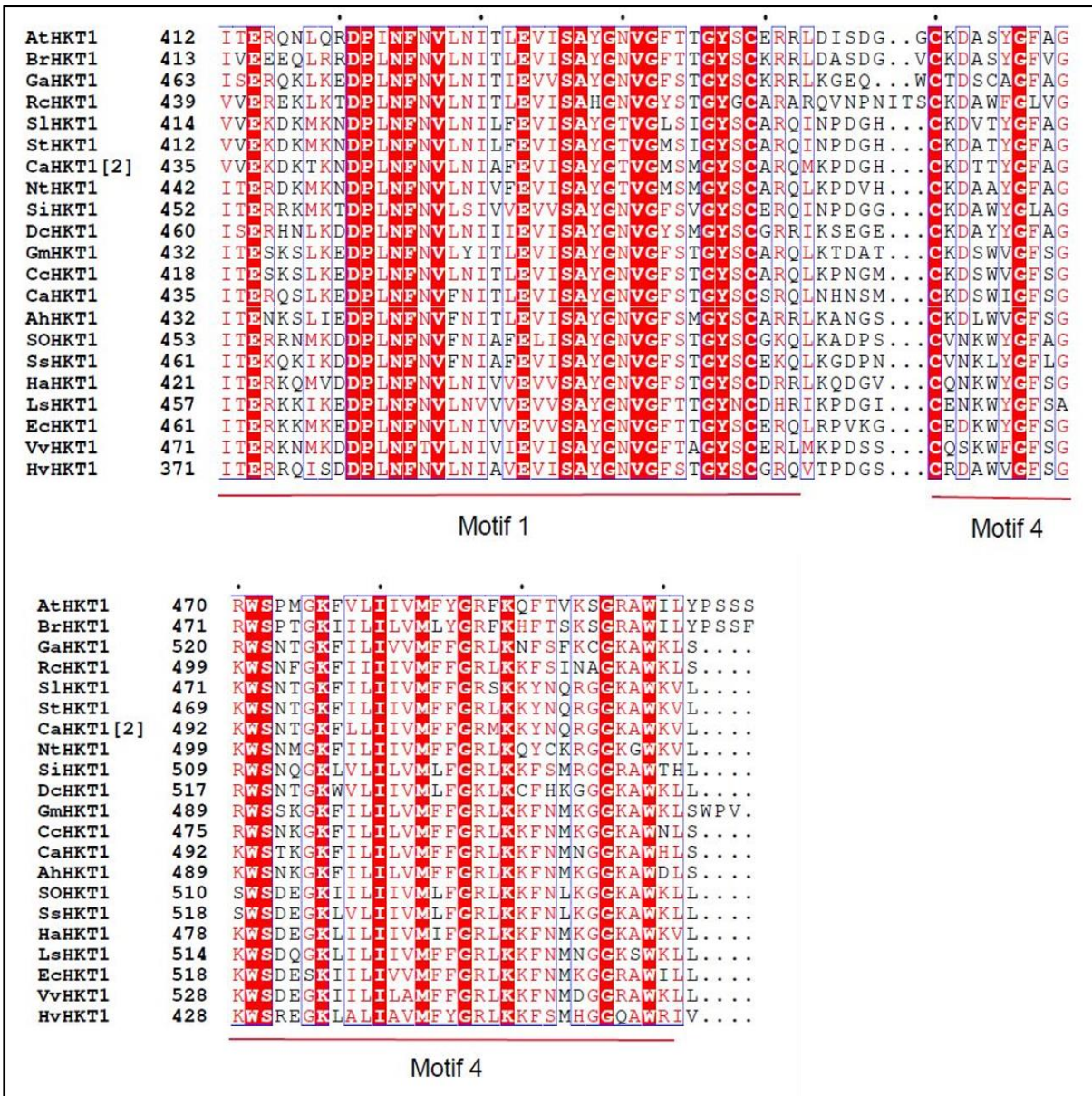


Figure 26: Highly conserved amino acid motif after multiple sequence alignment of HKT1 with motif 1 and motif 4

Table 10: Instability index and Helix Percentage of SOS1

*This protein combination are unstable

SL.No.	Motif's Sequence's	Instability Index	Helix Percentage (%)
1.	A1, A2, A3, A5, A4	35.35	80
2.	A1, A4, A5, A3, A2	37.09	77
3.	A2, A3, A4, A5, A1	37.58	77
4.	A4, A1, A2, A3, A5	37.79	75
5.	A4, A2, A3, A1, A5	37.79	75
6.	A5, A2, A3, A1, A4	37.79	77
7.	A2, A3, A5, A1, A4	37.79	77
8.	A1, A4, A2, A3, A5	37.79	77
9.	A4, A5, A1, A2, A3	37.79	78
10.	A5, A1, A4, A2, A3	37.79	78
11.	A4, A1, A5, A2, A3	37.79	80
12.	A4, A5, A2, A3, A1	37.79	81
13.	A4, A2, A3, A5, A1	37.79	81
14.	A2, A3, A1, A4, A5	37.79	81
15.	A1, A4, A5, A2, A3	37.79	81
16.	A3, A4, A5, A1, A2	37.97	73
17.	A5, A3, A2, A1, A4	38.17	69
18.	A4, A3, A5, A2, A1	38.17	71
19.	A4, A2, A5, A1, A3	38.17	71
20.	A3, A2, A4, A1, A5	38.17	72
21.	A3, A1, A4, A2, A5	38.17	72
22.	A4, A3, A2, A5, A1	38.17	73
23.	A4, A3, A1, A5, A2	38.17	73
24.	A2, A5, A1, A4, A3	38.17	73
25.	A2, A4, A3, A5, A1	38.17	73
26.	A5, A2, A4, A3, A1	38.17	74
27.	A4, A5, A2, A1, A3	38.17	74
28.	A3, A5, A1, A4, A2	38.17	74

29.	A3, A2, A4, A5, A1	38.17	74
30.	A5, A1, A4, A3, A2	38.17	75
31.	A4, A5, A3, A1, A2	38.17	75
32.	A4, A1, A3, A5, A2	38.17	75
33.	A4, A1, A5, A3, A2	38.17	75
34.	A3, A1, A4, A5, A2	38.17	75
35.	A5, A3, A1, A4, A2	38.17	76
36.	A5, A3, A1, A2, A4	38.17	76
37.	A5, A1, A3, A2, A4	38.17	76
38.	A4, A2, A1, A3, A5	38.17	76
39.	A4, A1, A2, A5, A3	38.17	76
40.	A2, A4, A5, A1, A3	38.17	76
41.	A4, A5, A1, A3, A2	38.17	77
42.	A4, A5, A3, A2, A1	38.17	77
43.	A4, A3, A1, A2, A5	38.17	77
44.	A4, A2, A1, A5, A3	38.17	77
45.	A3, A5, A2, A1, A4	38.17	77
46.	A3, A2, A5, A1, A4	38.17	77
47.	A2, A5, A3, A1, A4	38.17	77
48.	A3, A2, A1, A4, A5	38.17	78
49.	A5, A3, A2, A4, A1	38.17	78
50.	A5, A2, A1, A4, A3	38.17	78
51.	A5, A1, A2, A4, A3	38.17	78
52.	A4, A2, A5, A3, A1	38.17	78
53.	A3, A5, A1, A2, A4	38.17	78
54.	A3, A1, A5, A2, A4	38.17	78
55.	A2, A4, A5, A3, A1	38.17	78
56.	A1, A5, A3, A2, A4	38.17	78
57.	A2, A4, A3, A1, A5	38.17	79
58.	A1, A4, A3, A2, A5	38.17	79
59.	A2, A1, A4, A5, A3	38.17	80
60.	A4, A3, A5, A1, A2	38.17	80

61.	A4, A1, A3, A2, A5	38.17	80
62.	A3, A5, A2, A4, A1	38.17	80
63.	A2, A4, A5, A1, A3	38.17	80
64.	A2, A4, A1, A3, A5	38.17	80
65.	A2, A1, A4, A3, A5	38.17	80
66.	A1, A4, A3, A5, A2	38.17	80
67.	A3, A1, A2, A4, A5	38.17	81
68.	A4, A3, A2, A1, A5	38.17	81
69.	A5, A2, A4, A1, A3	38.17	81
70.	A1, A2, A4, A3, A5	38.17	81
71.	A1, A5, A2, A4, A3	38.17	82
72.	A1, A4, A2, A5, A3	38.17	82
73.	A1, A3, A5, A2, A4	38.17	82
74.	A1, A3, A2, A4, A5	38.17	83
75.	A1, A2, A4, A5, A3	38.17	84
76.	A2, A3, A4, A1, A5	38.67	79
77.	A1, A2, A3, A4, A5	38.67	80
78.	A5, A2, A3, A4, A1	38.67	80
79.	A1, A5, A2, A3, A4	38.67	80
80.	A5, A1, A2, A3, A4	38.76	84
81.	A3, A4, A5, A2, A1	39.05	66
82.	A2, A5, A3, A4, A1	39.05	68
83.	A3, A4, A2, A1, A5	39.05	71
84.	A3, A4, A1, A5, A2	39.05	72
85.	A5, A1, A3, A4, A2	39.05	73
86.	A3, A4, A2, A5, A1	39.05	73
87.	A1, A5, A3, A4, A2	39.05	73
88.	A1, A3, A4, A2, A5	39.05	73
89.	A5, A3, A4, A2, A1	39.05	75
90.	A5, A2, A1, A3, A4	39.05	76
91.	A3, A4, A1, A2, A5	39.05	76
92.	A2, A5, A1, A3, A4	39.05	76

93.	A1, A2, A5, A3, A4	39.05	77
94.	A2, A1, A5, A3, A4	39.05	77
95.	A1, A3, A4, A5, A2	39.05	78
96.	A2, A1, A3, A4, A5	39.05	81
97.	A5, A3, A4, A1, A2	39.05	83
98.	A2, A3, A1, A5, A4	39.79	76
99.	A5, A4, A1, A2, A3	39.79	77
100.	A5, A4, A2, A3, A1	39.79	78
101.	A2, A3, A5, A4, A1	39.79	78
102.	A1, 45, A4, A2, A3	39.79	82
103.	A2, A1, A3, A5, A4	40.18*	73
104.	A3, A1, A5, A4, A2	40.18*	73
105.	A1, A5, A4, A3, A2	40.18*	75
106.	A3, A5, A4, A1, A2	40.18*	76
107.	A1, A3, A5, A4, A2	40.18*	76
108.	A1, A2, A5, A4, A3	40.18*	76
109.	A5, A4, A2, A1, A3	40.18*	77
110.	A2, A5, A4, A3, A1	40.18*	77
111.	A5, A4, A1, A3, A2	40.18*	78
112.	A3, A2, A5, A4, A1	40.18*	78
113.	A3, A1, A2, A5, A4	40.18*	78
114.	A5, A4, A3, A2, A1	40.18*	79
115.	A2, A1 A5, A4, A3	40.18*	79
116.	A1, A3, A2, A5, A4	40.18*	79
117.	A5, A4, A3, A1, A2	40.18*	80
118.	A3, A2, A1, A5, A4	40.18*	81
119.	A3, A5, A4, A2, A1	40.18*	81
120.	A2, A5, A4, A1, A3	40.18*	83

#Here **A1**= FPYDWNWKTSLLLGGLLSATDPVAVVALLKELGASKKLSTIIEGESLMND;

A2= VMTLGMFYAAFARTAFKGESQQSLHHFWEMVAYIANTLIFILSGVVIAEG;

A3= ANIDPDLLLAVFLPALLFESSFSMEVHQIKRCMAQMV LLAGPGV;

A4= PRKLVTYFTVERLESACYIAAFLRAHRIARRQLHDFIGDS;

A5=VQAAYWEMPLDEG RITQTTANILMQSVDEALDLVS.

Table 11: Instability index and Helix Percentage of HKT1

*This protein combination are unstable

SL.No.	Motif's Sequence's	Instability Index	Helix Percentage (%)
1.	A1, A4, A5, A3, A2	22.69	57
2.	A1, A4, A5, A2, A3	23.01	61
3.	A4, A1, A5, A3, A2	23.01	57
4.	A3, A1, A4, A5, A2	23.01	60
5.	A5, A3, A2, A4, A1	23.01	62
6.	A3, A5, A1, A4, A2	23.05	59
7.	A5, A3, A1, A2, A4	23.05	58
8.	A2, A3, A1, A4, A5	23.14	59
9.	A4, A5, A3, A1, A2	23.14	60
10.	A4, A1, A2, A5, A3	23.14	55
11.	A2, A4, A5, A3, A1	23.14	54
12.	A1, A2, A4, A5, A3	23.14	48
13.	A2, A5, A3, A4, A1	23.14	44
14.	A5, A3, A4, A1, A2	23.14	56
15.	A5, A3, A1, A4, A2	23.18	55
16.	A2, A5, A3, A1, A4	23.18	52
17.	A1, A4, A2, A5, A3	23.18	42
18.	A4, A1, A5, A2, A3	23.34	56
19.	A4, A5, A2, A3, A1	23.34	56
20.	A3, A2, A4, A1, A5	23.34	63
21.	A3, A2, A4, A5, A1	23.34	56
22.	A4, A1, A3, A5, A2	23.34	60
23.	A4, A5, A1, A3, A2	23.34	57
24.	A4, A1, A3, A2, A5	23.34	60
25.	A3, A5, A2, A4, A1	23.34	61
26.	A5, A2, A4, A1, A3	23.34	50
27.	A1, A3, A2, A4, A5	23.34	60
28.	A5, A2, A3, A4, A1	23.34	58

29.	A3, A4, A1, A5, A2	23.34	66
30.	A1, A3, A4, A5, A2	23.34	64
31.	A5, A4, A1, A3, A2	23.34	61
32.	A3, A2, A5, A4, A1	23.34	60
33.	A5, A2, A3, A1, A4	23.38	56
34.	A3, A2, A5, A1, A4	23.38	49
35.	A1, A5, A3, A2, A4	23.38	52
36.	A2, A3, A4, A5, A1	23.46	45
37.	A4, A1, A2, A3, A5	23.46	58
38.	A4, A5, A1, A2, A3	23.46	61
39.	A3, A4, A5, A1, A2	23.46	62
40.	A2, A4, A5, A1, A3	23.46	56
41.	A2, A4, A5, A1, A3	23.46	56
42.	A2, A4, A1, A3, A5	23.46	55
43.	A3, A1, A2, A4, A5	23.46	63
44.	A2, A3, A4, A1, A5	23.46	50
45.	A1, A2, A3, A4, A5	23.46	58
46.	A3, A4, A1, A2, A5	23.46	60
47.	A5, A4, A1, A2, A3	23.46	61
48.	A2, A3, A5, A4, A1	23.46	58
49.	A3, A5, A4, A1, A2	23.46	59
50.	A2, A5, A4, A1, A3	23.46	55
51.	A2, A3, A5, A1, A4	23.50	50
52.	A1, A4, A2, A3, A5	23.50	53
53.	A5, A1, A4, A2, A3	23.50	56
54.	A3, A1, A4, A2, A5	23.50	58
55.	A4, A2, A5, A3, A1	23.50	49
56.	A1, A5, A3, A4, A2	23.50	55
57.	A1, A2, A5, A3, A4	23.50	39
58.	A5, A2, A1, A3, A4	23.52	52
59.	A2, A1, A4, A5, A3	23.64	49
60.	A5, A1, A3, A2, A4	23.70	64

61.	A3, A1, A5, A2, A4	23.70	58
62.	A1, A3, A5, A2, A4	23.70	62
63.	A1, A5, A2, A3, A4	23.70	54
64.	A1, A3, A2, A5, A4	23.70	59
65.	A1, A2, A3, A5, A4	23.82	51
66.	A4, A2, A3, A1, A5	23.82	52
67.	A4, A2, A3, A5, A1	23.82	52
68.	A4, A2, A5, A1, A3	23.82	57
69.	A3, A5, A1, A2, A4	23.82	62
70.	A5, A1, A2, A3, A4	23.82	58
71.	A5, A1, A3, A4, A2	23.82	66
72.	A3, A4, A2, A5, A1	23.82	58
73.	A1, A3, A4, A2, A5	23.82	58
74.	A2, A5, A1, A3, A4	23.82	49
75.	A2, A3, A1, A5, A4	23.82	56
76.	A5, A4, A2, A3, A1	23.82	60
77.	A1, A5, A4, A2, A3	23.82	49
78.	A3, A1, A5, A4, A2	23.82	58
79.	A1, A3, A5, A4, A2	23.82	60
80.	A3, A1, A2, A5, A4	23.82	57
81.	A4, A5, A3, A2, A1	23.84	56
82.	A3, A2, A1, A4, A5	23.84	52
83.	A5, A3, A2, A1, A4	23.88	45
84.	A4, A5, A2, A1, A3	24.16	52
85.	A3, A4, A5, A2, A1	24.16	57
86.	A3, A5, A2, A1, A4	24.20	55
87.	A2, A1, A3, A4, A5	24.28	44
88.	A4, A2, A1, A5, A3	24.32	58
89.	A5, A3, A4, A2, A1	24.32	60
90.	A2, A1, A5, A3, A4	24.32	52
91.	A3, A2, A1, A5, A4	24.52	59
92.	A4, A2, A1, A3, A5	24.65	49

93.	A3, A4, A2, A1, A5	24.65	64
94.	A2, A1, A3, A5, A4	24.65	55
95.	A5, A4, A2, A1, A3	24.65	59
96.	A3, A5, A4, A2, A1	24.65	57
97.	A5, A1, A4, A3, A2	24.77	59
98.	A1, A4, A3, A2, A5	24.77	49
99.	A1, A4, A3, A5, A2	24.77	59
100.	A2, A5, A1, A4, A3	24.89	46
101.	A4, A3, A2, A5, A1	25.09	54
102.	A4, A3, A1, A5, A2	25.09	59
103.	A5, A2, A4, A3, A1	25.09	58
104.	A1, A5, A2, A4, A3	25.09	44
105.	A1, A5, A4, A3, A2	25.09	53
106.	A2, A4, A3, A5, A1	25.22	50
107.	A4, A3, A1, A2, A5	25.22	60
108.	A5, A1, A2, A4, A3	25.22	62
109.	A2, A4, A3, A1, A5	25.22	50
110.	A4, A3, A5, A1, A2	25.22	63
111.	A1, A2, A4, A3, A5	25.22	54
112.	A1, A2, A5, A4, A3	25.22	51
113.	A2, A5, A4, A3, A1	25.22	55
114.	A5, A4, A3, A1, A2	25.22	57
115.	A5, A2, A1, A4, A3	25.59	54
116.	A2, A1, A4, A3, A5	25.72	51
117.	A4, A3, A5, A2, A1	25.92	62
118.	A4, A3, A2, A1, A5	25.92	60
119.	A5, A4, A3, A2, A1	25.92	64
120.	A2, A1 A5, A4, A3	26.04	52

#Here **A1**= LVIFILICITERKKLKEDPLNFNVLNITLEVISAYGNVGFSTGYSCARQ;

A2= TFSLFTTVSTFANCGFYPTNENMIYFKKNSGLLLILIPQVLGNTLFPPC;

A3= RPKDLDLFFTSVSATTVSSMSTVEMEVFSNTQLIFLITILMFLGGEVFTSM;

A4= CKDAWYGFSGKWSNTGKFILIIVMFFFGRLLKKNMKGKAWK;

A5= GLSS YQKLVGSLFQVVNSRHTGESVFDLSTISPAILVLFVMM

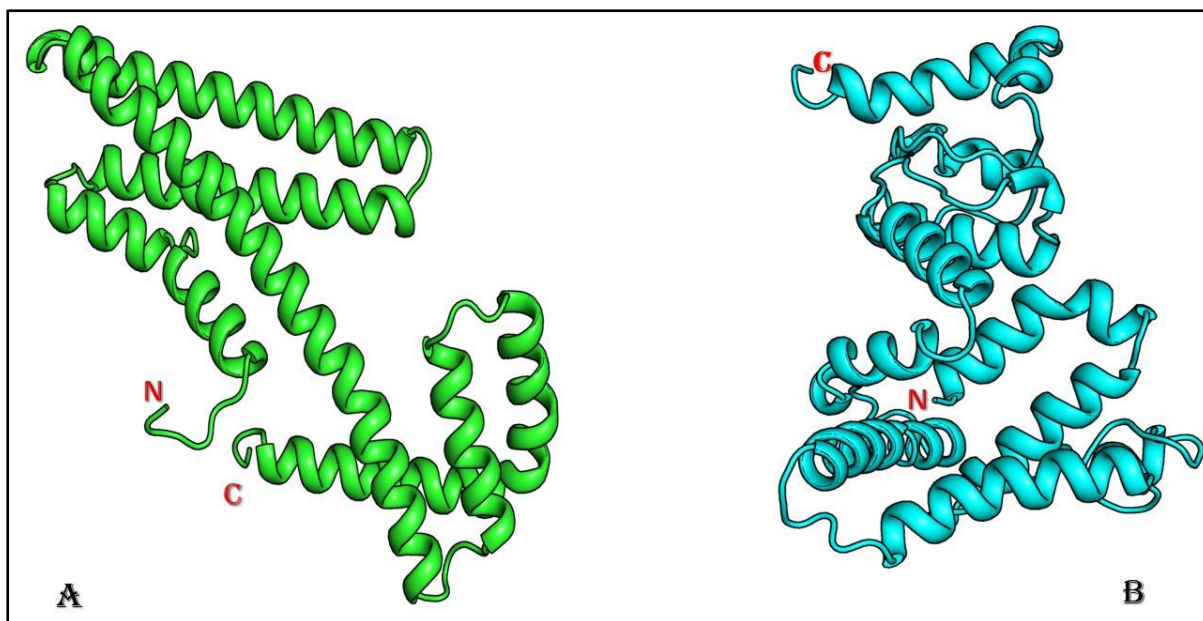


Figure 27: Peptide design of secondary structure before molecular dynamic simulation.
A. SOS1; B. HKT1

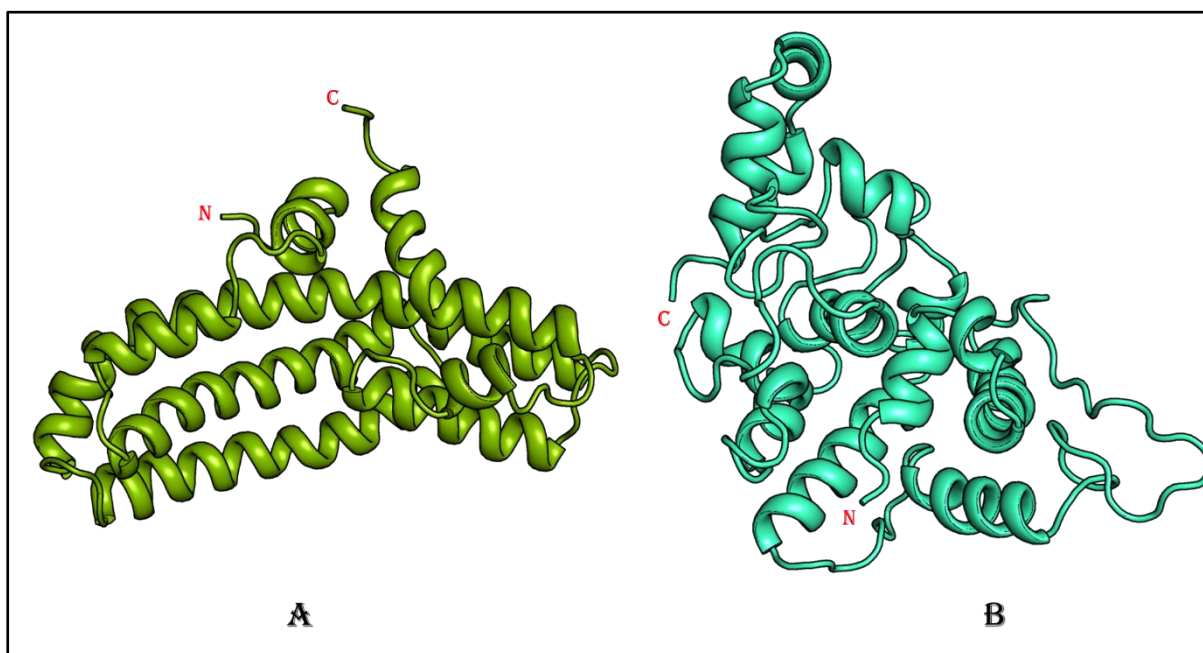


Figure 28: Peptide design of secondary structure after molecular dynamic simulation.
A. SOS1; B. HKT1

Table 12: Antigenicity of SOS1 by SVMTrip

*Recommend Score

Sl.No.	Location	Epitope	Score
1.	117-136	VFDLSTISPAILVLFVMMR	1.0*
2.	46-65	SCARQCKDAWYGFSGKWSNT	0.84
3.	211-230	FKKNSGLLLILIPQVLGNTL	0.708
4.	74-93	MFFFGRLKKFNMKGGKAWKG	0.571

Table 13: Antigenicity of HKT1 by SVMTrip

*Recommend Score

Sl.No.	Location	Epitope	Score
1.	4-23	DWNWKTSLLLGGLLSATDPV	1.0*
2.	52-71	MTLGMFYAAFARTAFKGESQ	0.773
3.	84-103	IANTLIFILSGVVIAEGANI	0.623
4.	194-213	ACYICAAFLRAHRIARRQLH	0.622
5.	143-162	GVVQAAYWEMLDEGRITQTT	0.474

Table 14: Antigenicity of SOS1 by ABCPred

*Recommend Score

Sl.No.	Rank	Sequence	Start position	Score
1.	1	AAYWEMLDEGRITQTT	147	0.84*
2.	2	STIIEGESLMNDVMTL	39	0.83
3.	3	LLSATDPVAVVALLKE	16	0.82

4.	4	AHRIARRQLHDFIGDS	204	0.8
5.	4	VDEALDLVSPRKLVTY	170	0.8
6.	5	KGESQQSLHHFWEMVA	67	0.79
7.	6	VIAEGANIDPDLLAV	96	0.78
8.	7	FYA AFARTAFKGESQQ	57	0.77
9.	7	HQIKRCMAQMVLLAGP	127	0.77
10.	8	EGRITQTTANILMQSV	155	0.75
11.	9	DWNWKTSLLLGGLLSA	4	0.69
12.	9	ACYICAAFLRAHRIAR	194	0.69
13.	9	MVLLAGPGVVQAAYWE	136	0.69
14.	10	VSPRKLVTYFTVERLE	177	0.65

Table 15: Antigenicity of HKT1 by ABCPred

*Recommend Score

Sl.No.	Rank	Sequence	Start position	Score
1.	1	TVSSMSTVEMEVFSNT	151	0.91*
2.	2	IFIILICITERKKLKE	3	0.88
3.	3	AWYGFSGKWSNTGKFI	54	0.87
4.	4	TGYSCARQCKDAWYGF	43	0.86
5.	5	GNVGFSTGYSCARQCK	37	0.85
6.	6	CITERKKLKEDPLNFN	9	0.84
7.	6	GGEVFTSMTFSLFTTV	178	0.84
8.	7	FVVMMPKDLDFFTS	131	0.79
9.	8	YPTNENMIYFKKNSGL	202	0.78
10.	8	FFTSVSATTVSSMSTV	143	0.78
11.	9	GKAWKGLSSYQKLVGS	88	0.76
12.	10	TTVSTFANCGFYPTNE	191	0.75
13.	11	VVNSRHTGESVFDLST	107	0.73

14.	12	LLLILIPQVLGNTLFP	217	0.7
15.	13	KKFNMKGGKAWKGLSS	81	0.67
16.	13	LEVISAYGNVGFSTGY	30	0.67
17.	14	GKFILIIVMFFFGRLK	66	0.66

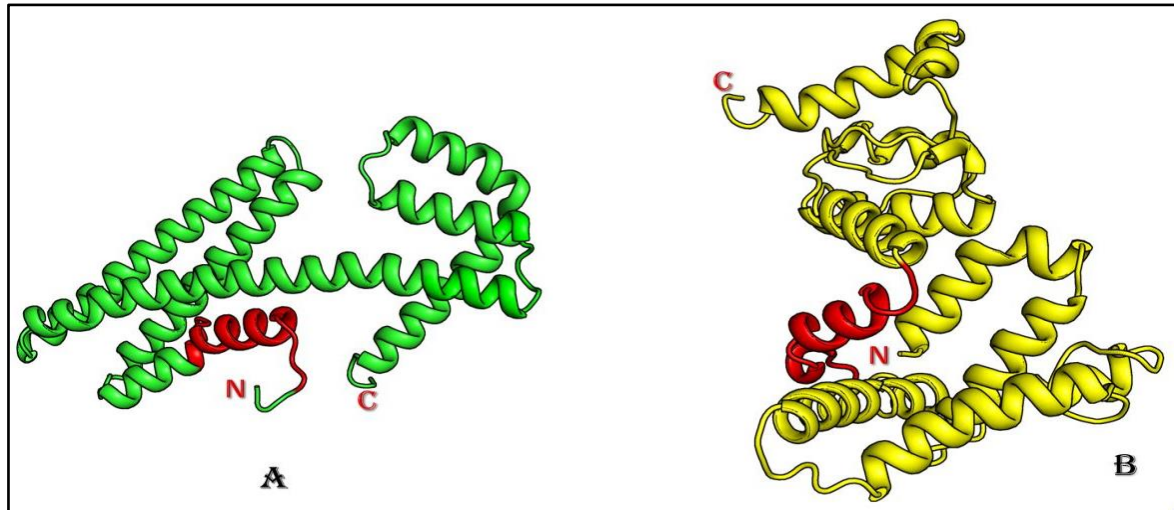


Figure 29: Antigenicity mapping of highest score **A.** SOS1 and **B.** HKT1 by SVMTrip server

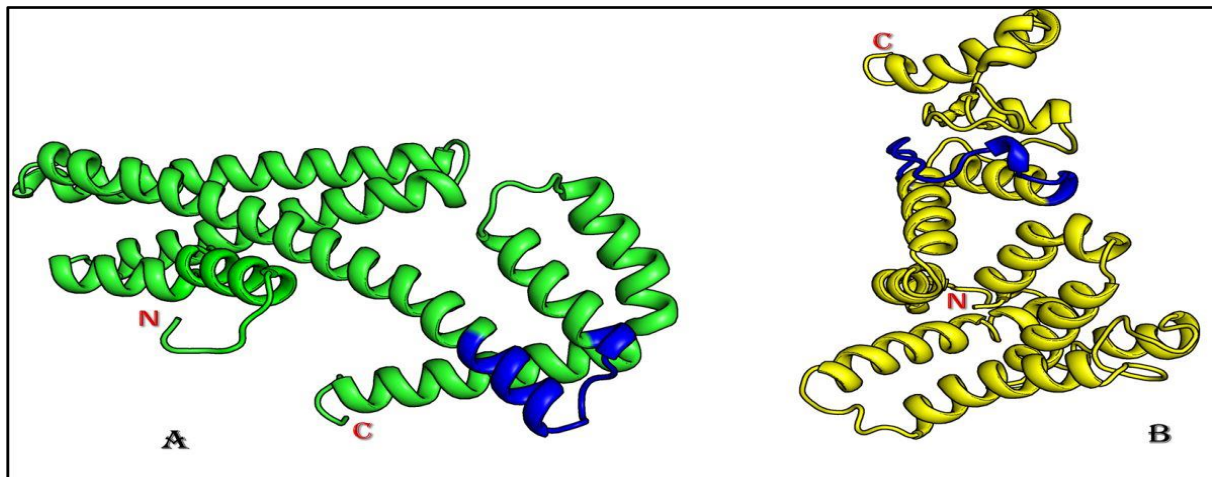


Figure 30: Antigenicity mapping of highest score **A.** SOS1 and **B.** HKT1 by ABCPred server

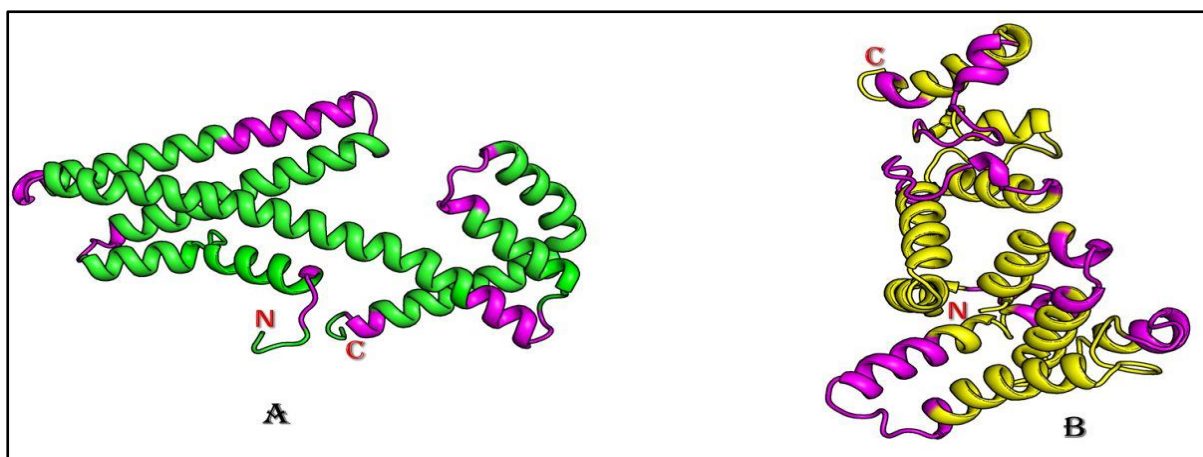


Figure 31: Antigenicity mapping of highest score **A. SOS1** and **B. HKT1** by BepiPred server

Table 16: Shows Physio-chemical properties of SOS1 by Bcepred server

Sl.No.	Sequence	Different properties	Selected Sequence
1.	FPYDWNWKTSLLLGGLLSATDPV AVVALLKELGASKKLSTIEGESL MNDVMTLGMFYAAFARTAF <u>KGE</u> <u>SQQS</u> LHHFWEMVAYIANTLIFILS GVVIAEGANIDPDLLLAVFLPALL FESSFSMEVHQIKRCMAQMVLLA GPGVVQAAYWEMLDEGRITQTT ANILMQSVDEALDLVSPRKLVTY FTVERLESACYICAAFLRAHRIAR RQLHDFIGDS	Hydrophilicity	KGESQQS

2.	FPYDWNWKTSLLLGGLLSATDPV AVVALLKEL <u>LGASKKL</u> STIIEGESL MNDVMTLGMFYAAFAR <u>TAFKGE</u> <u>SQ</u> SLHHFWEMVAYIANTLIFILS GVVIAEGANIDPDLAVFLPALL FESSFSMEVHQIKRCMAQMVLLA GPGVVQAAYWEMLDEGRITQTT ANILMQSVDEALDLVSPRKLVTY FTVERLESACYICAAFLRAHRIAR RQLHDFIGDS	Flexibility	LGASKKL/ TAFKGESQ
3.	F <u>PYDWNWKTS</u> LLLGGLLSATDPV AVVALL <u>KELGASKKLST</u> IIEGESL MNDVMTLGMFYAAFAR <u>RTAFKGE</u> <u>SQ</u> SLHHFWEMVAYIANTLIFILS GVVIAEGANIDPDLAVFLPALL FESSFSM <u>EVHQIKR</u> CMAQMVLLA GPGVVQAAYWEML <u>DEGRITQTT</u> ANILMQSVDEAL <u>DLVSPRKLVTY</u> FTVERLESACYICAAFL <u>RAHRIAR</u> <u>RQLHDF</u> IGDS	Accessibility	PYDWNWKTS/ KELGASKKLST/ RTAFKGESQSLH/ EVHQIKR/ DEGRITQT/ DLVSPRKLVTY/ RAHRIARRQLHDF
5.	FPYDWNWKTSLLLGGLLSATDPV AVVALLKELGASKKLSTIIEGESL MNDVMTLGMFYAAFAR <u>RTAFKGE</u> SQ <u>SLHHFWEMV</u> AYIANTLIFILS GVVIAEGANIDPDLAVFLPALL FESSFS <u>MEVHQIKRCM</u> AQMVLLA GPGVVQAAYW <u>EMLDEGR</u> ITQTT ANILMQSVDEALDLVSPRKLVTY <u>FTVERLESAC</u> YICAAFL <u>RAHRIAR</u> <u>RQLHDF</u> IGDS	Polarity	RTAFKGE/ SLHHFWEMV/ MEVHQIKRCM/ EMLDEGR/ FTVERLESAC/ AFLRAHRIARRQL HDF

6.	FPYDWNWKT <u>SLLLGGLLS</u> ATDPV AVVALLKELGASKKLSTIIEGESL MNDVMTLGMFYAAFARTAFKGE SQQSLHHFWEMVAYIAN <u>TLIFILS</u> <u>GVVIAEGANIDPDLLL</u> AVFLPALL FESSFSME <u>VHQIKRC</u> MAQMVLLA GPGVVQAAYWEMLDEGRITQTT ANILMQSVDEA <u>LDLVSPRKLVTY</u> <u>FTVE</u> RLESACYICAAFLRAHRIAR RQLHDFIGDS	Antigenic propensity	SLLLGGLLS/ TLIFILSGVVI/ IDPDLLL/ VHQIKRC/ LDLVSPRKLVTYFT VE
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Table 17: Shows Physio-chemical properties of HKT1 by Bcepred server

Sl.No.	Sequence	Different properties	Selected Sequence
1.	LVIFILICITERK KLK EDPLNFNVL NITLEVISAYGNVGFSTGYSC <u>ARQ</u> <u>CKDA</u> WYGFSGKWSNTGKFILIV MFFFGRLKKFNMKGGKAWKGLS SYQKLVGSLFQVV <u>NSRHTGES</u> VF DLSTISPAILVLFVVMMPKDLDL FFTSVSATTVSSMSTVEMEVFSNT QLIFLTILMFLGGEVFTSMTFSLFT TVSTFANCGFYPTNENMIYFKKNS GLLLILIPQVLGNTLFPPC	Hydrophilicity	ARQCKDA/ NSRHTGES
2.	LVIFIL <u>ICITERKKLK</u> EDPLNFNVL NITLEVISAYGNVGFSTGYSCARQ CKDAW <u>YGFSGKW</u> SNTGKFILIV MFFFGRL <u>KKFNMKG</u> GKAWKGLS SYQKLVGSLF <u>QVVNSRHTGES</u> VF DLSTISPAILVLFVVMMPKDLDL FFTSVSATTVSSMSTVEMEVFSNT QLIFLTILMFLGGEVFTSMTFSLFT	Flexibility	ICITERK KLK / YGFSGKW/ KKFNMKG/ QVVNSRHTG/ MIYFKKNS

	TVSTFANCGFYPTNEN <u>MIYFKKNS</u> GLLLILIPQVLGNTLFPPC		
3.	LVIFILI <u>CITERKCLKEDPLN</u> NFNVL NITLEVISAYGNVGFSTGYSC <u>ARQ</u> <u>CKDAW</u> YGFSG <u>KWSNTGK</u> FILIIV MFFF <u>GRLKKFNMKGGKAWK</u> GLS SYQKLVGSLFQVV <u>NSRHTGES</u> VF DLSTISPAILVLFVV <u>MMRPKDLDL</u> FFTSVSATTVSSMSTVEMEVFSNT QLIFLTILMFLGGEVFTSMTFSLFT TVSTFANCG <u>FYPTNENMIYFKKNS</u> <u>GLLLILIPQVLGNTLFPPC</u>	Accessibility	CITERKCLKEDPLNF N/ ARQCKDAW/ KWSNTGK/ GRLKKFNMKGGKA WK/ NSRHTGES/ MMRPKDLDL/ FYPTNENMIYFKKN SG
4.	LVIFILI <u>CITERKCLKEDPL</u> NFNVL NITLEVISAYGNVGFSTGYSCARQ CKDAWYGFSGKWSNTGKFILIIV MFFF <u>GRLKKFNMKG</u> GKAWKGLS SYQKLVGSLFQVVNSRHTGESVF DLSTISPAILVLFVVMMPRPKDLDL FFTSVSATTVSSMSTVEMEVFSNT QLIFLTILMFLGGEVFTSMTFSLFT TVSTFANCGFYPTNENMIYFKKNS GLLLILIPQVLGNTLFPPC	Exposed surface	CITERKCLKEDPL/ LKKFNMKG
5.	LVIFIL <u>ICITERKCLKEDPL</u> NFNVL NITLEVISAYGNVGFSTGYSCARQ CKDAWYGFSGKWSNTGKFILIIV MFFF <u>GRLKKFNMKG</u> GKAWKGLS SYQKLVGSLFQVV <u>NSRHTGES</u> VF DLSTISPAILVLFVVM <u>MRPKDLDL</u> FFTSVSATTVSSMSTVEMEVFSNT QLIFLTILMFLGGEVFTSMTFSLFT TVSTFANCGFYPTNENMIYFKKNS GLLLILIPQVLGNTLFPPC	Polarity	ICITERKCLKEDPL/ GRLKKFNMKG/ NSRHTGESV/ MRPKDLDL

6.	<u>LVIFIILICITE</u> RKKLKEDPLNFNVL N <u>ITLEVISA</u> YGNVGFSTGYSCARQ CKDAWYGFSGKWSNTG <u>KFILIIV</u> <u>MFFF</u> GRLKKFNMKGGKAWKGL <u>S</u> <u>SYQKLVGSLFQVVNS</u> RHTGES <u>VF</u> <u>DLSTI</u> SPAIL <u>VLFVMMR</u> PKDLD <u>L</u> <u>FFTSVS</u> ATTVSSMSTVEMEVFSNT <u>QLIFLTILMFL</u> GGEVFTSMT <u>FSLFT</u> <u>TV</u> STFANCGFYPTNENMIYFKK <u>NS</u> <u>GLLLILIPQLG</u> NTLFPPC	Antigenic propensity	LVIFIILICITE/ ITLEVISA/ KFILIIVMFFF/ LSSYQKLVGSLFQV VNS/ FDLSTI/ ILVLFVMMR/ LFFTSVS/ QLIFLTILMFL/ FSLFTTV/ NSGLLLILIPQLG
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